

The Agriculturists 16(2): 15-24 (2018)ISSN 2304-7321 (Online), ISSN 1729-5211 (Print)A Scientific Journal of Krishi FoundationIndexed JournalDOI: http://dx.doi.org/ 10.3329/agric.v16i02.40339Impact Factor: 0.568 (GIF, 2015)

Antifungal Efficacy of Plant Oils Containing Thymol and Carvacrol in Controlling Botrytis cinerea, the Causal Agent of Grape (Vitis vinifera) Gray Mould

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Received: 26 September 2018

Accepted: 25 December 2018

Abstract

The efficacy of a large number of plant extracts and essential oils in controlling plant diseases has been proven worldwide. *Botrytis cinerea* has attacked a wide host range causing severe loss in the field and at storage. In this study the antifungal efficacy of essential oils of three medicinal plants namely wild marjoram (*Zataria multifolia*), wild savory (perennial) (*Satureja mutica*) and savory (annual) (*Satureja hortensis*) possessing these compounds at three concentrations: 50, 100 and 200 ppm were evaluated in controlling the mycelial growth, spore germination and germ tube elongation of *B. cinerea*. All treatments except savory (annual) essential oil at 50 ppm showed significant differences with the control in inhibiting the mycelial growth as well as spore germination and germ tube elongation of *B. cinerea* (p=0.01). It was also noted that wild marjoram at 100 ppm and wild savory (perennial) essential oils at 200 ppm could completely (100%) inhibit the growth of the fungus. Essential oils from wild marjoram and wild savory had higher antifungal activity than annual savory. Their suitable formulations could be prepared and used as safe alternatives for controlling moulds of horticultural products during storage.

Keywords: Antifungal potential, essential oils, grape, gray mould.

1. Introduction

Grape is one of the main horticultural products of temperate regions in Iran. About 300000 hectare of land is under its cultivation with an average yield of about 25 tons/ha. A variety called Red Fakhri (thick skinnd) is grown in norther Iran with a good storage property. This product is harvested and kept in cold storage (about 1°C) during the winter and gradually sold until the spring (Ahmadi *et al.*, 2016).

Grape gray mould caused by *Botrytis cinerea* Pers. Ex. Fr. is one the most important decaying

fungi of this product in the wine yards and also in the store leading to rotting of the product (Elad *et al.*, 2007). Among several postharvest diseases of table grapes (*Vitis vinifera*) occurring during storage, gray mold is a particularly, severe disease because the causal agent, *B. cinerea*, grows at temperature as low as 0°C. Other postharvest diseases, caused by *Penicillium* spp. and *Aspergillus* spp., also often lead to deterioration of table grapes after harvest (Shin *et al.*, 2014). In Iran, during the storage period *B. cinerea.* causes more damage than those caused by *Penicillium* spp. (Ghafari *et al.*, 2011; Jalili-Marandi *et al.*, 2010). Extracts and essential oils of a huge number of plants possess chemical compounds such as alkaloids, tannins, quinines, phenolic compounds and phytoalexins with their approved antifungal efficacy (Fawcett and Spencer, 1966). The use of plant essential oil compounds such as thymol and linalool, to reducing postharvest diseases in several kinds of fruits, including table grapes and oranges, has received much attention in European countries.

Results of a research showed that among 21 plant essential oils, the thyme (Thymus capitatus) essential oil at 1%, completely controlled the mycelial growth as well as spore germination of B. cinerea under in vitro condition while essential oils of clove (Syzygium aromaticum) and Australian meant an (Prostanthera rotundifolia) had fungistatic activity of 93 and 65% respectively (Antonov et al., 1998). Siripornvisal et al. (2009) evaluated four medicinal plant essential oils against B. cinerea and reported that in a modified in- vitro micro-atmospheric test, vapors of clove, cinnamon and lemongrass oils exhibited strong inhibitory effects on B. cinerea, with minimal inhibitory quantity (MIO) equal to 15 uL while the vapors of galingale oil exhibited weaker inhibitory effect with MIQ of 25 µL. They reported that among four medicinal plant essential oils, clove, cinnamon and lemongrass oils exhibited fungicidal effect on the pathogen, while galingale oil exhibited some fungistatic properties and finally they concluded that the clove, cinnamon and lemongrass essential oils might be used as alternative options for the control of gray mould on postharvest organic strawberry fruits especially as a natural fumigant in closed container or packaging (Siripornvisal et al., 2009).

