

An Official Journal of Bangladesh Livestock Research Institute (BLRI)

Bangladesh Journal of Livestock Research

Journal Homepage: https://www.banglajol.info/index.php/BJLR



Review Article

Major respiratory diseases of goat and their epidemiology, prevention and control

Md Habibur Rahman¹, Enam Ahmed², Md Nurul Haque³, Md Zakir Hassan¹ and Md Zulfekar Ali^{1*}

¹Animal Health Research Division, Bangladesh Livestock Research Institute, Savar, Dhaka 1341, Bangladesh; ²Research Farm, Support Service Division, Bangladesh Livestock Research Institute, Savar, Dhaka 1341, Bangladesh; ³Goat Production Research Division, Bangladesh Livestock Research Institute, Savar, Dhaka 1341, Bangladesh

Abstract

Goats and sheep have the capacity to provide meat, milk, leather, and wool, making them valuable assets for the global livestock business, particularly in Southeast Asia, the Mediterranean region, and African nations. Respiratory diseases are the primary cause of death for these animals, accounting for about half of their deaths. Whatever their causes, goat and sheep infectious respiratory diseases account for 5.6% of all small ruminant diseases and disorders. Most of these have an infectious origin, particularly those that are bacterial, parasitic, fungal, or viral. PPR, caprine arthritis encephalitis, and bluetongue are the main viruses that cause respiratory diseases in goats. Through the use of a roadmap, the World Organization for Animal Health (WOAH) has established a global strategy for the eradication of PPR from the planet by 2030. Conversely, the two main bacterial causes of respiratory diseases are Pasteurella and Mycoplasma species, which can co-infect viral disorders or act as the primary cause. They can also exacerbate existing infections and lead to incorrect diagnoses. Numerous surveillances identified fungal infections, particularly Aspergillosis and Conidiobolomycosis, as well as parasite infections such as nasal myiasis and verminous pneumonia. In the review paper, we provide the most recent updated results on such diseases and their causative agents, taking into account elements such as distribution, transmission, spatiotemporal dynamics, possible risk factors, clinical signs and symptoms, treatment, avoidance, and control methods. Lastly, we offer some particular suggestions for addressing the respiratory conditions that affect goats.

Key words: Goat, Respiratory disease, Bluetongue, Pasteurellosis

Bang. J. Livs. Res. Vol. 29 (1&2), 2022: P. 1-20. https://doi.org/10.3329/bjlr.v29i1.72031

Introduction

Goats play an important role as providers of nutrition, food security and socio-economic status to their human owners in many low-income Asian countries, especially the landless small holder farmers in tropical countries (Liang *et al.*, 2019). In Bangladesh, the livestock sector has emerged as one of the key components of national as well as agricultural growth with

^{*}Corresponding: zulfekarvet@gmail.com

an annual contribution of 1.85% of the national GDP and 16.52% share in the agricultural GDP (DLS, 2023). Goat is one of the major economically important livestock species in Bangladesh with a population of about 26.95 million representing 47.15% of Bangladesh's total ruminant population (DLS, 2023). The popularity of goat rearing is being increased in Bangladesh for fulfilling the growing demand of animal protein. But, hot and humid conditions in Bangladesh favour various that impairs diseases productivity of animals and incurs veterinary costs (Rakib et al., 2022; Rahman et al., 2020; Munsi et al., 2018). Respiratory diseases of sheep goats and multifactorial (Lacasta et al., 2008) and are multiple etiological responsible for the respiratory disease complex. In small ruminants, respiratory infections are among the major causes of mortality and subsequent economic losses (Chakraborty et al., 2014). In the USA, the respiratory disease-associated economic loss was estimated to be more than one billion dollars (Nicolas et al., 2008). Interestingly, infectious respiratory disorders of sheep and goats are involved in 5.6% of the total small ruminant diseases and represent approximately 50% of the global death rate (Kumar et al., 2014). Goats and sheep exhibit increased susceptibility to respiratory diseases, including bacteria, viruses, parasites, and fungi, primarily due to inadequate management practices that render these animals more vulnerable to infectious agents. Diseases of the respiratory tract can affect individuals or groups in sheep and goat flocks, resulting in low body weights and an increased death rate (Chakraborty et al., 2014). The aforementioned phenomenon leads to significant economic losses for individuals

engaged in shepherding and goat keeping, reflecting a decline in the production of meat, milk, and wool, as well as a decrease in the number of offspring. The presence of unfavourable weather conditions, hence inducing stress frequently contributes to the development and manifestation of multiple diseases. The unfavorable situation arises when both bacterial and viral infections coexist, especially in adverse environmental circumstances (Lacasta et al., Furthermore, it has been observed that animals that are immunodeficient, being pregnant, lactating, or of old age are particularly susceptible to respiratory infections caused by various pathogens such as Streptococcus pneumoniae, Mannheimia haemolytica, Bordetella parapertussis, Mycoplasma Arcanobacterium species, pyogenes, and Pasteurella species (Brogden et al., 1998). Infections of this type present a significant challenge in the context of the intensive rearing of sheep and goats. Diseases such as Peste des Petits Ruminants (PPR), ovine pulmonary adenomatosis, and bluetongue have a detrimental impact on global trade (Chakraborty et al., 2014; Kumar et al., 2014), consequently hindering economic growth.

However, there is a limited amount of research available on respiratory disorders in goats, and there is a shortage of comprehensive information regarding the present prevalence and distribution of these diseases across a greater geographical area. This work presents a comprehensive overview of various important respiratory diseases affecting goats, with particular emphasis on the context of global as well as Bangladesh. Our effort in this subject has been focused on reviving the study of disease dynamics, covering epidemiology, pathophysiology, present situation, and

future possibilities, with a particular emphasis on diagnostics, treatments, and prophylactics, as well as forthcoming problems or threats. This study reviewed and summarized goat respiratory diseases literature to identify gaps and make recommendations for sustainable goat production in Bangladesh and abroad.

Methodology

The review of various studies holds significant importance in the development of control and preventative strategies. The authors carefully reviewed the currently available knowledge on these important respiratory diseases which were collected and downloaded from various sources. Then provided an in-depth discussion regarding the present prevalence and distribution of these diseases, present situation, particular emphasis on diagnostics, treatments, and prophylactics measures of their implications within the global and Bangladeshi scenario.

Respiratory Diseases of Goat

The classification of infectious respiratory diseases in goats and sheep depending on the involvement of the etiological agents (Kumar *et al.*, 2014; Bell, 2008):

- Bacterial diseases: Pasteurellosis, Mycoplasmosis, Ovine progressive pneumonia, Enzootic pneumonia, and Caseous lymphadenitis
- Viral diseases: PPR, Bluetongue, Caprine arthritis encephalitis (CAE) virus, and Parainfluenza
- Parasitic diseases: Nasal myiasis and Verminous pneumonia
- Fungal diseases: Fungal pneumonia (Conidiobolomycosis, Aspergillosis)

Frequently, the existence of environmental

stress, immunosuppression, and inadequate management procedures can lead to a greater susceptibility of sick individuals to secondary pathogens. Additionally, the occurrence of mixed infections involving many causes is a frequently observed phenomenon (Chakraborty *et al.*, 2014; Scott, 2011). These diseases mostly affect the respiratory tract, with lesions localized either in the upper or lower respiratory tract of the animals (Chakraborty *et al.*, 2014; Daniel *et al.*, 2006). Therefore, these diseases can be divided into the following categories: (Bell, 2008)

Diseases of the upper respiratory tract, namely, nasal myiasis and enzootic nasal tumors, mainly remain confined to sinus, nostrils, and nasal cavity. Various tumors like nasal polyps (adenopapillomas), squamous cell carcinomas. adenocarcinomas, lymphosarcomas, and adenomas are common in upper respiratory tracts of sheep and goats. However, the incidence rate is very low and only sporadic cases are reported.

