

Original Articles

Serological and Molecular Epidemiology of Human Brucellosis in Mymensingh region Bangladesh

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Abstract

Brucellosis is a major Bacterial Zoonotic disease worldwide. More than 500,000 new cases are reported globally every year with the annual incidence rates varying widely from <2 to >500 per 1000,000 population among different region of the world. This cross sectional descriptive study was carried out in the department of microbiology, Mymensingh medical college, Mymensingh, Bangladesh from 1st January to 31 December 2017, on total 600 patients of PUO with or without occupational exposure. Blood was taken as sample, serum was separated and screening test was done by Brucella specific latex agglutination test from all collected samples. A Titer of $\geq 1:160$ was taken as positive. ICT, PCR and real time PCR were done from all screening positive samples. Among the study population 65.83% (395/600) were male and 34.17% (205/600) were female. The sero prevalence of Human Brucellosis was found 13.33% (40/300) and 5% (15/300), respectively in risk and non risk group of population. Out of 55 Brucella specific latex agglutination test positive samples 27.27% (15/55) were ICT positive, 5.45% (3/55) were PCR positive and 3.64% (2/55) were real time PCR positive. No ICT, PCR or Real time PCR cases were found from sero negative samples. Among the ICT positive cases 86.67% (13/15) from risk group and 13.33% (2/15) from non risk group of study population. Among the 3 PCR positive cases 13.33% (2/15) were from ICT positive cases and 2.5 % (1/40) from ICT negative cases. All PCR and Real time PCR positive cases were found in risk group of study population. No reliable data for Human Brucellosis is available in our country. However the present study revealed that a considerable number of Human Brucellosis is present in risk group as well as non risk group of population in both rural and urban area in Mymensingh district of Bangladesh. In our country we use only Brucella specific latex agglutination test for diagnosis of human Brucellosis. It is a non specific test and gives many false positive results. So it is necessary to introduce newer tests with higher sensitivity and specificity which will help to diagnose the disease rapidly and more accurately.

Introduction

Brucellosis is a worldwide reported zoonotic and occupational disease in farmers, veterinarians, slaughter house workers, animal handlers and meat inspectors¹. It is caused by bacteria belonging to genus *Brucella*. They are small gram-negative, intracellular, non spore forming, non capsulated cocco-bacilli². Brucellosis is a very old zoonotic disease and recent evidence showed that it was recognized as a clinical entity from the times of Crimean

war³. The responsible micro-organism was isolated by David Bruce in 1887 from the spleen of a soldier who died from the disease⁴. There are six known species of Brucellae including *B.melitensis*, *B.abortus*, *B.suis*, *B.canis*, *B.ovis*, *B.neotomae* are identified at present⁵. According to World Health Organization every year more than 500,000 new cases are reported with the annual incidence rates varying widely from <2 to >500 per 1000,000 population among different region of the world^{1,6}. The disease remains endemic in many regions of the world including the Eastern Mediterranean basin, The Middle East, The Arabian

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peninsula, Mexico, Central and South America, Central Asia, and the Indian subcontinent⁷. International travelers visiting *Brucella* endemic region are at risk of infection¹. There are sporadic reports of prevalence of Human Brucellosis in Bangladesh and the true incidence of Brucellosis is not known. Serological tests done among cattle handlers of some villages in Bangladesh showed that the prevalence is in between 6-12.8 %⁸.

The most important way for transmission of *Brucella* to human is consumption of unpasteurized milk and milk products, contact with meat and blood of infected livestock, skin cuts, breathing dust contaminated with feces and urine of the infected animal. Transmission of the disease from Human to Human has rarely been happening but it may be transmitted through sexual intercourse (sperm), blood transfusion, bone marrow transplantation, shared drug needles among the addicted, intrauterine transfusion through the placenta and breast milk^{9,10}.

Human Brucellosis is a debilitating illness characterized by fever (78%), arthralgia (65%), myalgia (47%), and back pain (45%). Hepatomegaly (23%), splenomegaly (26%)¹¹. Human Brucellosis is often misdiagnosed or under diagnosed due to overlapping clinical manifestations with many other bacterial infections. It is one of the causes of fever of prolonged duration in endemic areas and one of the important causes of pyrexia of unknown origin¹².