In a survey, antifungal activity of 13 medicinal plant essential oils were studied against apple gray mold through pour plate method and production of volatile compounds. The results showed that essential oils of anise, cumin, caraway, ammin, pennyroyal, thyme and cinnamon at concentrations of 250, 500, 750

 μ L/L. had a significant effect against *B. cinerea* on PDA culture. By the volatile compounds production method, cumin, ammin, pennyroyal, dill, cinnamon, anise and caraway essential oils showed the most inhibitory effect and the others illustrated somewhat fungistatic power. (Behdani et al., 2012). Shin et al. (2014) reported that thymol (30 μ g/mL) and linalool (120 μ g/mL) significantly inhibited the mycelial growth and conidia germination of B. cinerea. The occurrence rate of gray mould and other unknown fungi were significantly reduced by fumigation with 30 µg/mL thymol in several table grape cultivars, such as Campbell early, Muscat Bailey A, Sheridan, and Geobong. However fumigation at such rate had no influence on the sugar content and hardness of grapes, but significantly reduced the fungal infection. Therefore, they suggested that 30 µg/mL thymol could be utilized to reduce deterioration of grapes caused by gray mold and other fungal infections during long-term storage. Mermer-Dogu and Zobar (2014) studied the effect of 7 different essential oils of grape seed (Vitis vinifera), thyme (Thymus sp.), rosemary (Rosmarinus officinalis), ozone oil (Olea europaea), peppermint (Mentha piperita), basil (Ocimum basilicum) and sage (Salvia spp.) against B. cinerea and reported that in term of priority, the thyme, peppermint and rosemary oils were effective against this pathogen.

Sesan et al. (2015) working on effect of several plant extracts of savory (Satureja hortensis), garlic (Allium sativum), peppermint (Mentha piperita), rosemary (Rosmarinus officinalis), wormwood (Artemisia dracunculus), yarrow (Achillea millefolium), and hyssop (Hyssopus officinalis) against B. cinerea of blackcurrant (Ribes nigrum L.) reported that 10 to 20% concentrations of savory, garlic, peppermint and hyssop extracts could control the pathogen between 80 to 100%, while the lowest antibotrytis activity or no efficiency was noticed by yarrow, wormwood, wormwood and rosemary extracts when even applied at 20%. In another study, savory (Satureja hortensis), wild marjoram (Zataria multiflora) and ajwain

(*Carum copticom*) essential oils were tested *in vitro* (on PDA) and *in vivo* (dipping inoculated fruit into essential oil solutions) against strawberry gray mold (*B. cinerea*) and authors concluded that a 200 ppm concentration of savory, wild marjoram and ajwain were able to effectively control the growth of the fungus in both methods (Etemadi *et al.*, 2012).

In the present study, the efficacy of wild marjoram (*Zataria multifolia*), annual savory (*Satureja hortensis*) and perennial or wild savory (*S. mutica*) essential oils at three concentrations (50, 100 and 200 ppm) in inhibiting the mycelial growth, spore germination and germ tube elongation of grape gray mould causal agent were compared.