Diseases of the lower respiratory tract, namely, PPR, parainfluenza, Pasteurellosis, Ovine progressive pneumonia, mycoplasmosis, caprine arthritis encephalitis virus, caseous lymphadenitis, verminous pneumonia, and many others which involve lungs and lesions, are observed in alveoli and bronchioles.

The morbidity and mortality rates in animals of all age groups can vary depending on the severity of the infections and the physical condition of the infected animals. These diseases, either alone or together with other related conditions, may appear as either acute or chronic in nature, and they pose a substantial threat to the livestock industry,

resulting in major economic losses (Chakraborty *et al.*, 2014). Therefore, respiratory diseases can also be categorized based on the disease onset and duration, as shown below (Bell, 2008):

- (1). Acute: PPR, Pasteurellosis, Bluetongue, and Parainfluenza,
- (2).Chronic: Mycoplasmosis, Verminous pneumonia, Nasal myiasis, and Enzootic nasal tumors,
- (3).Progressive: Ovine progressive pneumonia, Caprine arthritis encephalitis virus, Caseous lymphadenitis, and Pulmonary adenomatosis.

Important Viral Respiratory Diseases

Peste des petits ruminants (PPR)

Peste des petits ruminants (PPR) are a highly contagious viral disease akin to rinderpest, afflicting goats, sheep, and certain wild ruminants, with camels among the carriers (Kinne *et al.*, 2010). PPR is marked by high morbidity and mortality and it can reach up to 5–90% and 50–80%, respectively (Chowdhury *et al.*, 2014).

The major clinical signs of PPR affected depression, goats are high fever. mucopurulent discharges from the eye and nose and mouth; acute diarrhea; enteritis; bronchial pneumonia; and erosive stomatitis (Rahman et al., 2020; Chowdhury et al., 2014). A higher frequency of PPR infection is reported during the summer season than other seasons (Rahman et al., 2020). Crucially, it doesn't affect humans. It affects 30 million animals across 70 nations, causing annual economic losses of up to USD 1.2 to 1.7 billion, endangering the livelihoods, food security, and employment of 300 million families, while impeding national and regional development (FAO-OIE, 2015).

Nationwide intensive risk factors and space-time cluster analysis of PPR in Bangladesh found that 27.6% of cases occurred during the monsoon season, with varying proportions by division (Chowdhury *et al.*, 2014). Five space-time clusters and nine hotspots were identified, and PPR incidence remained high in certain regions.

The worldwide eradication of PPR by 2030 is a realistic and cost-effective endeavor. PPR is easily diagnosed, and a single vaccine dose confers lifelong immunity, while the virus has a short survival period outside a host, making it a prime target for eradication efforts (WOAH, 2016). International consensus, political support, technical feasibility, and the successful partnership between FAO and OIE, notably in eradicating rinderpest, all enhance the prospects for success in the global PPR eradication program. The program consists (four) stages, starting 04 epidemiological assessment (stage 1) and culminating in OIE-recognized freedom from PPR (stage 4) (Figure 1). Countries are categorized as "below stage 1" when no epidemiological data is available or "beyond stage 4" when OIE recognition is attained (FAO-OIE, 2015). Bangladesh is actively participating in the PPR Global Eradication Program (2015-2030) and has assessed its progress using the PPR Global Control and Eradication Strategy (GCES), placing it at Stage 2. The country aims to achieve PPR-free status by 2025 (Samad et al., 2019). Despite various control efforts, Bangladesh has experienced recurring PPR outbreaks, causing substantial economic predicted losses US\$25 million small-scale livestock farmers (Begum et al., 2018).



Figure 1. The phases of the worldwide PPR eradication initiative, as outlined by the World Organization for Animal Health (WOAH), leading to a country's official recognition as free from PPR

To effectively combat the disease, it's crucial understand to the virus's pathobiology and molecular evolution. Molecular analysis, particularly of the N gene sequence, has revealed the presence of lineage IV PPRV strains in Bangladesh closely linked to isolates from neighboring countries like Nepal, China, India, Bhutan, and Tibet indicating potential transboundary animal movement (Nooruzzaman et al., 2021; Rahman et al., 2018). PPRV has a single serotype but is divided into four genetic lineages (Banyard et al., 2010). Lineages I and II are in western and central Africa, lineage III is in eastern Africa and the Arabian Peninsula, and lineage IV spans South Asia, the Middle East, and recent parts of Africa (Clarke et al., 2018). Lineage IV reached South Asia, starting in 1987 in southern India and subsequently spreading to Bangladesh in 1993 (Islam et al., 2001), Pakistan in 1994 (Amjad et al., 1996), and Nepal in 1995 (Dhar et al., 2002). These lineages offer insights into the distribution of PPRV strains. Bangladeshi PPRVs, analyzed through phylogenetics, belong to lineage IV and are closely related to strains in China, Tibet (2007-2008), and India (2014-2018) (Nooruzzaman et al., 2021). Predictive analysis identified sixteen epitopes in N, M, F, and hemagglutinin (H) proteins. Notably, N and M protein epitopes match existing vaccines. suggesting Bangladeshi strains could serve as local vaccine options (Nooruzzaman et al., 2021; Rahman et al., 2018).

Vaccination is crucial for Peste des petits (PPR) prevention. Live ruminants attenuated vaccines, such as Nigeria 75/1 (used in Africa and the Middle East) and India's lineage IV strains like Sungri 96, have been effective (Diallo et al., 1989). Bangladesh has its own Titu strain, developed by the Bangladesh Livestock Research Institute, successfully used for PPR control for 20 years (Rahman et al., 2011). There is no particular remedy for PPR, but addressing bacterial and parasitic complications can reduce mortality rates in impacted groups of animals. Additionally, supportive providing care, such administering fluids, antibiotics, and PPR-specific hyperimmune serum, lower the mortality rate in goats infected with PPR (Begum et al., 2018). The global challenge of Peste des Petits Ruminants goat farming (PPR) in needs comprehensive and integrated approach involving the development of suitable diagnostic methods, immunization strategies, and therapeutic protocols for effective mitigation and control.

Bluetongue (BT)

Bluetongue (BT), caused by the Bluetongue virus (BTV), affects goats and sheep and spreads through *Culicoides spp.* primarily (Qin *et al.*, 2018). Infected ruminants, including domestic and wild species, are susceptible. While cattle and goats might carry BTV without symptoms, sheep and deer

often show clinical signs, sometimes leading to high fatality rates (Constable *et al.*, 2017). Goats infected with BTV-8 exhibit fever and mild symptoms like lameness and diarrhea. Trade restrictions and production losses, including mortality and reduced milk/meat output, result in significant economic impacts (Rushton and Lyons, 2015).

Bluetongue virus (BTV), comprising 27 serotypes and diverse strains, exhibits considerable genetic variability affecting its epidemiology. Classified as a "notifiable" disease by the WOAH, BTV impacts both animal health and the economy globally (WOAH, 2021). Present on all continents except Antarctica, the virus perpetuates in enzootic regions, maintaining transmission cycles among vectors and vertebrate hosts. Factors shaping BTV dynamics encompass host, vector, and virus genetics alongside environmental conditions. In goat flocks, morbidity averages 2.7%, mortality 1.2%, and a 45% case fatality rate, notably higher than in sheep (Allepuz et al., 2010). Bangladesh, bordering BTV-endemic India, faces susceptibility due to livestock and Culicoides midge movements. Local factors like species, density, climate, management practices significantly influence BTV transmission dynamics (Courtejoie et al., 2018; Ali et al., 2020). Temperature thresholds regulate outbreaks, while vector dispersal and host migrations facilitate disease transmission between herds.