The laboratory diagnosis of Human Brucellosis is based on microbiological, serological, or molecular methods, each having its own advantages and disadvantages¹³. Many serological tests such as rose bengal plate test (RBPT), complement fixation test (CFT) and serum agglutination test (SAT), Coombs test, ELISA are used for the diagnosis of Human Brucellosis¹⁴ but isolation of pathogen from blood culture remains the gold standard¹⁵. The molecular diagnosis of Human Brucellosis can be performed using genus specific polymerase chain reaction (PCR)¹⁶, molecular assay targeting the *bcsp31* gene coding for a 31kDa immunogenic outer membrane protein conserved among all *Brucella* spp are the most common molecular targets in clinical applications¹³ in comparison to PCR. Real time PCR is a technique for determination and quantification of nucleic acid¹⁷.

Materials and methods

This cross sectional descriptive type of study was carried

out in the Department of Microbiology, Mymensingh medical college, Mymensingh, Bangladesh, On patients suffering from pyrexia of unknown origin who were attending at outpatient and inpatient department of Mymensingh medical college hospital from 01/01/2017 to 31/12/2017. Total 600 (six hundreds) patients of either sex (300 from risk group/occupationally exposed and 300 from non risk group) were included in this study. 10 ml Whole blood was collected aseptically from each patient of which 5 ml were stored in-20°C for PCR and Real time PCR and serum were separated from rest of blood and used for *Brucella* specific latex agglutination test and ICT.

Laboratory procedure

At first screening test (*Brucella* specific latex agglutination test) were done with all collected samples and Titer \geq 1:160 were consider as screening positive¹⁸. The screening positive samples were then tested for ICT, PCR and real time PCR. The *Brucella* specific latex agglutination test (Spinreact SA/SAU. ctra Santa calona. 7E.17176 SA NT ESTEVE 1E BAS (G1) Spain) was performed on each sample according to the manufacturer's instructions. *Brucella* IgM/IgG LFA is an immune chromatographic lateral flow assay (one diagnostics, 1090HA, Amsterdam, The Netherlands). The assay was intended to be used as an aid in the sero diagnosis of Brucellosis. The *Brucella* IgM/IgG LFA consists of two devices, one for the detection of specific IgM antibodies and one for the detection of specific IgG antibodies and the test were done according to the manufacturer's instructions.

Polymerase chain reaction (PCR)

Detection of *Brucella* genome specific gene encoding outer membrane protein by PCR from blood was performed by using standard protocol with specific primer¹. The primers and reagent were purchased from Sigma Aldrich co (St. Louis. MO, USA) The reaction mixture 25 μ l consisted of 12.5 μ l of ready mix taq buffer with MgCl₂, (Sigma Aldrich, co, St. Louis, MO, USA), 0.5 μ l forward primer (10 pmole/ μ l), 0.5 μ l reverse primer (10 pmole/ μ l), 10 μ l of nuclease free water, To this mixture 1.8 μ l of template DNA was added.

Protocol of thermal cycle

Initial Denaturation at 95°C for 3 minutes
 Denaturation at 95°C for 45 seconds
 Primer annealing at 60°C for 45second
 Extension at 72°C for 2 minutes and
 Final extension at 72°C for 10 minutes

} 35 cycles (1)

Primer used for PCR

The molecular diagnosis of Human Brucellosis can be performed using genus specific primers by polymerase chain reaction (PCR)¹⁶; molecular assay targeting the bcp31 gene coding for a 31-kDa immunogenic outer membrane protein conserved among all *Brucella* spp are the most common molecular targets in clinical applications¹³.

Brucella specific 31KDa antigen specific primers¹³

Prime name	Sequence of primer	Product size
B4	5'-TGGCTCGGTTGCCA-ATATCAA-3'	223 bp
B5	5'-CGCGCTTGCCTTTC-AGGTCTG-3'	

Visualization and documentation

The PCR products were run in 1.5% agarose gels stained with ethidium bromide and visualized under the UV transilluminator and photographs was taken by using digital camera.