2. Materials and Methods

2.1 Essential oil extraction

Matured plant parts of wild marjoram, annual savory and perennial savory were washed and dried in shade at room temperature. They were then powdered by grinder and 100 g. of each were mixed with 500 ml distilled water. Essential oils were extracted by steam distillation method using a clevenger apparatus for 5 hr. Measurements of essential oils were performed using а QP-5050A (Shimadzu) gas chromatograph coupled with a VG Autospect Mass Spectrometer (MS) at 70 eV, 40-550 amu with a fused silica capillary column using helium as a carrier gas and with temperature programming from 60 °C/5 minutes to 300 °C/2 minutes. The MS was operated using an interface temperature of 240 °C, and an electron impact ionization of 70 eV with a scan mass range of 40-350 m/z (sampling rate of 1.0 scan/s). Identification of the compounds was conducted by comparing their linear retention indices (LRI) with literature values and their mass spectral data with those from the MS data system. Linear retention indices were calculated using GC data of a homologous series of saturated aliphatic hydrocarbons (C_8 to C_{40}) separated on the same column using the same conditions as for GC analysis of the essential oils (Adams, 2007).

These essential oils prepared as 100% pure materials were kept in cold storage for further studies.

2.2 Preparation of selective media for isolation of B. cinerea from infected grape berries

For preparation of *Botrytis* selective medium, 1 g NaNO3, 1.2 g K2HPO4, 0.2 g MgSO₄.7H₂O, 0.15 g KCl, 20 g glucose and 25 g agar were added to 1 L of distilled water and autoclaved at 121°C for 20 min. The prepared medium was cooled to 60° C and then under sterilized condition, 0.015 g PCNB, 0.01 g Maneb, 0.05 g Chloramphenicol, 2.2 g CuSO₄ and 5 ml of tannic acid were added to 5.5 with 1N NaOH (Kritzman and Netzer, 1978).

2.3 Isolation and purification of B. cinerea

Infected grape bunches were collected from grape wine yards and were transferred to the laboratory. They were thoroughly washed in running water, and then small berry portions from in between healthy and affected parts were separated and were surface sterilized with sodium hypocheloride 0.5% for 30 sec. After washing 2-3 times with sterilized distilled water they were dried in between sterilized filter papers and cultured in Botrytis selective medium as mentioned by Kritzman and Netzer (1978). After 3-4 days, using hyphal tip method, small portions of newly growing hypha with medium were transferred to petri plates containing potato dextrose agar (PDA) medium and kept at 20 °C in an incubator for its proper growth. Petri plates were then sealed with parafilm and kept at 4°C in a refrigerator for further use.

2.4 Pathogenicity test

Botrytis spores were collected from 2 weeks old culture grown on PDA into an Erlenmayer flask filled with sterilized distilled water using a sterilized scalpel. The Erlenmayer flask was then kept on a shaker for 20 min to gain a homogenized spore suspension. Using a hemocytometer, the spore concentration was adjusted to 10^5 spores/mL. Before inoculating the grape berries with spore suspension, they

were surface sterilized with Ethanol 70% and then slightly injured with a sterilized needle. Inoculated grape berries were then kept inside sterilized desiccator at 20 °C and were inspected daily for appearance of gray mold symptoms. After appearance of symptoms and production of abundant number of spores the causal agent was reisolated and identified using standard identification key (Beever and Weeds, 2004) to fulfill the Koch's postulates.

2.5 Effect of different essential oil concentrations on mycelial growth of B. cinerea

For estimating the antifungal effect of three concentrations (50, 100 and 200 ppm) of tested essential oils on mycelial growth of the fungus, poisoned food technique was employed (Soylu et al., 2010). Briefly the technique is, after cooling the autoclaved PDA up to 50 °C, under sterilized condition, the estimated amount of essential oils was added to the flasks of the medium. For gaining a homogenous mixture of the culture medium and the essential oil about 0.5% of Tween-80 was added and shaken gently. After pouring the prepared medium into petri plates, 5 mm discs of 7 days old B. cinerea cut with a sterilized borer was placed at the centre of petri plates. For the control only the Tween-80 was added into PDA plates. Three replications were used for each treatment and inoculated petri plates were kept inside an incubator at 20 °C.

