

## Finalized Article

# Nasal colonization of Methicillin resistant *Staphylococcus aureus* among patients during hospital admission--emergence of community-associated MRSA strains.

Shahana Khanam<sup>1</sup>, Mohammad Jobayer<sup>2</sup>, SM Shamsuzzaman<sup>2</sup>, Jalaluddin Ashraful Haq<sup>3</sup>,  
Md Motlabur Rahman<sup>4</sup>, Kazi Zulfiquer Mamun<sup>5</sup>.

<sup>1</sup>Department of Microbiology, MH Samorita Medical College, Dhaka. <sup>2</sup>Department of Microbiology, Dhaka Medical College.

<sup>3</sup>Department of Microbiology, Ibrahim Medical College, Dhaka. <sup>4</sup>Department of Medicine, Dhaka Medical College.

<sup>5</sup>Department of Microbiology, Popular Medical College, Dhaka

### Abstract

Patients colonized with Methicillin resistant *Staphylococcus aureus* (MRSA) in hospital are considered as one of the risk factors for infection with MRSA. Worldwide spread of MRSA in both hospital setting and community poses public health threat. This study was undertaken to determine the frequency of MRSA colonization among patients at time of hospital admission. Five hundred adult patients were screened within 24 hrs of admission in different wards in Dhaka Medical College Hospital by taking nasal swabs from anterior nares and were analyzed. All isolated *Staphylococcus aureus* were screened to detect methicillin resistance by modified Kirby-Bauer disc diffusion method using oxacillin and ceftiofloxacin disc and then all MRSA isolates were subjected for MIC testing against oxacillin by agar dilution method and PCR for *mecA* gene detection. Out of 500 patients *Staph aureus* nasal colonization was observed among 112 (22.4%) patients and among those 7.6% was MRSA. MRSA colonization rate was 23.29% among patients who had history of prior hospitalization and was 4.92% among community residents who had no previous hospitalization history in last 12 month. A significant number of patients (7.6%) were colonized with MRSA at the time of admission. Screening for MRSA carriers among this population is necessary for hospital acquired infection control.

**Key words:** MRSA, *Staphylococcus aureus*, PCR

### Introduction:

Colonization with *Staphylococcus aureus* has been identified as an important risk factor for the development of *Staph aureus* infections in both community and hospital settings<sup>1-3</sup>. Anterior nares are the most consistent site of *Staph aureus* colonization<sup>4</sup>. *Staph aureus* first developed resistance to penicillin in the 1940s and then to methicillin in early 1960s. Methicillin resistant *Staph aureus* (MRSA) is resistant to methicillin and other  $\beta$ -lactamase-resistant penicillins (oxacillin, nafcillin) and cephalosporins<sup>5</sup>. Number of MRSA infections has doubled in the last 10 years, and number of

deaths in the United States owing to complications of this infection is higher than the number of deaths from AIDS<sup>6</sup>. MRSA infection is largely confined to hospitals and long term care facilities, typically linked to persons with health care associated risk factors such as hospitalization or nursing home care, chronic dialysis, antibiotic treatment, or exposure to invasive devices or procedures, called healthcare-associated MRSA (HA-MRSA) infection<sup>7</sup>. The frequency of community-associated MRSA (CA-MRSA)<sup>8</sup> is increasing. CA-MRSA is an emerging pathogen diagnosed from an outpatient or within 48hrs of hospitalization if the patient lacks healthcare-associated MRSA risk factors<sup>7</sup>. These infections have been associated with carriage of *Staphylococcus aureus* cassette chromosome (SCC) *mec* type IV complex and genes encoding Panton-Valentine leukocidin toxin<sup>9,10</sup>. Different studies<sup>11,12</sup> reported that PVL genes were differentially distributed among CA-MRSA strains and PVL is not only the key virulence determinant of CA-MRSA.

---

#### ✉ Corresponding author:

- Dr. Shahana Khanam
- Assistant Professor
- Department of Microbiology, MH Samorita Medical College
- Dhaka, Bangladesh. Tel: +8802-9143757
- Mobile: 01717546296
- Email: shahana77\_dr@yahoo.com

Other virulence factors are associated with CA-MRSA, such as phenol-soluble modulins (PSMs) and  $\alpha$ -hemolysin<sup>13,14</sup>. In Bangladesh, the rate of MRSA infection ranges from 32% to 63% in hospitals<sup>15</sup>. The frequency of MRSA is alarming here due to indiscriminate and incomplete uses of antibiotics<sup>16,17</sup>. Recognition and isolation of persons either colonized or infected with MRSA is recommended for minimizing the spread of MRSA within hospitals. In Bangladesh, there is no adequate information on MRSA nasal colonization that is the important risk factor for both HA-MRSA and CA-MRSA infection. The present study was designed to determine the MRSA nasal colonization in patients at the time of admission to the hospital and to evaluate CA-MRSA carriage.