Interpretation

Samples were score as positive when a PCR product of 223 bp in case of 31 KDa antigen genes could be detected in PCR.

Real time PCR

To detect *Brucella* spp by real time PCR from samples we used geneaid, genomic DNA purification kit for whole blood DNA isolation and purification¹⁹.

Programming the real time PCR instrument¹⁹

- 1) Select the filter pairs to be used (FAM and HEX).
- 2) Identification of unknown samples, positive and negative control.
- 3) Select the correct thermal protocols.

Protocol of thermo cycler¹⁹

- 1) Initial denaturation for activation the hotstar Taq DNA polymerase at 95°C for 12.30 minutes.
- 2) Denaturation at 97°C for 00.30 minutes.
- 3) Annealing and synthesis at 54°C for 1.30 minutes.
- 4) Terminal hold at 22°C for 5 minutes.

(Point 2 and 3 run for 50 cycles)

Result analysis: After the end of program, the graphics were seen. In negative control it was running along with the bottom and for positive control it gave a curve in the soft ware graphics.

Results

Table-1 Age and sex distribution of total study population (n=600).

Parenthesis indicates percentage

Table 1: Showing the total distribution of study population regarding age and sex. Among the study population 65.83% (395/600) were male and 34.17% (205/600) were female. Majority of the study population 49.17% (295/600) were in the age group of 20-40 years

Age distribution (years)	Male	Female	Total
<20	105 (68.63%)	48 (31.37%)	153 (25.5%)
20-40	180 (60.02%)	115 (38.98%)	295 (49.17%)
41-60	80 (76.19%)	25 (23.81%)	105 (17.5%)
>60	30 (63.83%)	17 (36.17%)	47 (7.83%)
Total	395 (65.83%)	205 (34.17%)	600 (100%)
Ratio	Male: Female =1.93 : 1		

Table 2: Result of *Brucella* specific latex agglutination test in risk and non risk group of study population

Titer	Risk group		cumulative percentage	Non risk group		cumulative percentage
	no of cases	percentage		no of cases	percentage	
1:20	115	38.33%	--	155	51.67%	--
1:40	80	26.67%	--	105	35%	--
1:80	65	21.67%	--	25	8.33%	--
1:160	36	12%	13.33%	15	5%	5%
1:320	04	1.33%			--	
Total	300			300		

Table 2: showing the result of *Brucella* specific latex agglutination test in both group of study population. In risk group 12% (36/300) and 1.33% (4/300) having a titer of 1:160 and 1:320 respectively, the cumulative percentage were 13.33% and in non risk group 5% (15/300) population has a titer of 1:160 .

Table 3: Distribution of ICT positive cases in latex agglutination test positive cases (n=55)

Group	ICT positive	ICT negative
Risk (n=40)	13 (32.50%)	27 (67.5%)
Non Risk (n=15)	02 (13.33%)	13 (86.67%)
Total (n=55)	15 (27.27%)	40 (72.73%)

Table 3: shows the distribution of ICT positive cases among the *Brucella* specific latex agglutination test positive cases. Total 15(27.27%) cases were ICT positive. Among them 13(32.5%) were from risk group and (13.33%) from non risk group of study population.

Table 4: Distribution of PCR positive cases among the *Brucella* specific latex agglutination test positive cases in both group of study population (n=55)

Group	PCR positive	PCR negative
Risk group (n=40)	03	37
Non risk group (n=15)	00	15
Total (n=55)	03 (5.45%)	52 (94.55%)

Table 4 : shows the distribution of PCR positive cases among the *Brucella* specific latex agglutination test positive cases. Total 5.45% (3/55) cases were found PCR positive; all PCR positive cases were found in risk group and no cases were found positive in non risk group of study population.

Table 5: Real time PCR positive cases among the PCR positive and PCR negative cases

Case	Real time PCR positive	Real time PCR negative
PCR positive (n=3)	02	01
PCR negative (n=52)	00	52
Total (n=55)	02 (3.64%)	53 (96.36%)

Table 5: showing the distribution of real time PCR positive cases among the PCR positive and PCR negative cases. Out of 55 cases 3.64% (2/55) were real time PCR positive. No Real time PCR positive cases were found from PCR negative cases. All PCR and Real time PCR positive cases were found in risk group of study population.