Bluetongue virus (BTV) affects goats globally, with diverse seroprevalence rates reported across countries. BTV misdiagnosis is common due to clinical similarities with diseases like peste des petits ruminants (PPR), foot rot, foot and mouth disease, pox, and contagious ecthyma (Rahman *et al.*, 2018). Diagnosis involves

molecular methods like ELISA, PCR, serum neutralization tests (SNT), and real-time aiding in early detection serotype-specific identification, albeit with varied sensitivities, costs. and requirements (Mars et al., 2010). These diagnostic tools are vital for monitoring and controlling BTV in livestock populations. Effective bluetongue control involves prevention, supportive care, and vaccination movement restrictions around infected farms helped manage outbreaks (Ensoy et al., 2013). An inactivated vaccine from an Italian bluetongue virus isolate demonstrated success in vaccinated animals (Emidio et al., 2004). Treatment for affected animals focuses on rest, soft food, and managing secondary infections during the healing phase.

Caprine Arthritis and Encephalitis (CAE)

Caprine arthritis-encephalitis is a fatal, widespread multisystemic inflammatory viral disease which is responsible for major production loss in goats (Olech et al., 2020). The causative agent of the disease, caprine arthritis-encephalitis virus (CAEV), is a lentivirus within the group of small ruminant lentivirus (SRLV) belonging to the subfamily retroviridae family and Orthoretrovirinae (Jesse et al., 2018; Thomann et al., 2017). Lentivirus infections cause progressing and fetal infections in different target organs includes carpal joints, mammary glands, lungs and central nervous system (Thomann et al., 2017; Didugu et al., 2016). This might have made a detrimental impact on the amount of milk produced and raised the possibility of developing mastitis (Leitner et al., 2010). In kids, the disease presents as encephalitis, whereas adult goats may experience severe indurative mastitis, arthritis, and occasionally interstitial

pneumonia (Rahman et al., 2023; Jesse et al., 2018). Other clinical manifestations of CAEV infections encompass enlargement of the joint resulting in lameness, synovitis, and decreased growth rate have further been mentioned (Jesse et al., 2018). The arthritic form exhibits a greater occurrence rate in goats, resulting in lameness and an enlargement of joint area (Souza et al., 2015). The economic losses associated with the disease can be attributed to factors such as mortality resulting from clinical ailments, the reduced value of culled animals, the adverse effects of subclinical disease on productivity, and a reduction in the economic lifespan (Norouzi et al., 2015).

Typically, the infection caused by the CAE virus has been observed to spread by both vertical and horizontal transmission routes (Jesse et al., 2018; Thomann et al., 2017). It has been observed that the CAE virus spreads both vertically normally horizontally. Horizontal transmission arises through direct contact with infected animals, bodily fluids, and excretions; vertical transmission arises by consumption of contaminated milk or colostrums (Jesse et al., 2018; Thomann et al., 2017). However, CAEV's of the mechanism sexual transmission remains unclear, in contrast to other lentiviruses (Al Ahmad et al., 2008). CAEV cause disease mainly in goats. Once the virus enters the body, the host becomes permanently infected. In mixed populations, there have also been reports of natural cross-species transmission from sheep to goats and from goats to sheep (Gjerset et al., 2009). There is no proof that the caprine arthritis-encephalitis virus (or any SRLV) may infect people. Following its first discovery in 1974 in goats, reports of CAE virus infection have been made in various countries globally (Cork et al., 1974). The

prevalence of infection in endemic regions can vary widely, ranging from < 5% to more than 60% and has been recorded from Thailand 5.52% (Nyi Lin *et al.*, 2011), Turkey 7.5% (Azkur *et al.*, 2011), Mexico 3.6% (Torres-Acosta *et al.*, 2003), Italy 40% (Gufler and Baumgartner, 2007), Malaysia 8.8% (Jesse *et al.*, 2018), Jordan 18. 5% (Hailat *et al.*, 2022), Brazil 8.2% (Bandeira *et al.*, 2009), Iraq 8.69% (Hamzah and Mosa, 2020), India 4.5% (Didugu *et al.*, 2016), America 31% (De Groot *et al.*, 1993), Norway 42% (Nord and Ådnøy, 1997) and also in Bangladesh 4.26% (Rahman *et al.*, 2023).

The disease diagnosis incorporates a blend of clinical manifestations, postmortem results and histopathological observations (Jesse et al., 2018). Serological testing is the primary tool used to detect the MV/CAEV virus because it takes into account the longevity of circulating antibodies against the virus. For serological diagnosing of CAEV, these procedures consist of indirect immunofluorescence, enzyme-linked immunosorbent assay (ELISA), and agar gel immuno diffusion (AGID) (Norouzi et al., 2015). In practical applications, the most common serological test used to diagnose CAEV infection is the enzyme-linked immunosorbent assay (ELISA). Since ELISA is more sensitive than AGID, it is the preferred method (Jesse et al., 2018).

There is currently no effective treatment or vaccination for this disease, but improvements in the quality and efficacy of diagnostic tests may be able to eradicate it (Norouzi *et al.*, 2015). Hence, there is neither a vaccination nor a treatment for CAE, so preventative measures are crucial. These include (i) separating kid from their seropositive dams as soon as they are born,

(ii) using heat-treated or SRLV-negative colostrum, (iii) increasing the culling rate, and (iv) conducting routine serological surveillance and separating seropositive goats right away (Cirone *et al.*, 2019). Antibiotics may be prescribed for secondary bacterial infections in cases of mastitis or pneumonitis (www.cfsph.iastate.edu). Therefore, the rapid diagnosis of the disease via a serological test remains an essential technique for controlling, preventing, and eliminating CAEV infection (Brinkhof *et al.*, 2010).

Important Bacterial Respiratory Diseases

Pneumonic Pasteurellosis

8

Pneumonic pasteurellosis, commonly known as respiratory mannheimiosis, is a widespread and acute respiratory tract infection in ruminant animals, leading to significant financial losses due to death, reduced weight gain, delayed marketing, treatment costs, and reduced overall health among survivors (Dereje et al., 2014). It is caused by bacteria from the Pasteurellaceae family, including Mannheimia haemolytica, Biberstinia trehalosi, and Pasteurella multocida, which become problematic when the host's defense mechanisms compromised. These bacteria are typically commensals in the tonsils nasopharyngeal microflora of healthy goats (Legesse et al., 2018; Belege et al., 2017). Pneumonic pasteurellosis is prevalent in temperate both and tropical regions worldwide, affecting goats in hot, humid lowland areas as well as highlands, leading to high morbidity and mortality rates (Belege et al., 2017). While M. haemolytica is mostly limited to ruminants, P. multocida has a broader host range (Quinn et al., 2002). Many species within the Pasteurellaceae family colonize the mucous

membranes of the genital, respiratory, and alimentary tracts in various mammals, birds, and reptiles. Controlling and preventing pneumonic pasteurellosis is essential for preserving the health and productivity of ruminant livestock (Dousse et al., 2008). Pasteurellosis, or pneumonic pasteurellosis, spreads through inhalation of infected droplets coughed up or exhaled by infected animals, affecting both carriers and clinical cases. Mannheimia haemolytica Pasteurella multocida. typically upper respiratory can commensals. become opportunistic pathogens when animals are immunocompromised (Belege et al., 2017). Aerosols serve as a means of exogenous transmission (Quinn et al., 2002). These bacteria are highly sensitive environmental factors, making conditions crucial in disease spread. Goats, with their group-rearing and huddle-prone behavior, are particularly susceptible to contagious diseases (Chakraborty et al., Pneumonic pasteurellosis in goat manifests with a wide range of clinical symptoms, including intermittent coughing, sudden death, or acute respiratory distress (Afata, 2018). Clinical signs usually appear within 10-14 days of stress exposure, although earlier onset is common. In acute outbreaks, the clinical course is relatively brief, lasting 2-3 days, resulting in death, recovery, or treatment-related outcomes. Infected goats exhibit dullness, reduced appetite, depression, high fever, coughing, nasal discharge, and anorexia, with productive coughs developing later in most cases (Legesse et al., 2018; Mohamed and 2008). Dyspnea with Abdelsalam, expiratory grunt can be observed in advanced stages. Pneumonic pasteurellosis may cause severe bronchopneumonia, pleurisy, septicemia, and acute febrile courses. Young goats are more susceptible,

