

5-Bromo-3-(3-hydroxyprop-1-ynyl)-2H-pyran-2-one targets prostate cancer cells by down-regulating inflammation-related genes

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Abstract

The present study was performed to examine the effect of 5-bromo-3-(3-hydroxyprop-1-ynyl)-2H-pyran-2-one (BHP) on the rate of cell proliferation and apoptosis induction in the PC3, human prostate carcinoma cell line. The cell viability was assayed by using sulphorhodamine B staining and apoptosis by annexin V and flow cytometry analyses. The results revealed that BHP treatment in PC3 cells caused a significant reduction in the rate of cell proliferation in dose- and time-dependent manner. Compared to the untreated cells, the formation of HUVEC tubes was markedly inhibited on treatment with BHP at a concentration of 30 μ M. Further investigation revealed the expression of HMGB1, IL-6 and IL-8, pro-inflammatory cytokines was also inhibited on treatment with BHP. Therefore, BHP treatment plays an important role in inducing apoptosis in the prostate cells and can be of therapeutic value for the prostate cancer treatment.

Introduction

Prostate cancer, one of the frequent detected causes of mortality in men, accounts for more than 25,000 deaths in the United States alone every year (Jemal et al., 2010). The treatment strategies generally used for the treatment of prostate cancer in its primary stage include, androgen ablation, surgical removal of the tumor mass and or radiotherapy. However, use of hormones ultimately induces resistance. Therefore, screening of the natural products for investigation and development of the novel treatment strategies for prostate cancer is preferred. In the process of prostate carcinoma cell metastases to adjacent tissues angiogenesis and invasion have been reported to exhibit vital role (Weidner et al., 1993; Wakui et al., 1992). Angiogenesis in prostate carcinoma cells influences the expression levels of p53, VEGF and MMP-9. Another important factor with role in the progress of cancer inflammation which is initiated and regulated by cytokines and chemokines (Lu et al., 2006; Coussens and Werb, 2002).

It has been shown that concentration of IL-6 and IL-8, cytokines is higher in the patients with hormone resistant than in hormone-sensitive disease (Veltri et al., 1999; Pfizenmaier et al., 2003; Uehara et al., 2005).

2-Pyrones, the cyclic 6-membered esters are the natural isolates obtained from the extracts of plants, bacteria and other animals by using conventional chromatography (Jensen and Fenical, 1994; Faulkner, 2001). Their role in providing protection to the organisms against various other attackers is well established (McGlacken and Fairlamb, 2005). The dihydro-2-pyrones isolated from microbial sources have shown a diverse range of biological activities. Some of their activities include anticancer, anti-HIV, antifungal, neurotoxic, cytotoxic, phytotoxic, and activities (Dickinson, 1993, Thaisrivongs et al., 1996; Poppe et al., 1997; Turner et al., 1998). The mechanism of anticancer activity of 2-pyrones involves inhibition of DNA synthesis in the cells (Trachootham et al., 2008). In the human leukemic cells, 2-pyrones have been shown to inhibit cell growth

through suppression of DNA synthesis (Trachootham et al., 2008).

The present study was performed to investigate the effect of 5-bromo-3-(3-hydroxyprop-1-ynyl)-2H-pyran-2-one (BHP), an analog of 2-pyrone, in the inhibition of cell growth and induction of apoptosis in the prostate cancer cells.

Materials and Methods

Chemicals and reagents

BHP and DMEM was obtained from Sigma-Aldrich (St. Louis, MO). Cell viability assay kit and annexin V-FITC apoptosis detection kit were purchased from Santa Cruz Biotechnology (USA). The other common chemicals were purchased from Sigma-Aldrich (St. Louis, MO).

Cell lines and cell culture

PC3 cell line was obtained from Korean Cell Line Bank (South Korea) and cultured in DMEM medium supplemented with 10% fetal bovine serum. The cells were maintained at 37°C, 5% CO₂ environment.