After 6 days when the fungal growth reached at the edge of the petri plates, the mycelial growth on the medium in different petri plates was measured horizontally and vertically and was noted down.

2.6 Effect of different essential oil concentrations on spore germination and germ tube elongation of B. cinerea

In this section the method of spore germination assay used by Soylu *et al.* (2010) was employed. A very thin layer of PDA as prepared previously were poured in petri plates so that germinating spores were easily seen under light microscope. About 150 μ l of spore suspension of the fungus at about 10⁴ spores/mL were spread over the thin layer of petri-plate and were kept inside the incubator at 20°C for 12 hr. Three drops of lactophenol cotton blue were spread over the petri plates. About 100 spores were observed and their germ tubes' length was measured. Three replications were used for each treatment and in the case of control only Tween-80 was used.

2.7 Data analysis

Experiments were conducted using 10 treatments (Table 1) and three replications in a completely randomized design. Data were analyzed using Mstat-C software and means were compared using Dunkan's multiple range test at 1 and 5% level of significance.

Treatment code Plant Essential oil concentration (ppm) 1 Wild marjoram 50 2 Wild savory 50 3 50 Savory 4 Wild marjoram 100 5 Wild savory 100 6 Savory 100 7 Wild marjoram 200 8 Wild savory 200 9 Savory 200 10 Control (check)

Table 1. Medicinal plants with the concentration of essential oil used as treatments

3. Results and Discussion

3.1 Amount of Essential oils and their main compounds

The percentage of the main compounds of wild marjoram, wild savory (perennial) savory (annual) essential oils are shown in Table 2.

3.2 Pathogenicity test of B. cinerea on grape berries

After conducting the pathogenicity test on grape berries (Var. Askari), (Fig. 1), the re-isolated fungus was identified as *B. cinerea* with the use of standard identification key provided by Beever and Weeds (2004).

3.3 Effect of different essential oil concentrations on mycelial growth of B. cinerea

All treatments except 50 ppm concentration of savory, essential oil had significant differences in reducing the mycelial growth of *B. cinerea* compared to control (p=0.01). In these test essential oils of wild marjoram and wild savory

at 200 ppm completely (100%) inhibited the mycelial growth of the fungus although at 100 ppm some mycelial growth was observed. Among the plants wild marjoram and wild savory performed better antifungal activity as compared to savory (Table 3 and Fig. 2).

3.4 Effect of different essential oil concentrations on spore germination and germ tube elongation of B. cinerea

In this experiment, the spore germination of *B. cinerea* was reduced by different concentrations of all three medicinal plant essential oils and all treatments had significant difference compared to control except that of savory at 50 ppm which was statistically similar with the control (p = 0.01). With the increase of concentrations, spore germination of the *B. cinerea* was significantly suppressed in such a way that both 100 ppm and more concentration of wild marjoram and wild savory essential oils completely (100%) stopped the spore germination of the fungus and no germ tube was observed (Table 3 and Fig. 3).

Table 2. Tested medicinal plants and their main compounds

Name of medicinal plant	Scientific name	% of main compounds
Wild marjoram	Zataria multifolia	Thymol: 31.10, Carvacrol: 27.49
Wild savory	Satureja mutica	Thymol: 4.04, Carvacrol: 44.10
Savory	Satureja hortensis	Thymol: 0.66, Carvacrol: 50.46



Figure 1. Growth of B. cinerea on grape berries (Var. Askari) during pathogenicity test