**Methods:**

This cross sectional study was carried out in the Department of Microbiology in Dhaka Medical College during the period of January 2010 to December 2011.

Five hundred adult patients were screened within 24 hours of their admission to Dhaka Medical College Hospital by taking nasal swab from both anterior nares and were analyzed. Data related to age, sex, history of prior hospitalization (within past 12 month) or directly from home and their medical history, such as-diabetes mellitus, chronic obstructive pulmonary disease, cerebrovascular disease, chronic kidney disease were collected from hospital records or directly from patients using predesigned data collection form.

**Case definition:**

**Community acquired MRSA:**

MRSA strains isolated from the patients having the following criteria were considered as community acquired MRSA (CA-MRSA).

- Samples collected within 24 hours after admission to the hospital.
- No history of hospitalization in the past year (within 12 month).
- No indwelling catheters or medical devices that pass through skin into the body.

**Healthcare associated MRSA:**

MRSA strains isolated from the patients having history of hospitalization within last 12 months were considered as healthcare associated MRSA (HA-MRSA).

**Collection of nasal swab:** A single sterile cotton swab was moistened with sterile normal saline and was then inserted into each nostril and nasal septum and immediately processed for culture. Nasal swab samples were plated on blood agar media and incubated at 37°C. Isolates were identified as *Staph aureus* by colony morphology, Gram staining and

biochemical tests (catalase, coagulase and mannitol fermentation test)<sup>18</sup>.

**Detection of MRSA:** *Staph aureus* isolates were screened for methicillin resistance by disc diffusion method using oxacillin (1µg) and cefoxitin (30µg) disc and by determination of minimum inhibitory concentration (MIC) of oxacillin by agar dilution method as per recommendation of CLSI method<sup>19</sup> and by detection of *mec-A* gene by PCR.

**Polymerase chain reaction (PCR):** PCR for detection of *mec-A* and PVL genes was performed using specific primers. DNA was extracted from bacterial pellets by simple boiling method<sup>20</sup>.

**DNA amplification:** Isolated DNA was amplified by using specific primers for *mec-A*<sup>21,22</sup>. The following oligonucleotide primers were used:

Primer	Oligonucleotide sequence (5'-3')	Amplicon Size
<i>mec-A</i>	Forward- AAAATCGATGGTAAAGGTTGGC	533 bp
	Reverse- AGTTCTGCAGTACCGGATTTTGC	

PCR was performed in a final reaction volume of 25µl, containing 12.5 µl Master mix, 1.5µl of each primer, 2µl of extracted DNA and 6.5µl nuclease free water (Promega Corporation, USA).

**Visualization and Interpretation of results:** After staining with ethidium bromide (0.5µg/ml) and destaining, gel was observed under UV Transilluminator (Gel Doc, Major science, Taiwan) and DNA bands were identified according to their molecular size by comparing with 100 bp DNA ladder. Samples showing the presence of specific DNA band corresponding to 533 bp were considered positive for presence of *mec-A* gene.

**Antimicrobial susceptibility test:** All MRSA isolates were tested for susceptibility against ceftriaxone (30µg), ciprofloxacin (5µg), doxycycline (30µg), erythromycin (15µg), gentamycin (10µg), rifampicin (5µg), vancomycin (30µg), fusidic acid (10µg) and linezolid (30µg) by disc diffusion method as recommended by CLSI<sup>19</sup>. The discs from each batch were standardized by testing against reference strain of *Staph aureus* ATCC-25923.

**RESULTS:**

After screening 500 nasal swabs, 255 (51%) were culture positive for *Staphylococcus*. Out of 255 *Staphylococcus*, 112 (22.4%) were *Staph aureus* and 143 (28.6%) were coagulase negative *Staphylococcus* (Table I).

Out of 112 *Staph aureus*, 38 (33.93%) strains were detected as MRSA and 74 (66.07%) strains were detected as MSSA by different phenotypic method and by detection of *mec-A* gene by PCR (Table II).