Cell proliferation assays

For the determination of cell viability MTT [12 µL, 5 mg/mL in phosphate buffered saline (PBS)] assay was used. PC3 cells were grown in DMEM supplemented with 10% FBS and cultured in an incubator with humidified atmosphere of 5% CO₂ at 37°C. The cells were dispersed on to 96-well plates at a density of 2.5 × 10⁵ cells per well. To each of the well different concentrations of BHP were added and incubated for 48 hours where as the untreated and control cells were treated with DMSO. After incubation, MTT reagent (Sigma-Aldrich) was added to each of the well and incubation was continued for 4 hours more. The contents dissolved in dimethyl sulfoxide were subjected to the measurement of absorbance at 455 nm wavelength using an ELISA reader (BD Biosciences, USA).

Determination of apoptosis by fluorescent microscopy

For the purpose of determination of apoptosis in PC3 cells annexin V-FITC apoptosis detection kit was used. The apoptotic cells were differentiated from the necrotic cells by using propidium iodide staining. PC3 cells were allowed to attach on to the culture glass slides and then the cells were exposed to BHP. Following 48 hours of incubation with BHP the cells were treated with 1X binding buffer, annexin V-FITC and propidium iodide reagents. After incubation for 20 min at 37°C under dark conditions fluorescent microscopy (Leica Microsystems GmbH, Germany) was used to analyze the apoptotic cells.

Tube formation assay

In order to determine the tube formation property,

polarization of the matrigel (BD Biosciences, USA) was performed at 37°C for 40 min. HUVECs at a density of 2.5 × 10⁵ cells per ml were put in ECGM2 medium supplemented with 50 ng/mL VEGF. The cell suspension (0.2 mL) was put in to each of the matrigel-coated well and treated with BHP or DMSO as control for 24 hours. The phase-contrast microscope was used to examine the alteration in the morphology and formation of tubes whereas the images were captured at magnification ×200.

Cell invasion and motility assay

The invasive ability of the tumor cells was determined using matrigel invasion chamber. The membranes composed of polycarbonate bearing 8 µm sized pores were coated with matrigel (10 µg/cm²) and then incubated overnight. The BHP treated cells after incubation for 48 hours were put in the upper chamber at a density of 2.5 × 10⁶ cells in RPMI-1640 supplemented with 1% BSA. The lower chamber used as a chemoattractant contained RPMI-1640 medium supplemented with 10% FBS. Following 24 hours incubation, the membranes were fixed in 4% paraformaldehyde. After 30 min the membranes were stained with eosin washed with PBS and removed from the inserts. The membranes were cleared from the upper side and membranes were mounted using glycerol. For the calculation of invaded cells five regions were randomly selected and the cells were calculated three times in each region.

Western blot analysis

The BHP treated cells were washed three times with cold PBS and then treated with 120 µL radioimmunoprecipitation assay buffer [50 mM Tris-HCl, pH 6.8; 0.1% SDS, 150 mM NaCl, 1 mM EDTA, 0.1 mM Na₃VO₄, 1 mM sodium fluoride (NaF), 1% Triton X-100, 1% NP-40, 1 mM dithiothreitol, 1 mM PMSF, 1 µg/mL aprotinin, 1 µg/mL leupeptin and 1 µg/mL pepstatin A]. The cell lysates were collected into 1.5-mL culture tubes and then centrifuged at 12,000× g for 30 min to collect the supernatants. Bicinchoninic acid assay (Sigma-Aldrich) was used for the determination of concentration of proteins. Electrophoresis on 10% (w/v) SDS-polyacrylamide gel was used for the isolation of the proteins. The proteins were then transferred onto polyvinylidene difluoride membranes which were blocked with 5% skimmed milk in buffer [10 mM Tris-HCl (pH 7.6), 100 mM NaCl and 0.1% (v/v) Tween-20]. Incubation of the membranes with primary antibodies was performed overnight. The membranes were washed with Tris-buffered saline and Tween-20 followed by incubation with the secondary antibodies for 1 hour. Tanon Gel Imager system (Tanon, China) was used for the semi-quantitative analysis of the proteins.