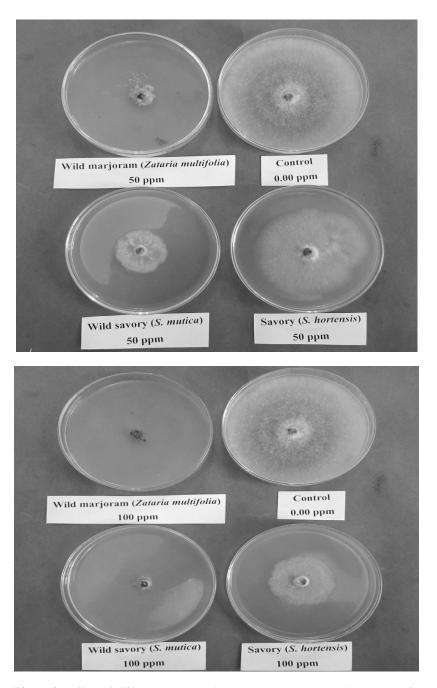


Figure 2. Effect of different essential oil concentrations on mycelial growth of *B. cinerea* compared to control

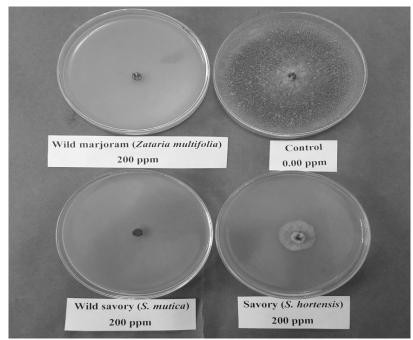


Figure 2 (continued). Effect of different essential oil concentrations on mycelial growth of *B. cinerea* compared to control

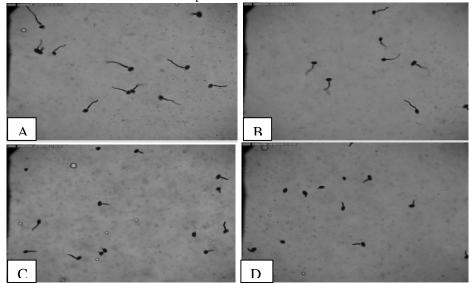


Figure 3. Effect of different essential oil concentrations (50, 100 and 200 ppm respectively) on spore germination and germ tube elongation of *B. cinerea* in PDA compared to control (magnification at 300 X): A, B and C (savory), D, E and F (wild savory), G, H and I (wild marjoram) and J (control)

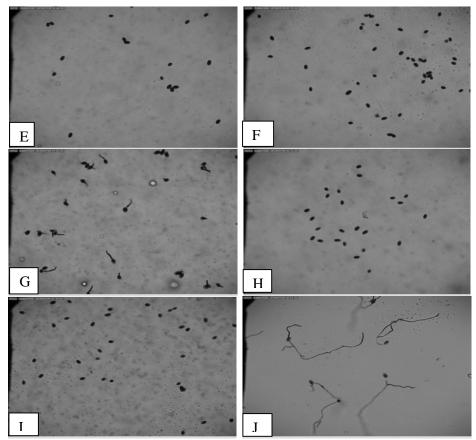


Figure 3 (continued). Effect of different essential oil concentrations (50, 100 and 200 ppm respectively) on spore germination and germ tube elongation of *B. cinerea* in PDA compared to control (magnification at 300 X): A, B and C (savory), D, E and F (wild savory), G, H and I (wild marjoram) and J (control)

 Table 3. Comparison of different treatments means as effect on mycelial growth, spore germination and germ tube elongation of *B. cinerea*

Treatment	Mean mycelial	Mean spore	Mean germ tube
	growth (cm)	germination (%)	length (µ)
Control	9.00 a	100 a	114.4 a
Savory -50 ppm	8.56 a	100 a	44.68 b
Savory -100 ppm	5.21 b	75.00 b	19.93 c
Wild savory -50 ppm	3.70 c	50.80 c	5.15 d
Savory -200 ppm	2.40 d	37.33 d	7.33 d
Wild marjoram-50 ppm	0.98 e	45.00 e	4.50 d
Wild savory -100 ppm	0.81 e	0.00 f	0.00 e
Wild marjoram-100ppm	0.66 e	0.00 f	0.00 e
Wild savory-200 ppm	0.00 f	0.00 f	0.00 e
Wild marjoram-200 ppm	0.00 f	0.00 f	0.00 e

Values in each column followed by the same letter are not significantly different ($P \le 0.01$) according to Duncan's multiple range test.