Among 73 patients having previous history of hospitalization, 23 (31.50%) *Staph aureus* were isolated, of them 17 (23.29%) were MRSA. Of the 427 patients who had no history of previous hospitalization, 89 (20.84%) were *Staph aureus*, of them 21 (4.92%) were MRSA (Table III).

Both health-care related and community-associated-MRSA strains were resistant to anti-staphylococcal β-lactam antibiotics (oxacillin, cefoxitin, ceftriaxone). Both CA-MRSA and HA-MRSA strains were highly resistant to ciprofloxacin (90.47% and 94.11% respectively). HA-MRSA colonization strains showed resistance to erythromycin (88.23%), gentamycin (82.35%) and doxycycline (70.59%). Rate of resistance to both vancomycin and rifampicin were 17.65% and fusidic acid was 23.53% among health-care related MRSA isolates. Most (76.19%) of CA-MRSA strains were resistant to erythromycin. Most of CA-MRSA isolates were susceptible to rifampicin (95.24%), fusidic acid (90.48%) and doxycycline (47.62%). All CA-MRSA strains were susceptible to vancomycin and linezolid (Table IV).

**Table I : Isolation rate of Staphylococcus from nasal swab sample (n=500)**

Staphylococci	No. (%)
Staphylococcus aureus	112 (22.40)
Coagulase -ve Staphylococcus	143 (28.60)
<b>Total</b>	<b>255 (51.00)</b>

**Table II : Shows isolation rate of MRSA and MSSA among Staphylococcus aureus (n=112).**

<i>Staphylococcus aureus</i>	No. (%)
MRSA	38 (33.93)
MSSA	74 (66.07)
<b>Total</b>	<b>112 (100)</b>

**Table III : Isolation of Staph aureus and MRSA among previously hospitalized patients and patients from community (n=500).**

Study population	<i>Staph aureus</i>	<i>MRSA</i>
	No. (%)	No. (%)
Previously hospitalized patients (n=73)	23 (31.50)	17 (23.29)
Patients from community (n=427)	89 (20.84)	21 (4.92)
<b>Total (n= 500)</b>	<b>112 (22.40)</b>	<b>38 (7.60)</b>

**Table IV: Antimicrobial susceptibility pattern of healthcare associated-MRSA and community associated-MRSA strains.**

Antimicrobial agents	HA (n=17) Resistant No. (%)	Sensitive No. (%)	CA (n=21) Resistant No. (%)	Sensitive No. (%)
Ceftriaxone	16 (94.12)	01 (5.88)	19 (90.47)	02 (9.53)
Ciprofloxacin	16 (94.12)	01 (5.88)	19 (90.47)	02 (9.53)
Erythromycin	15 (88.24)	02 (9.53)	16 (94.12)	05 (23.81)
Gentamycin	14 (82.35)	03 (17.65)	15 (88.24)	06 (28.57)
Doxycycline	12 (70.59)	05 (23.81)	11 (50.38)	10 (47.62)
Vancomycin	03 (17.65)	14 (82.35)	00 (0.00)	21 (100)
Rifampicin	03 (17.65)	14 (82.35)	01 (5.88)	20 (95.24)
Fusidic acid	04 (23.53)	13 (76.47)	02 (9.53)	19 (90.47)
Linezolid	01 (5.88)	16 (94.12)	00 (0.00)	21 (100)

**Discussion:**

Methicillin-resistant *Staph aureus* (MRSA) is not only confined to healthcare facilities or healthcare associated, but also a significant number of persons carry this organism without having any history of hospitalization or any risk factor, called community-associated MRSA (CA-MRSA)<sup>23</sup>. MRSA is a serious threat to hospitalized patients globally and now represents a challenge for public health, as community-acquired infections appear to be on the increase in various regions and countries<sup>24, 25</sup>. Nasal colonization is important risk factor for both hospital and community acquired MRSA infection<sup>1</sup>. It is necessary to take steps to prevent the spread of MRSA infection in hospital and community.