Real-time quantification PCR

A total of 2.5 × 10⁶ cells in 100-mm dishes were treated

with BHP for 48 hours followed by cell lysis. The RNeasy Plus Mini kit (Qiagen, USA) was used to extract the cell RNA and Superscript III First-strand Synthesis Super Mix for qRT-PCR (Invitrogen, USA) for RT-PCR. The NanoDrop1000 (Thermo Fisher Scientific, USA) was used for the determination of concentration of cDNA. For PCR an initial incubation was performed at 50°C for 2 min, then denaturation at 95°C for 10 min followed by 50 cycles of 95°C for 15 sec and 60°C for 1 min. The expression level of HMGB1, IL-6, IL-8 and (NAG-1) genes was analyzed using a SYBR green based gene specific real-time PCR quantification kit (SA Biosciences).

Statistical analysis

All the data were analyzed using ANOVA and Student-Newman-Keuls test. The results expressed the mean \pm SD. $p < 0.05$ was considered statistically significant.

Results

Effect on the growth and proliferation of PC3

Examination of the effect of BHP on the rate of cell proliferation in PC3 and DU-145 cells showed a concentration and time dependent inhibition. Increase in the concentration of BHP from 10 to 40 μ M significantly increased the inhibition of cell growth. The inhibition of cell proliferation in PC3 and DU-145 cells at 40 μ M concentration of BHP was 76 and 74%, respectively after 48 hours (Figure 1).

Effect on the induction of apoptosis in PC3 cells

The results from annexin V-FITC and propidium iodide staining showed a concentration dependent effect of BHP on apoptosis in PC3 cells after 48 hours (Figure 2). With the increase in concentration of BHP from 10 to 40 μ M, the proportion of apoptotic cells increased from 12

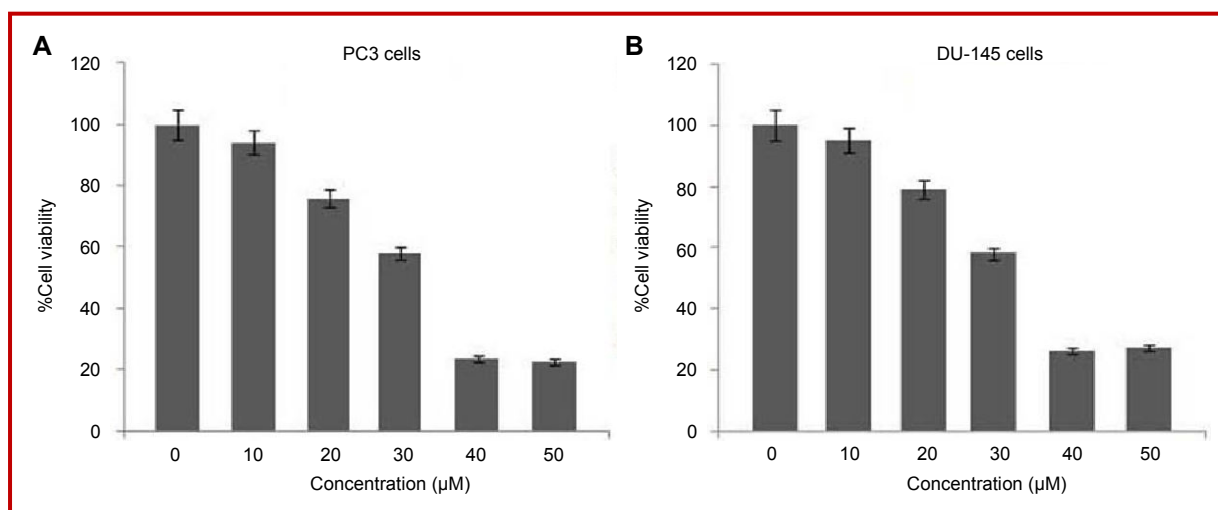


Figure 1: Effect of BHP on inhibition of proliferation in PC3 and DU-145 cells. The viability of the cells was determined by MTT assay. BHP caused significant reduction in the viability of PC3 and DU-145 cell lines