The thymol and carvacrol were found to be effective against *B. cinerea*; this result was supported by previous research (Lee *et al.*, 2007; Numpaque *et al.*, 2011; Shin *et al.*, 2014; Villanueva-Bermejo *et al.*, 2015; Gavaric *et al.*, 2015). These compounds could be used as effective antifungal compounds and can be safe alternatives for controlling a wide spectrum of microbial plant pathogens. The anti-microbial mechanism of these compounds are not well known but it is believed that such materials cause disorders in vital activity of fungal cell walls or destroy their cell walls (Isman and Machial, 2006).

Results of the current research showed that wild marjoram and wild savory containing more percentage of thymol and carvacrol compounds and had better antifungal activity against B. cinerea. These findings are in agreement with several works by Antonov et al. (1998) who reported that among several plant essential oils, thyme (Thymus capitatus) essential oil at 1% was the best in inhibiting B. cinerea of apple and even clove oil and oil of an Australian mint could not control it as such. Also Etemadi et al. (2012) stated that all three essential oils of Satureja hortensis, Zataria multiflora and Carum copticom at 200 ppm and more were able to effectively control the growth of Botrytis mycelium on both PDA and the actual strawberries fruit. Findings of Shin et al. (2014) also showed that thymol antifungal activity was 4 times more than that of linalool in controlling the gray mold mycelium on table grapes because thymol at 30 µg/mL was equal to linalool at 120 µg/mL in controlling the fungal mycelia. Other workers such as Mermer-Dogu and Zobar (2014) from Turkey and Sesan et al. (2015) from Romania also approved the effectiveness of these compounds in controlling B. cinerea the causal agent of horticultural products.

4. Conclusions

As essential oils produced from wild marjoram and wild savory had inhibitory effect against gray mold fungus (*B. cinerea*), therefore, their suitable formulations such as fumigants could be prepared and be used as safe alternatives to SO_2 smoke for controlling molds of horticultural products in storage.

5. Acknowledgement

We are most grateful to Shahrood Agricultural Research Center authorities, Shahrood, Iran for their financial support to complete this research.

References

- Adams R. P. 2007. Identification of essential oil components by gas chromatography/mass spectroscopy. Allured Publishing corporation. Carol stream. USA. 804pp.
- Ahmadi K., Gholizadeh H., Ebadzadeh H. R., Hatami F., Hosseinpour R., Kazemifrd, R., Abdeshah, H. 2016. Agricultural data for the year 2014-2015. Horticultural products. 3rd Pub. Ministry of Agriculture. Center of information and communication technology. Tehran. Iran. 240 pp. (In Persian).
- Antonov A., Stewart A., Walter M. 1998. Inhibition of conidium germination and mycelial growth of *Botrytis cinerea* by natural products. *Journal of Food Microbiology*, 23: 13-33.
- Beever R. E., Weeds P. L. 2004. Taxonomy and genetic variation of *Botrytis* and *Botryotinia*. pp. 9-52 In: *Botrytis*, *Biology*, *Pathology and Control*, (Eds.) Y. Elad, B. Williamson, P. Tudzynski, N. Delen, Kluwer Academic Publisher. The Netherlands.
- Behdani M., Pooyan M., Abbasi S. 2012. Evaluation of antifungal activity of some medicinal plant essential oils against *Botrytis cinerea*, causal agent of postharvest apple rot, *in vitro*. *International Journal of Agriculture and Crop Sciences*, 4(14): 1012-1016.
- Elad Y., Williamson B., Tudzynski P., Delen N. 2007. Botrytis: Biology, Pathology and

Control. Springer, The Netherlands. 393pp.