This study revealed that 38 (7.6%) patients were colonized with MRSA at the time of hospital admission which was similar to the study of from the USA<sup>26</sup> and Santos *et al* (2010)<sup>27</sup> from Brazil where colonization rate was 7.3% and 6.1% respectively. The prevalence of MRSA at admission was 3.4% and 1.1% in patients from the USA and from Saudi Arabia<sup>28, 29</sup> respectively which is lower than this study. Very low prevalence (0.03%) of MRSA nasal carriage at the time of hospitalization was observed in Netherland<sup>30</sup>. Such lower isolation rate in the studies of different countries was probably due to the fact that in those countries MRSA control program is well established and irrational antibiotic prescribing is restricted. This higher rate of MRSA colonization in this study is probably due to lack of MRSA control program, poor knowledge about personal hygiene among general population and overcrowding environment.

This study also evaluated that MRSA colonization rate was 23.29% among patients who had previous history of hospitalization in contrast to patients who had no previous hospitalization history (4.92%). Chatterjee *et al* (2009)<sup>31</sup> from India, Santos *et al* (2010)<sup>27</sup> from Brazil reported similar

results who also found significant relationship between MRSA colonization and hospitalization. The reason of higher rate of MRSA colonization in patients with history of previous hospitalization may be explained by the fact that, health care system including hospital personnel (patients and health care workers) act as important reservoir of MRSA acquisition that may be transmitted to other patients.

In this study, 427 patients were admitted directly from community who had no history of hospitalization in last 12 months. In this community group of patients isolation rate of MRSA (CA-MRSA) was 4.92%. This result was similar to the study of Chatterjee *et al.*(2009)<sup>31</sup> in India where CA-MRSA carriage rate was 3.16%. A study by Hidron *et al.*(2005)<sup>26</sup> from the USA reported that 2.2% of patients colonized with MRSA were admitted directly from community, which was lower than this study. Lower rate of MRSA colonization in the community people may be explained by the fact that they are less exposed to the source of MRSA as there is less chance of contact with health-care system and less exposure of antibiotics.

Out of 38 MRSA-colonization strains, 21 (55.26%) and 17 (44.74%) were community and health-care associated MRSA strains, respectively. In this study both CA-MRSA and HA-MRSA strains were resistant to anti-staphylococcal  $\beta$ -lactam antibiotics (oxacillin, cefoxitin, ceftriaxone). Both CA-MRSA and HA-MRSA strains were highly resistant to ciprofloxacin (90.47% and 94.12% respectively). CA-MRSA strains were resistant to erythromycin (76.17%) and gentamycin (76.49%) which was in agreement with study of Neela *et al.*(2008)<sup>32</sup> from Malaysia and Kim *et al.*(2004)<sup>33</sup>. All CA-MRSA strains were susceptible to vancomycin and linezolid. Most of CA-MRSA isolates were susceptible to rifampicin (95.24%), fusidic acid (90.48%) and doxycycline (47.62%). On the other hand, health-care related MRSA colonization strains showed resistance to erythromycin (88.24%), gentamycin (82.35%) and doxycycline (70.59%). Rate of resistance to rifampicin and fusidic acid were 17.65% and 23.53% respectively among health-care related MRSA isolates, which were higher than resistance rate of 4.76% and 9.52% among community associated MRSA isolates.

Antimicrobial susceptibility by disc diffusion method showed that 3 (17.65%) isolates were resistant to vancomycin. CA-MRSA isolates tend to become susceptible to non- $\beta$ -lactam antibiotics than HA-MRSA<sup>34</sup>. Chura *et al.*(2011)<sup>35</sup> reported that there was significant diversity in MRSA clones arising in the community worldwide, because geographical differences in typical antimicrobial resistance profiles. A study in Bangladesh reported that widespread and suboptimal use of

antimicrobial agents was an important factor for high prevalence of resistant strains<sup>17</sup>.

#### Conclusion:

This study demonstrated that a significant number of MRSA (7.6%) carrier patients are seeking admission everyday in Dhaka Medical College Hospital. MRSA colonization rate was 4.92% among community residents who had no hospitalization history. Carrier patients can transmit MRSA to other inpatients in hospital by skin-to-skin contact or by contact with contaminated items. So, early detection of MRSA carrier, contact isolation and decolonization may prevent MRSA transmission in hospital and community. MRSA colonization rate was higher among patients who had history of previous (within 12 month) hospitalization. So, maintaining clean environment of hospital and hygiene practices among hospital personnel and patient attendant during handling the patients may prevent MRSA transmission from hospital to hospital or to community.

#### Financial support:

The work was supported financially by the department of Microbiology, Dhaka Medical College, Dhaka, Bangladesh.

#### Acknowledgement:

We acknowledge different wards of Dhaka Medical College, Dhaka, Bangladesh for providing the sample collection facility.