and severe infections can lead to sudden death (Mohamed and Abdelsalam, 2008). The disease primarily arises from physical or physiological stress due to unfavorable environmental and climatic factors, such as extreme weather. poor management, crowded living conditions, inadequate transportation, or prior infections with respiratory viruses or mycoplasma (Belege et al., 2017; Dereje et al., 2014). Pneumonic pasteurellosis mainly affects animals with compromised lung defenses and can lead to morbidity rates of up to 35% and case fatality rates ranging from 5% to 10% in small ruminants (Belege et al., 2017). The prevalence of pneumonic pasteurellosis varies in different countries, with rates reported from 22.8% to 58.13% in India (Makwana et al., 2022), 4% for Pasteurella multocida and 2.6% for Mannheimia haemolytica in Iraq (Ahmed et al., 2022), and rates ranging from 21.9% to 44% in Ethiopia (Assefa al..et 2018). Bangladesh, various studies have reported the presence of Pasteurella spp. and P. multocida and Mannheimia haemolytica in pneumonic goats, with prevalence rates of 16.67% (Ahmed et al., 2019), 20% (Sen et al., 2018), 15% (Rashid et al., 2013), 20% (Momin et al., 2011), and 48.57 % (Rahman et al., 2023) in different regions.

Pneumonic pasteurellosis is a significant concern affecting goat populations in many countries, leading to economic losses and necessitating proper management and control measures. These bacteria typically part of the upper respiratory tract's microbiota and often coexist with other pathogens. Antibiotic resistance growing concern, making sensitivity testing (Kehrenberg et al., Gentamicin and Ceftriaxone group were comparatively more sensitive group of

antibiotics and some symptomatic treatments should be given (Rahman et al., 2023). Effective control strategies include stress reduction, early diagnosis, antibiotic treatment, vaccination, and biosecurity measures (Afata, 2018). Vaccination is a and cost-effective approach, practical particularly in developing countries. although challenges exist due to multiple serotypes of Pasteurella and Mannheimia without cross-protection. Integrating these techniques is essential for effective pneumonic pasteurellosis control in goats and sheep (Belege et al., 2017).

Contagious caprine pleuropneumonia (CCPP)

Contagious caprine pleuropneumonia (CCPP) is a highly transmissible and fast-spreading mycoplasmal respiratory disease affecting goat populations and causing significant financial damage in many parts of the world, especially in Africa and Asia (Yatoo et al., 2019). The disease is caused by Mycoplasma capricolum subsp. capripneumoniae, formerly strain F38, a member of the Mycoplasma mycoides cluster (Ahaduzzaman, 2021). It is the smallest fastidious bacteria size is only 300nm which has a triple layer membrane but lacks cell wall (Wazir et al., 2016). CCPP induced catastrophic harm through direct production losses like high infection rates, death of animals and reduced productivity or indirect losses such as treatment costs, monitoring, disease diagnosis, restrictions on trade and minimal animal price (Solangi et al., 2023; Yatoo et al., 2019). CCPP resulting from Mccp contributes to 100% morbidity in goat herds and 60-80% of mortality occurs (Solangi et al., 2023). The disease is listed by the World Organisation for Animal Health (WOAH, 2020; Ahaduzzaman, 2021) as one of the

diseases that must be reported. An estimated US\$ 507 million is spent annually on CCPP worldwide (Yatoo et al., 2019). Acute infection is clinically characterised by a high fever (40.5-41.5°C), anorexia, depression, dyspnea, difficulty breathing, coughing, grunting, and, within five days, the death of unrecovered animals (Teshome et al., 2019; Yatoo et al., 2019). Chronic and per-acute types are less frequent. When an infection is per-acute, animals can pass away from it in 1 to 3 days with no or few clinical symptoms; when it is chronic, symptoms are less apparent and include just minor nasal discharge and coughing that occurs after physical activity (Ahaduzzaman, 2021). Only domestic and wild ruminants are susceptible to CCPP and domestic goats are the primary CCPP hosts (Ahaduzzaman, 2021; Yatoo et al., 2019). However, Teshome et al. (2019) have recorded cases of CCPP in sheep that are kept in close vicinity to goats. All ages and sexes of animals may be affected (Bölske et al., 1996). Close contact with infected animals may transmit the highly contagious CCPP virus quickly through oculo-nasal discharge, bruising production, bare joints, feces, urine, and inhaling droplets that are contaminated (Ahaduzzaman, 2021; Yatoo et al., 2019). In the acute stage of the disease, profuse mucopurulent to fibrinopurulent exudate develops in fibrinous pleuropneumonia, indicating that CCPP's pathogenicity is strictly restricted to the lungs and pleura (Wesonga et al., 2004). No zoonotic importance exists for CCPP (Ahaduzzaman, 2021).

In CCPP, different methods are used for diagnosis. Pleural fluid, which has a high concentration of the pathogenic organism, and a portion of the infected lung at the interface between the affected and healthy tissues are the preferable samples for the diagnosis of CCPP (Ahaduzzaman, 2021). Appropriate diagnostic tests are then used to detect the antigen or antibody. Clinical samples from the nose, throat, pleural fluid, and lung tissue are subjected to the fast and effective polymerase chain reaction (PCR) method (Bölske et al., 1996). Complement fixation (CFT), test counter-immunoelectrophoresis, enzyme-linked immunosorbent (ELISA), indirect hemagglutination (IH), and latex agglutination test (LAT) are further diagnostic tests for CCPP. Although culturing bacteria is challenging, is regarded isolation of bacteria confirmation of the causal agent. There is currently little epidemiological research on the frequency of CCPP. CCPP is a significant transboundary disease, reports of the occurrence and prevalence of CCPP vary widely. The prevalence of CCPP was recorded from Somalia 49% (Abdi et al., 2020), Ethiopia 7.1% (Gelagay et al., 2007), Oman 28% (Hussain et al., 2021), India 8.11% (Jain et al., 2015), Nepal 3.37% (Adhikari et al., 2022), Kenya 48.6% (Kipronoh et al., 2016) and Tajikistan 10.1% (Amirbekov et al., 2010). Because certain regions are endemic and some are neglected, prevalence estimations are frequently affected by the local epidemiological factors as well as the specificity and sensitivity of the diagnostic tests applied (Ahmed et al., 2020). Different authors have reported on a variety of therapeutic studies using antibiotics with varying degrees of success, such as oxytetracycline and streptomycin, fluoroquinolones (e.g., danofloxacin), and the macrolide family (Dereje and Teshale, 2021; Yatoo et al., 2019). But for treating local and systemic Mycoplasma infections in sheep, goats, cattle, and swine,

macrolides, particularly tylosin are thought to be the most effective drug against Mccp (Yatoo et al., 2019). Additionally, it was found that danofloxacin was very useful in treating clinical CCPP in goats (Ozdemir et al., 2006). The primary methods used for prevention control **CCPP** and vaccination, quarantine, limitations on movement, slaughtering exposed and sick and complete cleaning disinfection of the premises (Dereje and 2021: Yatoo *et al.*, 2019). However, in most of these countries are not available currently. vaccines Worldwide efforts are needed to develop suitable diagnostic. preventive. therapeutic strategies for the prevention and control of CCPP in order to combat this new and serious danger to goat husbandry internationally.

Important Parasitic Respiratory Disease

Verminous Pneumonia

Lungworm infection, known as verminous bronchitis or pneumonia, affects the lower respiratory tract due to various nematodes like Dictyocaulus filarial, Protostrongylus rufescens, and Muellerius capillaris (Zafari et al., 2022). Clinical signs include coughing, dyspnea, and weight loss, varying based on infection severity, age, and immune status (Asmare et al., 2018). Its prevalence in tropical regions averages 19-22%, with limited studies in Bangladesh. Pakistan and Turkey reported infection rates of 31% in sheep, 11% in goats (Ahmed, 2020), and 62.5% in post-mortem sheep exams, 45.1% in faecal investigations (McCarthy et al., 2022). Diagnosis involves Baermann technique and ELISA tests, although interpretation complexities exist (Zafari et al., 2022; Asmare et al., 2018). Anthelmintic drugs and oral vaccinations

are primary management methods, emphasizing preventive measures via grazing management to curb reinfection through larvae surviving on pasture or intermediate hosts like snails and slugs (McCarthy *et al.*, 2022).