- Etemadi N. A., Behdad M., Zeinali H. 2012. Antifungal effects of three plant essential oils against *Botrytis cinerea*: The cause of gray mold on strawberry. *Journal of Research in Agricultural Science*, 8(2): 165-170.
- Fawcett C. H., Spencer D. M. 1966. Antifungal phenolic acids in apple fruits after infection with *Sclerotinia fructigena*. *Annals of Applied Biology*, 60: 87-96.
- Gavaric N., Mozina S. S., Kladar N., Bozin B. 2015. Chemical profile, antioxidant and antibacterial activity of Thyme and Oregano essential oils, Thymol and Carvacrol and their possible synergism. *Journal of Essential Oil Bearing Plants*, 18(4): 1013-1021.
- Ghafari Z., Kazemi-Ghahfarokhi N., Rahimi E. 2011. Presence of ochratoxin a in red and white grape juice commercialized in Iran. *American-Eurasian Journal of Toxicological Sciences*, 3(4): 228-230.
- Isman M. B., Machial C. M. 2006. Pesticides based on plant essential oils: From traditional practice to commercialization. pp. 29-44 In: *Naturally occurring bioactive compounds*, (Eds) M. Rai, M. C. Carpinella, Elsevier, Amsterdam, The Netherlands.
- Jalili-Marandi R., Hassani A., Ghosta Y., Abdollahi A., Pirzad A., Sefidkon F. 2010. *Thymus kotschyanus* and *Carum copticum* essential oils as botanical preservatives for table grape. *Journal of Medicinal Plants Research*, 4(22): 2424-2430.
- Kritzman G., Netzer D. 1978. A selective medium for isolation and identification of *Botrytis* spp. from soil and onion seed. *Phytoparasitica*, 6: 3-7.
- Lee S.O., Choi G. J., Jang K. S., Lim H. K., Cho K. Y., Kim J. C. 2007. Antifungal activity of five plant essential oils as fumigant

against postharvest and soilborne plant pathogenic fungi. *Plant Pathology Journal*, 23(2): 97-102.

- Mermer-Dogu D., Zobar D. 2014. Effects of some plant essential oils against *Botrytis cinerea* and *Tetranychus urticae* on grapevine. *Turkish Journal of Agricultural and Natural Sciences*. Special Issue 1: 1268–1273.
- Numpaque M. A., Oviedo L. A., Gil J. H., García C. M., Durango D. L. 2011. Thymol and carvacrol: biotransformation and antifungal activity against the plant pathogenic fungi *Colletotrichum acutatum* and *Botryodiplodia theobromae*. *Tropical Plant Pathology*, 36 (1): 3-13.
- Şesan T. E., Enache E., Iacomi B. M., Oprea M., Oancea F., Iacomi C. 2015. Antifungal activity of some plant extracts against *Botrytis cinerea* Pers. in the Blackcurrant crop (*Ribes nigrum* L.). Acta Sci. Pol., Hortorum Cultus, 14(1): 29-43.
- Shin M. H., Kim J. H., Choi H. W., Keum Y. S., Chum S. C. 2014. Effect of Thymol and Linalool fumigation on postharvest diseases of table grapes. *Mycobiology*, 42(3): 262–268.
- Siripornvisal S., Rungprom W., Sawatdikarn S. 2009. Antifungal activity of essential oils derived from some medicinal plants against grey mould (*Botrytis cinerea*). *Asian Journal of Food and Agro-Industry*. Special Issue, S229-S233.
- Soylu E. M., Kurt S., Soylu S. 2010. *In vitro* and *in vivo* antifungal activities of the essential oils of various plants against tomato grey mould disease agent *Botrytis cinerea*. *International Journal of Food Microbiology*, 143(3): 183-189.
- Villanueva-Bermejo D., Angelov I., Vicente G., Stateva, R. P., Rodriguez García-Risco M., Reglero G., Ibañez E., Fornari T. 2015. Extraction of thymol from different varieties of thyme plants using green solvents. *Journal of Science of Food and Agriculture*, 95(14): 2901-290.