#### Conflict of interest:

We do not have any potential conflicts of interest.

#### REFERENCES:

1. Kluytmans J, Van Belkum A, Verbrugh H. Nasal carriage of *Staph aureus*: epidemiology, underlying mechanisms, and associated risks. *Clin Microbiol Rev* 1997; 10: 505-20.
2. Gorwitz RJ, Kruszon-Moran D, McAllister SK, McQuillan G, McDougal LK, Fosheim GE. Changes in the prevalence of nasal colonization with *Staph aureus* in the United States. *J Infect Dis* 2008; 197: 1226-34.
3. Von-Eiff C, Becker K, Machka K. Nasal carriage as a source of *Staph aureus* bacteremia. *N Engl J Med* 2001; 344: 11-6.
4. Sanford MD, Widmer AF, Bale MJ, Jones RN, Wenzel RP. Efficient detection and long term persistence of the carriage of methicillin-resistant *Staph aureus*. *Clin Infect Dis* 1994; 19: 1123-8.
5. McDougal LK and Thornsberry C. The role of  $\beta$ -lactamase in Staphylococcal resistance to penicillinase-

- resistant penicillins and cephalosporins. *J Clin Microbiol* 1986; 23:832-9.
6. Como-Sabeti KC, Harimon KH, Buck JM, Glennen A, Boxrude DJ, Lancefield R. Community-associated methicillin-resistant *Staph aureus*: trends in case and isolate characteristics from six years of prospective surveillance. *Public Health Rep* 2009; 124: 427-35.
  7. Klevens RM, Morrison MA, Nadle J, Petit S, Gershman K, Ray S. Invasive methicillin-resistant *Staph aureus* infections in the United States. *JAMA* 2007; 298: 1763-71.
  8. Chambers HF. The Changing Epidemiology of *Staph aureus*? *Emerg Infect Dis* 2001; 7: 172-82.
  9. Boyle-Vavra S and Daum SR. Community-acquired methicillin resistant *Staph aureus*: the role of Panton-Valentine leukocidin. *Lab Invest* 2007; 87: 3-9.
  10. Deleo FR, Otto M, Kreiswirth BN, Chambers HF. Community-associated methicillin-resistant *Staph aureus*. *Lancet* 2010; 375: 1557-68.
  11. Said-Salim B, Mathema B, Kreiswirth BN. Community-acquired methicillin-resistant *Staph aureus*: an emerging pathogen. *Infect Control Hosp Epidemiol* 2003;24: 451-5.
  12. Voyich JM, Otto M, Mathema B, Braughton KR, Whitney AR, Welty D, et al. Is Panton-Valentine leukocidin the major virulence determinant in community associated methicillin-resistant *Staph aureus* disease? *J Infect Dis* 2006; 194: 1761-70.
  13. Bubeck WJ, Bae MT, Otto F, Deleo R, Schneewind O. Poring over pores: alpha hemolysin and Panton-Valentine leukocidin in *Staph aureus* pneumonia. *Nat Med* 2007; 13:1405-6.
  14. Wang FD, Chen YY, Chen TL, Liu CY. Risk factors and mortality in patients with nosocomial *Staph aureus* bacteremia. *Am J Infect Control* 2008; 36: 118-22.
  15. Haq JA, Rahman MM, Asna SM, Hossain MA, Ahmed I, Haq T, *et al*. Methicillin-resistant *Staph aureus* in Bangladesh: a multicentre study. *Int J Antimicrobiol Agents* 2005; 25: 276-7.
  16. Khan HA, Shamsuzzaman AKM, Paul SK, Alam MM, Mahmud MC, Musa AKM, *et al*. Antimicrobial susceptibility and coagulase typing of MRSA strains at Mymensingh Medical College. *Bangladesh J Microbiol* 2007; 1: 56-60.
  17. Mamun KZ, Shears P, Tabassum S, Hart CA. Antimicrobial use and antimicrobial resistance in rural Bangladesh. *Trans Roy Soc Trop Med Hygn* 1996; 90: 213.
  18. Cheesbrough M. Microscopical techniques used in microbiology. In: *District laboratory practices in tropical countries, Part II*. Cambridge University Press; UK: 2000; pp 39-41.
  19. Clinical and Laboratory Standards Institute. Performance standards for antimicrobial susceptibility testing; Seventeenth informational Supplement. Document M100-S17. Wayne, PA. CLSI 2007; 27.
  20. Reischl U, Pulz M, Ehret W, Wolf H. PCR based detection of mycobacteria in sputum samples using a simple and reliable DNA extraction protocol. *Biotechniques*. 1994; 17: 844-45.
  21. Rahimi F, Bouzari M, Maleki Z, Rahimi F. Antibiotic susceptibility pattern among *Staphylococcus* spp. With emphasis on detection of *mecA* gene in methicillin resistant *Staph aureus* isolates. *Iranian J Clin Infect Dis* 2009; 4:143-150.
  22. Siripornmongkolchai T, Chomvarin C, Chaicumpar K. Evaluation of different primers for detecting *Mec -A* gene by PCR in comparison with phenotypic methods for discrimination of Methicillin-resistant *Staph aureus*. *Southeast Asian J Trop Med Public Health* 2002 ;33 :758-63
  23. Centers for Disease Control and Prevention. Active Bacterial Core Surveillance: methodology-case definition and ascertainment. 2007. Available at <http://www.cdc.gov/ncidod/dbmd/abcs/meth-case.htm>. Accessed on 21/06/ 2010.
  24. Vandenesch F, Naimi T, Enright MC, Lina G, Nimmo GR, Heffernan H, Liassine N. Community-acquired methicillin-resistant *Staph aureus* carrying Panton-Valentine leukocidin genes: worldwide emergence. *Emerg Infect Dis* 2003; 9: 978-84.
  25. Layton M C, Hierholzer WJ, Patterson JE. The evolving epidemiology of methicillin resistant *Staph aureus* at a university hospital. *Infect Control Hosp Epidemiol* 1995; 16: 12-7.
  26. Hidron AI, Kourbatova EV, Halvosa J, Terrell BJ, Mc Dougal LK, Tenover FC, *et al*. Risk factors for colonization with methicillin-resistant *Staph aureus* in patients admitted to an urban hospital: emergence of community-associated MRSA nasal carriage. *Clin Infect Dis* 2005; 41: 159-66.
  27. Santos HB, Machado DP, Camey SA, Kuchenbecker RS, Barth AL, Wagner MB. Prevalence and acquisition of MRSA amongst patients admitted to a tertiary-care hospital in Brazil. *BMC Infect Dis* 2010; 10: 2-7.