Important Fungal Respiratory Disease

Aspergillosis

Pneumonia caused by Aspergillus spp. has been documented in goats, particularly in young individuals, resulting in symptoms such as lethargy, loss of appetite, coughing, difficulty breathing, and discharge from the nose. The fungi discussed in this study are responsible for producing many diseases, such sinonasal and sino-orbital as aspergillosis, diarrhoea, pneumonia, mastitis, and abortion in ruminant animals (Carmo et al., 2014). The aetiology and epidemiology of a given phenomenon are of paramount importance in academic research and analysis. Understanding the underlying causes and factors contributing to the development and occurrence of a Fungi belonging to the taxonomic genus Aspergillus (order Eurotiales. family Trichocomaceae) are saprophytic organisms that have a broad distribution across several natural habitats. The transmission of spread takes place by the dispersion of spores in the form of aerosols, which can be found in soil, decomposing plant matter, and, on occasion, animal tissues. Aspergillosis, a disorder observed in goats, can manifest as a nasal form, leading to a decline in physical well-being due to necrosis of the nasal mucosa and nasal turbinate bones. This pathological process can ultimately result in significant respiratory distress (Elad and Segal, 2018). Cutaneous nodules, typically ranging from 0.3 to 3 cm in diameter,

occasionally exhibiting ulceration, can be detected in the dorsal nasal region and ears. Prolonged utilisation of antimicrobials, concurrent presence of chronic ailments, and consumption of feed contaminated with mould have been identified as contributing factors in the development of pulmonary aspergillosis in ruminant animals. Mastitis caused by Aspergillus spp. has also been documented in goats. The hyphae of Aspergillus species exhibit a slender morphology, measuring between 3 to 12 um in width. They are characterised by their distinct separation, dichotomous branching pattern, and parallel margins. The staining of these entities can be achieved using the Periodic Acid-Schiff (PAS) and Gomori Methenamine Silver (GMS) techniques, as well as visualized in sections stained with Hematoxylin and Eosin (H&E) (Baker and Bennett, 2007).

Conclusion and Recommendations

Goats with respiratory diseases perform poorly in production and mortality, which costs farmers a lot of economic loss. Animal welfare is also significantly impacted by the diseases. In goats and sheep, respiratory are frequently caused disorders unfavorable weather, physiological stress, bacterial, viral and parasitic infections. For the common respiratory diseases that afflict goat, it is crucial to evaluate clinical diagnostic techniques, treatment choices, and preventative measures critically. This paper reviewed the overall scenario of important respiratory diseases of goats in global and Bangladesh perspective. However, we can share specific recommendations for the prevention and control of respiratory diseases of goats are as follows:

• Keep the herd dry, clean and follow the good farm practice.

- Advances in early diagnosis of respiratory diseases of small ruminants lead to identification of affected animals and isolation from the herd.
- Control of the route of transmission of pathogens through proper farm biosecurity guidelines and herd management plan.
- Follow the proper quarantine guidelines during incorporating new animals to the herd.
- Strengthen and easy access to veterinary services of the farmers.
- Surveillance: regular surveillance of diseases and monitoring of disease trends is significant for correct policy decisions to prevent the respiratory diseases.
- Vaccination: regular vaccination against endemic diseases with a proper vaccination plan is quite important.

References

- Abdi, O.M., Jama A.M.S., Jama, A.A., Omar, A.A., Dubad, A.B. 2020. Sero-Prevalence of Contagious Caprine Pleuropneumonia in Goats in Afgoye District Lower Shabelle Region, Somalia. J. Vet. Res., 5(2):000203.
- Adhikari, B.K., Subedi, D., Jyoti, S., Kaphle, K., Kharel, C.N. and Khanal, D.R. 2022. Seroprevalence of contagious caprine pleuropneumonia (CCPP) in Rupandehi and Palpa Districts of Nepal. Vet. Sci.: Research and Rev., 8(1):23-29.
- Afata A.W. 2018. Isolation and identification of Mannheimia haemolytica, Bibersteinia trehalosi and Pasteurella multocida from cattle and sheep from selected areas of

- Ethiopia. MSc Thesis, Addis Ababa University, Ethiopia.
- Ahaduzzaman, M.D. 2021. Contagious caprine pleuropneumonia (CCPP): A systematic review and meta-analysis of the prevalence in sheep and goats. Transbound. Emerg. Dis., 68(3):1332-1344.
- Ahmed, A.G.M., Bakri, E.O., Hussien, M.O., Taseen, M.E.E., Ahmed, A.M. and Abdalla, M.A. 2020. Molecular detection and risk factors of african horse sickness virus (ahsv) in different governorates of Sudan. J. Anim. Health Prod., 8(4):199-205.
- Ahmed, B.A., Abdullah, M.A. 2022. Isolation and molecular diagnosis of the main bacterial species causing Pneumonia in small ruminants in the Duhok Abattoir-Kurdistan region of Iraq. Microb. Biosyst., 7(2):66-73.
- Ahmed, S. 2020. Parasites of markhor, urial and Chiltan wild goat in Pakistan. Ann. Parasitol., 66(1).
- Ahmed, S.J., Hasan, A., Islam, M.R., Shawan, M.M.A.K., Uddin, F., Rahman, N., Hossain, M., 2019. Incidence and Antibiotic Susceptibility Profile of Pasteurella maltocida Isolates Isolated from Goats in Savar Area of Bangladesh. Agric. Sci. Dig., 39:357-360.
- Al Ahmad, M.A., Fieni, F., Pellerin, J.L., Guiguen, F., Cherel, Y., Chatagnon, G., Bouzar, A.B., Chebloune, Y. 2008. Detection of viral genomes of caprine arthritis-encephalitis virus (CAEV) in semen and in genital tract tissues of male goat. Theriogenol., 69: 473-480.
- Ali, M.Z., Carlile, G., Giasuddin, M. 2020.

- Impact of global climate change on livestock health: Bangladesh perspective. Open Vet. J., 10(2):178-188.
- Allepuz, A., García-Bocanegra, I., Napp, S., Casal, J., Arenas, A., Saez, M., González, M.A. 2010. Monitoring bluetongue disease (BTV-1) epidemic in southern Spain during 2007. Prev. Vet. Med., 96(3-4):263-271.
- Amirbekov, M., Murvatulloev, S., Ferrari, G. 2010. Contagious caprine pleuropneumonia detected for the first time in Tajikistan. EMPRES Transboundary Anim Dis Bull, 35:20-22.
- Amjad, H., Qamar Ul, I., Forsyth, M., Barrett, T., Rossiter, P.B. 1996. Peste des petits ruminants in goats in Pakistan. Vet. Rec. 139, 118–119.
- Anna Rovid Spickler. Veterinary Specialist, Center for Food Security and Public Health.(https://www.cfsph.iastate.edu/ Factsheets/pdfs/maedi_visna_and_cap rine_arthritis_encephalitis.pdf) (Accessed on 30 October, 2023).
- Asmare, K., Sibhat, B., Haile, A., Sheferaw, D., Aragaw, K., Abera, M., Abebe, R., Wieland, B., 2018. Lungworm infection in small ruminants in Ethiopia: Systematic review and meta-analysis. Vet. Parasitol.: Regional Studies and Reports, 14: 63-70.
- Assefa, G.A., Kelkay, M.Z. 2018. Goat pasteurellosis: serological analysis of circulating Pasteurella serotypes in Tanqua aberegelle and Kola Tembien Districts, Northern Ethiopia. BMC Res. Notes, 11(1): 1-5.