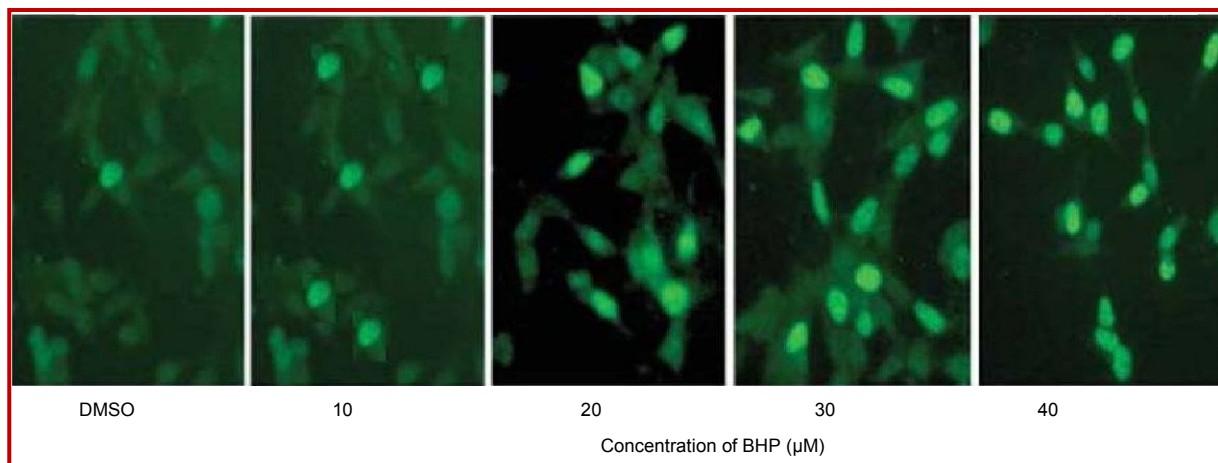


Figure 2: BHP treatment induces apoptosis in PC3 prostate carcinoma cells. PC3 cells after incubation with BHP or DMSO as control for 48 hours were stained using annexin V-FITC and propidium iodide. The fluorescence microscopy was used for the visualization of apoptotic cells

to 62%. The increase in the proportion of apoptotic cells on enhancing the concentration of BHP was further confirmed by flow cytometry analyses (Figure 3).

Effect on the tube formation of HUVEC and invasive ability of PC3 cells

Examination of the effect of BHP on tube formation of HUVEC cultured with BHP treated PC3 prostate cancer revealed marked inhibition in tube formation on matrigel (Figure 4). On the other hand, HUVEC cultured with control PC3 prostate cancer cells showed

extensive formation of tubes. The results from matrigel trans well assay revealed that BHP treatment in PC3 cells caused a significant decrease in the percentage of cells penetrating through matrigel coated transwell filters.

Effect on the expression of p65, VEGF and MMP-9 in PC3 cells

The process of angiogenesis and the ability of cancer cells to invade other tissues is controlled and regulated by NF- κ B (p65), VEGF and matrix metallo-proteinase-9