28. Davis KA, Stewart JJ, Crouch HK, Florez CE, Hospenthal DR. Methicillin-resistant *Staph aureus* (MRSA) nares colonization at hospital admission and its effect on subsequent MRSA infection. *Clin Infect Dis* 2004; 39: 776-82.
29. Panhotra BR, Saxena AK, Abdulrahman S, Mulhim A. Prevalence of methicillin-resistant and methicillin sensitive *Staph aureus* nasal colonization among patients at the time of admission to the hospital. *Ann Saudi Med* 2005; 25: 304-8.
30. Wertheim HF, Vos MC, Boelens HA, Voss A, Vandembroucke-Grauls CM, Meester MH, *et al*. Low prevalence of methicillin-resistant *Staph aureus* (MRSA) at hospital admission in the Netherlands: the value of search and destroy and restrictive antibiotic use. *J Hosp Infect* 2004; 56: 321-5.
31. Chatterjee SS, Ray P, Aggarwal A, Das A, Sharma M. A community-based study on nasal carriage of *Staph aureus*. *Ind J Med Res* 2009; 130: 742-8.
32. Neela V, Sasikumar M, Ghaznavi GR, Zamberi S, Mariana S. In vitro activities of 28 antimicrobial agents against methicillin-resistant *Staph aureus* from a clinical setting in Malaysia. *Faculty Med Health Sci* 2008; 39: 885-92.
33. Kim HB, Jang H, Nam HJ. In vitro activities of 28 antimicrobial agents against *Staph aureus* isolates from tertiary-care hospitals in Korea: a national wide survey. *Antimicrobiol Agents Chemother* 2004; 48: 1124-7.
34. Helen C, Maltezou A, Helen G. Community acquired methicillin-resistant *Staph aureus* infections. *Int J Antimicrobiol Agents* 2006; 27: 87-96.
35. Chura K, Laurent F, Coombs G, Grayson ML, Howden BP. Not Community-associated methicillin-resistant *Staph aureus* (CA-MRSA)! A clinician's guide to community MRSA- its evolving antimicrobial resistance and implications for therapy. *Clin Infect Dis* 2011; 52: 99-114.