- Azkur, A.K., Gazyagci, S., Aslan, M.E. 2011. Serological and epidemiological investigation of bluetongue, Maedi-Visna and caprine arthritis-encephalitis viruses in small ruminant in Kirikkale District in Turkey.
- Baker, S.E., Bennett, J.W. 2007. An overview of the genus Aspergillus. The Aspergilli: Genomics, medical aspects, biotechnology, and research methods. 7: 3-13.
- Bandeira, D.A., de Castro, R.S., Azevedo, E.O., Melo, L.D.S.S., de Melo, C.B. 2009. Seroprevalence of caprine arthritis—encephalitis virus in goats in the Cariri region, Paraiba state, Brazil. Vet. J. VET, 180(3): 399-401.
- Banyard, A.C., Parida, S., Batten, C., Oura, C., Kwiatek, O., Libeau, G., 2010. Global distribution of peste des petits ruminants virus and prospects for improved diagnosis and control. J. Gen. Virol., 91:2885–2897.
- Begum, S., Nooruzzaman, M., Parvin, M., Mohanto, N., Parvin, R., Islam, M.R., Chowdhury, E.H. 2018. Peste des petits ruminants virus infection of Black Bengal goats showed altered haematological and serum biochemical profiles. Onderstepoort J. Vet. Res., 85:e1–e10.
- Belege, T., Alamirew, K., Ketema, A., Kiflie, W., Endashaw, M. 2017. Ruminant pneumonic pasteurellosis: Review on epidemiology, pathogenesis and virulence mechanism. Acad. J. Anim. Dis., 6(2): 30-39.
- Bell, S. 2008. Respiratory disease in sheep: 2. Treatment and control. In practice,

- 30(5):278-283.
- Bölske, G., Mattsson, J.G., Bascuñana, C.R., Bergström, K., Wesonga, H., Johansson, K.E. 1996. Diagnosis of contagious caprine pleuropneumonia by detection and identification of Mycoplasma capricolum subsp. capripneumoniae by PCR and restriction enzyme analysis. J. Clin. Microbiol., 34(4):785-791.
- Brinkhof, J.M.A., Moll, L., Van Maanen, C., Houwers, D.J. 2010. Use of serology and polymerase chain reaction for the rapid eradication of small ruminant lentivirus infections from a sheep flock: A case report Res. Vet. Sci., 88(1):41-43.
- Brogden, K.A., Lehmkuhl, H.D. and Cutlip, R.C. 1998. Pasteurella haemolytica complicated respiratory infections in sheep and goats. Vet. Res., 29(3-4):233-254.
- Carmo, P.M.S.D, Portela, R.A., de Oliveira-Filho, J.C., Dantas, A.F.M., Simões, S.V.D., Garino Jr, F., Riet-Correa, F. 2014. Nasal and cutaneous aspergillosis in a goat. J. Comp. Pathol., 150(1): 4-7.
- Chakraborty, S., Kumar, A., Tiwari, R., Rahal, A., Malik, Y., Dhama, K., Pal, A., Prasad, M., 2014. Advances in diagnosis of respiratory diseases of small ruminants. Vet. Med. Int., 2014.
- Chowdhury, E.H., Bhuiyan, A.R., Rahman, M.M., Siddique, M.S.A., Islam, M.R. 2014. Natural peste des petits ruminants virus infection in Black Bengal goats: virological, pathological and immunohistochemical investigation. BMC Vet. Res.,10(1):1-10.

- Cirone, F., Maggiolino, A., Cirilli, M., Sposato, A., De Palo, P., Ciappetta, G., Pratelli, A. 2019. Small ruminant lentiviruses in goats in southern Italy: Serological evidence, risk factors and implementation of control programs. Vet. Microbiol., 228:143-146.
- Clarke, B.D., Islam, M.R., Yusuf, M.A., Mahapatra, M., Parida, S. 2018. Molecular detection, isolation and characterization of Peste-des-petits ruminants virus from goat milk from outbreaks in Bangladesh and its implication for eradication strategy. Transbound. Emerg. Dis. 65:1597–1604.
- Constable, P.D., Hinchcliff, K.W., Done, S.H., Grünberg, W. 2017. Veterinary Medicine: A Textbook of the Diseases of Cattle, Horses, Sheep, Pigs, and Goats, 11th ed. Saunders/ Elsevere, London, UK.
- Cork, L.C., Hadlow, W.J., Crawford, T.B., Gorham, J.R., Piper, R.C. 1974. Infectious leukoencephalomyelitis of young goats. J. Infect. Dis., 129(2): 134-141.
- Courtejoie, N., Zanella, G., Durand, B., 2018. Bluetongue transmission and control in Europe: A systematic review of compartmental mathematical models. Prev. Vet. Med., 156:113-125.
- Daniel, J.A., Held, J.E., Brake, D.G., Wulf, D.M., Epperson, W.B. 2006. Evaluation of the prevalence and onset of lung lesions and their impact on growth of lambs. Am. J. Vet. Res., 67(5):890-894.
- De Groot, B., Batra, J.C., Soni, R.L. 1993. Approaches and constraints in

- veterinary health care for improvement of goat production in Rajasthan In: Management of Small Ruminants Diseases Proceedings of a workshop.
- Dereje, A., Tesfaye, S., Tadesse, B. 2014. Isolation and identification of Mannhemia and Pasturella species from pneumonic and apparently healthy cattle and their antibiogram susceptibility pattern in Bedelle District, Western Ethiopia. J. Bacteriol. Res., 6(5):32-41.
- Dereje, T., Teshale, S. 2021. Contagious caprine pleuropneumonia: A review. Vet. Anim. Sci., 13(3):132-143.
- Dhar, P., Sreenivasa, B.P., Barrett, T., Corteyn, M., Singh, R.P., Bandyopadhyay, S.K. 2002. Recent epidemiology of peste des petits ruminants virus (PPRV). Vet. Microbiol., 88:153–159.
- Didugu, H., Sagi, S., Reddy, C.E.N., Kishore, K.N., Reddy, M.V., Vishnu, P.G. 2016. First report of Maedi-Visna and Caprine Arthritis-Encephalitis viruses in Krishna district, Andhra Pradesh, India. J Anim Res., 6(1):177-179.
- DLS. 2023. Livestock Economy at a glance 2022-2023, Department of Livestock Service (DLS), Ministry of Fisheries and Livestock, Bangladesh. http://www.dls.gov.bd/site/page/22b1 143b-9323-44f8-bfd8-647087828c9b/ Livestock-Economy (accessed on 15 August, 2023).
- Dousse, F., Thomann, A., Brodard, I., Korczak, B.M., Schlatter, Y., Kuhnert, P., Miserez, R., Frey, J. 2008. Routine phenotypic identification of bacterial

- species of the family Pasteurellaceae isolated from animals. J. Vet. Diagn. Invest., 20(6):716-724.
- Elad, D., Segal, E. 2018. Diagnostic aspects of veterinary and human aspergillosis. Front. Microbiol., 9:1303.
- Emidio, B., Nicolussi, P., Patta, C., Ronchi, G.F., Monaco, F., Savini, G., Ciarelli, A., Caporale, V. 2004. Efficacy and safety studies on an inactivated vaccine against bluetongue virus serotype 2. Vet. Ital., 40(4): 640-644.
- Ensoy, C., Aerts, M., Welby, S., Van der Stede, Y., Faes, C. 2013. A dynamic spatiotemporal model to investigate the effect of cattle movements on the spread of bluetongue BTV-8 in Belgium. PloS One 8: e78591.
- Food and Agriculture Organization (FAO); World Organization for Animal Health (OIE). Global Strategy for the Control and Eradication of PPR; FAO: Rome, Italy; OIE: Paris, France, 2015.
- Gelagay, A., Teshale, S., Amsalu, W., Esayas, G. 2007. Prevalence of contagious caprine pleuropneumonia in the Borana pastoral areas of Ethiopia. Small Rumin. Res., 70(2-3): 131-135.
- Gjerset, B., Rimstad, E., Teige, J., Soetaert, K., Jonassen, C.M. 2009. Impact of natural sheep–goat transmission on detection and control of small ruminant lentivirus group C infections. Vet. Microbiol., 135(3-4):231-238.
- Gufler, H., Baumgartner, W. 2007. Overview of herd and CAEV status in dwarf goats in South Tyrol, Italy. Vet Q., 29(2): 68-70.
- Hailat, N.Q., Algharaibeh, T.B., Al-Eitan,