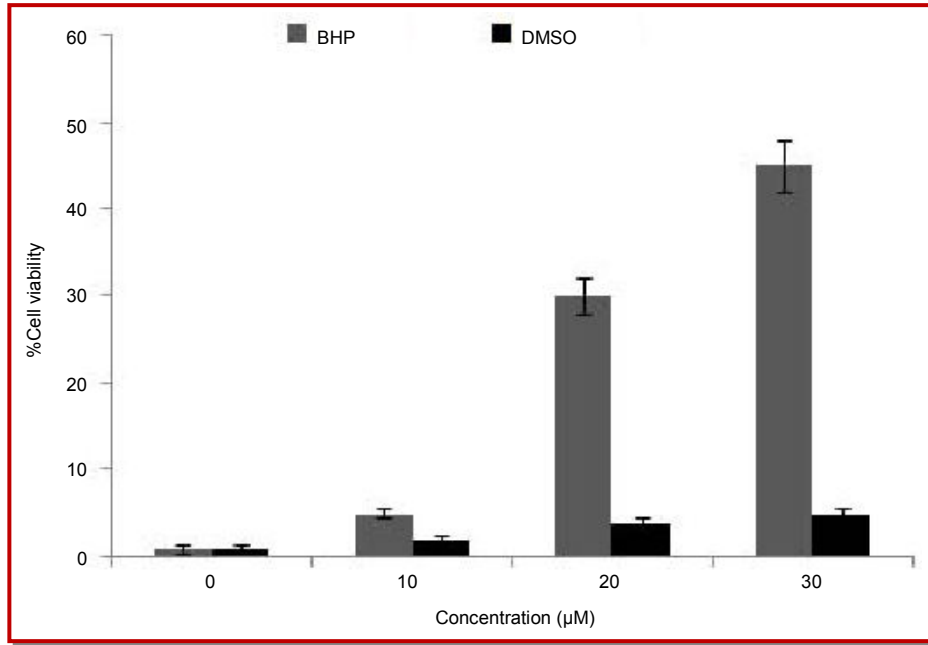


Figure 3: BHP treatment induces cell death in PC3 prostate carcinoma cells. PC3 cells treated with BHP for 48 hours followed by PI staining and flow cytometry analysis

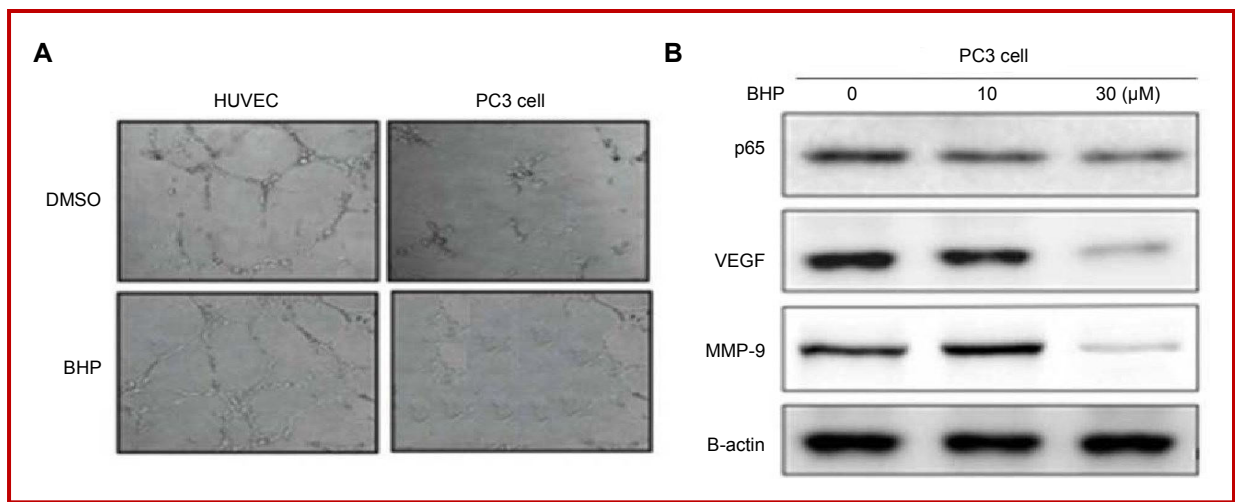


Figure 4: BHP exposure inhibited HUVEC tubulogenesis and PC3 cell invasion. (A) BHP influences tube formation by the endothelial cells through involvement of angiogenic factors in PC3 cells. PC3 cells cultured in the presence of BHP on matrigel pre-coated transwells plates for 24 hours showed inhibition in invasive ability on matrigel. (B) PC3 cells exposed to BHP for 24 hours showed inhibition in the expression of p65, VEGF and MMP-9 by immunoblot analysis