123 4 5 6 7 8 9 10 11 12 13 14 15 16 17 18

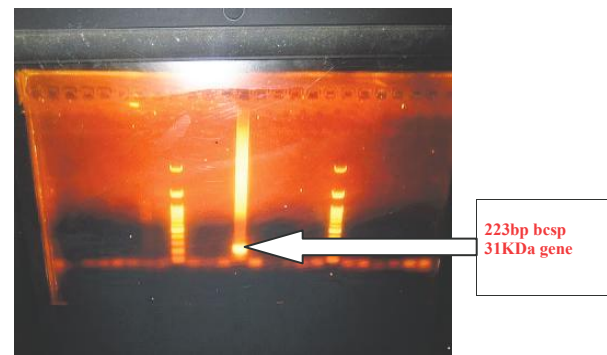


Figure 1: showing the PCR product of 31 KDa outer membrane gene

Discussion

In developing countries, Sub Saharan countries and African countries, Brucellosis is a major bacterial zoonotic disease found in human and domestic live stock. this affects the poor people livelihood by affecting their domestic live stock and thus causing them to compromise their health, survival and economic status²⁰. In practice blood culture is positive in 10%-30% of Human Brucellosis cases and remainder is diagnosed serologically and by molecular methods²¹. Although no single test provides 100% sensitivity and specificity, a variety of serological tests and molecular tests have been applied for diagnosis of Human Brucellosis. In the present study majority of the cases 49.17% (295/600) were in the age group between 20-40 years and total 65.83% (395/600) were male and 34.17% (205/600) were female. Male female ratio is 2:1 (table 1). The predominance of study population in age group (20-40yrs) and male was due to people in this age group being more active and as there are the main earning member of the family in our society. So they have more chance to occupational exposure⁴. The sero prevalence of Brucellosis by *Brucella* specific latex agglutination test in risk group was 13.33 % (40/300) and non risk group 5%(15/300) respectively (Table-2). Some other study in our country showed that the sero prevalence of Brucellosis were 6 to 12.8%⁸ in risk group. The increased prevalence rates in risk group of population may be due to occupational contact with domestic animal and lack of personal hygiene. The prevalence of Brucellosis by using IgM/IgG (ICT) kit was 27.27% (15/55) (table-3). All the positive cases were found positive by IgM kit. No IgG kit were found positive. This indicates the cases were acute or recent infection. In a study at Borana and from Hamar,

Ethiopia showed that 34.1% and 29.4% patients were sero positive for Brucellosis using *Brucella* immunoglobulin M (IgM), lateral flow assay, which was almost similar to our current study. It appears, no data available regarding the use of ICT IgM/IgG device for detection of Human Brucellosis in Bangladesh. In our present study we detected *Brucella* genome specific gene encoding outer membrane protein (LPS). (Table 4). Out of 55 sero positive samples 5.45% (3/55) were PCR positive. All positive cases were found in risk group. No cases from non risk group were found positive by PCR. The present study also detected the *Brucella* genus species DNA by real time PCR method; the findings of real time PCR from the sero positive cases were showed in (table 5). Out of 55 samples 3.64% (2/55) were found positive. The cut off value of real time PCR test were 0.849195 indicating a positive reaction in both situations. Out of 3 PCR positive cases 2 were real time PCR positive. The low prevalence of PCR and real time PCR positive cases in our country may be due to Brucellosis not being endemic in our country.

Conclusion and Recommendations

The present study revealed that a considerable number of Human Brucellosis is present in risk group as well as non risk group of population in both rural and urban area in Mymensingh district. More sensitive and specific test is required to diagnose the disease more rapidly and accurately, as latex agglutination test is non specific and gives many false positive results. Consequently Human Brucellosis should be included as a differential diagnosis in PUO cases especially in PUO with history of occupational exposure. Further study on the subject from different region of Bangladesh should be carried out in long scale to find out the true prevalence of the disease.

Limitations

Non availability as well as high cost of the reagent in local market limited the number of case in this study.

Time constrain, the study was done in a limited period of time.

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