- L.N., 2022. Pathological, molecular, and serological study of small ruminant lentiviruses in Jordan. Vet. World, 15(6): 1423.
- Hamzah, K.J., Mosa, A.H. 2020. Clinical and serological diagnosis of caprine arthritis encephalitis virus (CAEV) in goats of middle Iraq regions. Life Sci. Arch., 6:1771–7.
- Hussain, M.H., Asi, M.N., Al-Uahmadi, S.S.R., Al-Subhi, A.H.A., Al-Senaidi, N.Y.A., Al-Subhi, R.S.N., Al-Beloushi, M.K.I., Al-Sinani, F.S.S., Al-Riyami, B.S.T., Mansoor, M.K., Saqib, M. 2021. Seroprevalence and Associated Risk Factors of Contagious Caprine Pleuropneumonia in the Small Ruminants of Oman. Pak. Vet. J., 41(1).
- Islam, M.R., Shamsuddin, M., Rahman, M.A., Das, P.M., Dewan, M.L. 2001. An outbreak of peste des petits ruminants in Black Bengal Goats in Mymensngh, Bangladesh. Bangladesh Vet 18:14–19.
- Jain, U.D.I.T., Verma, A.K., Pal, B.C. 2015. Occurrence of mycoplasma infection in Barbari goats of Uttar Pradesh, India. Haryana Vet., 54(1): 53-55.
- Jesse, F.F.A., Bitrus, A.A., Abba, Y., Raju, V.N., Hambali, I.U., Peter, I.D., Lila, M.A.M., Norsidin, J.M. 2018. Seroprevalence of small ruminant caprine arthritis encephalitis lentivirus among goats from selected small ruminant farms in Selangor, Malaysia. Vet. World, 11(2):172.
- Kehrenberg, C., Schulze-Tanzil, G., Martel, J.L., Chaslus-Dancla, E., Schwarz, S. 2001. Antimicrobial resistance in Pasteurella and Mannheimia:

- epidemiology and genetic basis. Vet. Res., 32(3-4):323-339.
- Kinne, J., Kreutzer, R., Kreutzer, M., Wernery, U., Wohlsein, P. 2010. Peste des petits ruminants in Arabian wildlife. Epidemiol. Infect., 138:1211–1214.
- Kipronoh, A.K., Ombui, J.N., Kiara, H.K., Binepal, Y.S., Gitonga, E. and Wesonga, H.O. 2016. Prevalence of contagious caprine pleuro-pneumonia in pastoral flocks of goats in the Rift Valley region of Kenya. Trop. Anim. Health Prod., 48:151-155.
- Kumar, A., Rahal, A., Chakraborty, S., Verma, A.K., Dhama, K. 2014. Mycoplasma agalactiae, an etiological agent of contagious agalactia in small ruminants: a review. Vet. Med. Int., 2014.
- Kumar, A., Tikoo, S.K., Malik, P., Kumar, A.T., 2014. Respiratory diseases of small ruminants. Vet. Med. Int., 2014.
- Kumar, N., Maherchandani, S., Kashyap, S.K., Singh, S.V., Sharma, S., Chaubey, K.K., Ly, H. 2014. Peste des petits ruminants virus infection of small ruminants: A comprehensive review. Viruses, 6:2287–2327.
- Legesse, A., Abayneh, T., Mamo, G., Gelaye, E., Tesfaw, L., Yami, M., Belay, A. 2018. Molecular characterization of Mannheimia haemolytica isolates associated with pneumonic cases of sheep in selected areas of Central Ethiopia. BMC Microbiol., 18:1-10.
- Leitner, G., Krifucks, O., Weisblit, L., Lavi, Y., Bernstein, S., Merin, U. 2010. The effect of caprine arthritis encephalitis

- virus infection on production in goats. Vet. J. Vet., 183(3): 328-331.
- Liang, J.B., Paengkoum, P. 2019. Current status, challenges and the way forward for dairy goat production in Asia–conference summary of dairy goats in Asia. Asian J. Anim. Sci., 32(8):1233.
- Makwana, P.M., Desai, D.N., Patel, D.R., Kalyani, I.H., Sakhare, P.S., Muglikar, D.M., Patel, N. 2022. Detection of antibodies to peste des petits ruminants (PPR) virus and Manheimia haemolytica from pneumonic goats of South Gujarat region. Haryana Vet., 61:122-124.
- Mars, M.H., Van Maanen, C., Vellema, P., Kramps, J.A., Van Rijn, P.A. 2010. Evaluation of an indirect ELISA for detection of antibodies in bulk milk against bluetongue virus infections in the Netherlands. Vet. Microbiol., 146(3-4):209-214.
- McCarthy, C., Vineer, H.R., Morgan, E.R., van Dijk, J. 2022. Predicting the unpredictable? A climate-based model of the timing of peak pasture infectivity for Dictyocaulus viviparus. Vet. Parasitol., 309:109770.
- Mohamed, R.A., Abdelsalam, E.B. 2008. A review on pneumonic pasteurellosis (respiratory mannheimiosis) with emphasis on pathogenesis, virulence mechanisms and predisposing factors. Bulg. J. Vet. Med., 11(3):139-160.
- Momin, M.A., Islam, M.A., Khatun, M.M., Rahman, M.M. 2011. Characterization of bacteria associated with pneumonia in Black Bengal goats. Bangladesh J. Vet. Med., 9(1):67-71.

- Munsi, M.N., Ershaduzzaman, M., Akther, S., Rahman, M.M., Rahman, M.H., Rahman, M.M., 2018. Incidence of clinical diseases and disorders in goats at Bangladesh Livestock Research Institute. Asian J. Med. Biol. Res., 4(4):351-361.
- Nicolas, J., Ayling, R., McAuliffe, L. 2008. Mycoplasma Diseases of Ruminants. CAB International (ISBN 978-0-85199-012-5).
- Nooruzzaman, M., Akter, M.N., Begum, J.A., Begum, S., Parvin, R., Giasuddin, M., Islam, M.R., Lamien, C.E., Cattoli, G., Dundon, W.G., Chowdhury, E.H. 2021. Molecular insights into peste des petits ruminants virus identified in Bangladesh between 2008 and 2020. Infect. Genet. Evol., 96:105163.
- Nord, K., Ådnøy, T. 1997. Effects of infection by caprine arthritis-encephalitis virus on milk production of goats. J. Dairy Sci., 80(10):2391-2397.
- Norouzi, B., Razavizadeh, A.T., Azizzadeh, M., Mayameei, A., Mashhadi, V.N.N. 2015. Serological study of small ruminant lentiviruses in sheep Khorasan-e-Razavi population of province in Iran. In Vet Res Forum. (Vol. 6, No. 3, p. 245). Faculty of Veterinary Medicine. Urmia University, Urmia, Iran.
- Nyi Lin, T., Ngarmkum, S., Oraveerakul, K., Virakul, P., Techakumphu, M. 2011. Seroprevalence and risk factors associated with caprine arthritis-encephalitis virus infection in goats in the western part of Thailand. Thai J. Vet. Med., 353-360.