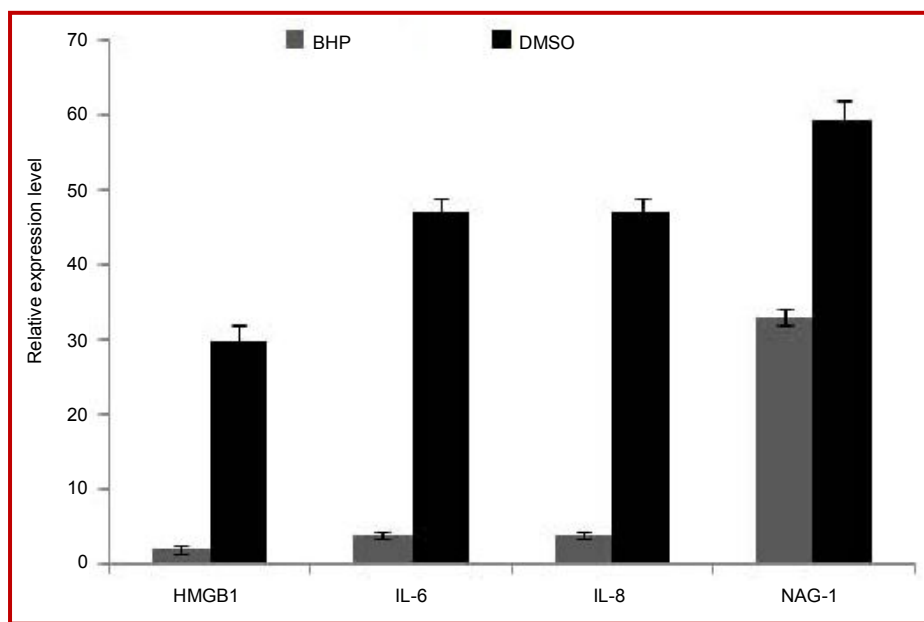


Figure 5: Inhibition of HMGB1, IL-6 and IL-8 expression and promotion of NAG-1 expression by BHP treatment in PC3 cells. The PC3 cells were incubated with BHP or DMSO as the control for 48 hours at 37°C. After incubation, expression levels of HMGB1, IL-6, IL-8 and NAG-1 genes were quantified by real-time PCR

(MMP-9). Results from the immune blot analysis showed that exposure of PC3 cells to BHP significantly inhibited the p65, VEGF and MMP-9 expression levels compared to the control cells.

Effect on the expression of inflammatory genes

The results from the quantitative real-time RT-PCR analysis revealed that BHP treatment in PC3 cells decreased the HMGB1, IL-6, and IL-8 expression levels and promoted the NAG-1 expression compared to the control cells (Figure 5).

Discussion

The results from our study using annexin V binding revealed that BHP inhibited cell proliferation through apoptotic cell death. The metastases of the carcinoma cells to the distant tissues are promoted by the processes like angiogenesis and invasion (Weidner et al., 1993; Wakui et al., 1992). The results from the present study showed that BHP treatment inhibited the invasive property and angiogenesis in PC3 cells. BHP treatment also caused inhibition in the expression of p65, VEGF and MMP-9. Thus, the inhibition of invasive property and angiogenesis in the PC3 cells by BHP involves reduction in the expression of p65, VEGF and MMP-9. Thus, BHP can be a promising candidate for the treatment of prostate carcinoma.

The factors like cytokines and chemokines are responsible for the creation of feasible environment for progress of the cancer. Inflammation is associated with

facilitating the progress of cancer (Lu et al., 2002; Coussens and Werb, 2002). The patients with hormone resistant disease show higher concentration of IL-6 and IL-8 in the serum (Veltri et al., 1999; Pfitzenmaier et al., 2003; Uehara et al., 2005). The results from the present study demonstrated that BHP treatment inhibited the expression of HMGB1, IL-6 and IL-8 in PC3 cells. Reduction in the expression of HMGB1 has been reported to induce apoptosis in the prostate cancer cell (Gnanasekar et al., 2009).

Conclusion

BHP treatment inhibits the cell proliferation, induces apoptosis, inhibits the tube formation and invasive ability of the prostate carcinoma cells.

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