- Olech, M., Osiński, Z., Kuźmak, J. 2020. Seroprevalence of small ruminant lentivirus (SRLV) infection in wild cervids in Poland. Prev. Vet. Med., 176:104905.
- Ozdemir, U., Loria, G.R., Godinho, K.S., Samson, R., Rowan, T.G., Churchward, C., Ayling, R.D., Nicholas, R.A.J. 2006. Effect of danofloxacin (Advocin A180) on goats affected with contagious caprine pleuropneumonia. Trop. Anim. Health Prod., 38:533-540.
- Peixoto, de C.T., de Lima, E.B., Farias, S.S., Ferreira, M.M., Macêdo, A.G., Nakazato, L., Pescador, C.A., d'Avila, M.S.M.S., Carvalho, V.A.N. and Madureira, K.M., 2017. Surtos de conidiobolomicose ovina por Conidiobolus lamprauges no Estado da Bahia, Nordeste do Brasil. Braz. J. Vet. Med., 39(4):252-263.
- Qin, S., Yang, H., Zhang, Y., Li, Z., Lin, J., Gao, L., Liao, D., Cao, Y., Ren, P., Li, H., Wu, J. 2018. Full genome sequence of the first bluetongue virus serotype 21 (BTV-21) isolated from China: evidence for genetic reassortment between BTV-21 and bluetongue virus serotype 16 (BTV-16). Virol., Arch. 163:1379-1382.
- Quinn, P.J., Markey, B.K., Carter, M.E., Donnelly, W.J.C., Leonard, F.C. 2002. Veterinary microbiology and microbial disease. Blackwell science.
- Rahman, A.A., Islam, S.S., Sufian, M.A., Talukder, M.H., Ward, M.P., Martínez-López, B., 2020. Peste des Petits Ruminants risk factors and space-time clusters in Bangladesh.

- Front. Vet. Sci., 7:572432.
- Rahman, H., Bhuiyan, A., Parvin, R., Giasuddin, M., Haque, M., Sayem, S., Islam, M., Chowdhury, E. 2011. Immune response of goats to thermostable PPR vaccine in Bangladesh. SAARC J. Agri., 9:73–81.
- Rahman, M.H., Akther, S., Alam, M.S., Ali, M.Z. and Ahmed, S. 2023. Caprine arthritis and encephalitis virus infection in goats of Bangladesh: Serological detection and its associated risk factors. Vet. World, 16(11):2256-2262.
- Rahman, M.H., Akther, S., Alam, S., Hassan, Z., Sarker, S., Ali, Z., Ahmed, S. 2023. Prevalence and identification of caprine pasteurellosis in pneumonic goats in Bangladesh. J. Adv. Vet. Anim. Res., 10(3):538-544.
- Rahman, M.H., Akther, S., Ali, M.Z., Hassan, M.Z. 2020. Incidence of diseases in goats in Bangladesh. Bangladesh Vet., 37:14-20.
- Rahman, M.Z., Haider, N., Gurley, E.S., Ahmed, S., Osmani, M.G., Hossain, M.B., Islam, A., Khan, S.A., Hossain, M.E., Epstein, J.H., Zeidner, N. 2018. Epidemiology and genetic characterization of Peste des petits ruminant virus in Bangladesh. Vet. Med. Sci., 4(3): 161-171.
- Rakib, M.R., Ahmed, S., Desha, N.H., Akthe,r S., Rahman, M.H., Pasha, M.M., Dhakal, A., Sultana, N., Hemayet, M.A. 2022. Morphometric features and performances of Black Bengal goat in Bangladesh. Trop. Anim. Health Prod., 54(6):341.

- Rashid, M.M., Ferdoush, M.J., Dipti, M., Roy, P., Rahman, M.M., Hossain, M.I., Hossain, M.M. 2013. Bacteriological and pathological investigation of goat lungs in Mymensingh and determination of antibiotic sensitivity. Bangladesh J. Vet. Med., 11(2).
- Rushton, J., Lyons, N. 2015. A review of the effects on production. Vet. Ital., 51: 401-406.
- Samad, M.A., Yousuf, M.A., Siddiky, M.N.A., Giasuddin, M., Sarker, N.R., Samanata, A. K., Bokhtiar, S.M. 2019. Training manual on molecular diagnosis and laboratory surveillance of PPR. In: Bangladesh Livestock Research Institute, Savar, Dhaka, 1341, p. 84.
- Scott, P.R. 2011. Treatment and control of respiratory disease in sheep. Vet. Clin. North Am. Food Anim., 27(1):175-186.
- Sen, S.K., Chowdhury, M.R., Mahbub-E-Elahi, A.T.M., Siddique, A.B. 2018. Bacteriological and histopathological investigation of pneumonia in black Bengal goat. Dairy Vet. Sci. J., 6(4): 001-007.
- Solangi, G.M., Nizamani, Z.A., Tariq, M., Leghari, Z.A., Kamboh, A.A., Talpur, B.R. 2023. Seroprevalence of contagious caprine pleuropneumonia in goats from selected endemic areas of Sindh. J. Anim. Health Prod., 11(1):56-61.
- Souza, T.S.D., Pinheiro, R.R., Costa, J.N., de Lima, C.C., Andrioli, A., de Azevedo, D.A., dos Santos, V.W., Araújo, J.F., Sousa, A.L.M.D., Pinheiro, D.N., Fernandes, F. 2015.

- Interspecific transmission of small ruminant lentiviruses from goats to sheep. Braz. J. Microbiol., 46:867-874.
- Teshome, D., Sori, T., Sacchini, F., Wieland, B. 2019. Epidemiological investigations of contagious caprine pleuropneumonia in selected districts of Borana zone, Southern Oromia, Ethiopia. Trop. Anim. Health Prod., 51:703-711.
- Thomann, B., Falzon, L.C., Bertoni, G., Vogt, H.R., Schüpbach-Regula, G., Magouras, I. 2017. A census to determine the prevalence and risk factors for caprine arthritis-encephalitis virus and visna/maedi virus in the Swiss goat population. Prev. Vet. Med., 137: 52-58.
- Torres-Acosta, J.F.J., Gutierrez-Ruiz, E.J., Butler, V., Schmidt, A., Evans, J., Babington, J., Bearman, K., Fordham, T., Brownlie, T., Schroer, S., Cámara-G, E. 2003. Serological survey of caprine arthritis-encephalitis virus in 83 goat herds of Yucatan, Mexico. Small Rumin. Res., 49(2): 207-211.
- Wazir, I., Hussain, I., Khan, M.A., Ali, M.I., Rahman, H.U., Ashraf, F., Khan, S., Khan, B., Ullah, S., Ullah, Q. 2016. Seroepidemiological analysis of contagious caprine pleuropneumonia through cELISA in selected districts of Khyber Pakhtunkhwa-Pakistan. Am. Sci. Res. J. Eng. Technol. Sci. (Asrjets), 26:274-281.
- Wesonga, H.O., Bölske, G., Thiaucourt, F., Wanjohi, C., Lindberg, R. 2004. Experimental contagious caprine

- pleuropneumonia: a long term study on the course of infection and pathology in a flock of goats infected with Mycoplasma capricolum subsp. capripneumoniae. Acta Vet. Scand., 45: 1-13.
- WOAH (World Organization for Animal Health). 2004. Bovine brucellosis. Manual of Diagnostic Tests and Vaccines for Terrestrial Animals, OIE, World Organisation for Animal Health, (Part 2), p.3.
- WOAH (World Organization for Animal Health). 2016. Peste des Petits Ruminants Global Eradication Programme. In Contributing to Food Security, Poverty Alleviation and Resilience; Food and Agriculture Organization of the United Nations: Rome, Italy; World Organisation for Animal Health: Paris, France, 2016.
- WOAH (World Organization for Animal Health). 2020. OIE-Listed diseases, infections and infestations in force in 2020.
- WOAH (World Organization for Animal Health). 2021. Terrestrial Manual. Bluetongue (infection with bluetongue virus). Chapter 3.1.3.
- Yatoo, M.I., Parray, O.R., Bashir, S.T., Bhat, R.A., Gopalakrishnan, A., Karthik, K., Dhama, K. and Singh, S.V., 2019. Contagious caprine pleuropneumonia—a comprehensive review. Vet. Q., 39(1):1.
- Zafari, S., Mohtasebi, S., Sazmand, A., Bahari, A., Sargison, N.D., Verocai, G.G. 2022. The prevalence and control of lungworms of pastoral ruminants in Iran. Pathogens, 11(12):1392.