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Antidiabetic potential of *Gaultheria trichophylla* in mice

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Introduction

Diabetes is a predominant public health concern and its prevalence is increasing all over the world. To treat type 2 diabetes mellitus and associated obesity, the potent glucosidase inhibitor from the natural source can be used (Ozaki et al., 2008). The people are diverting toward the herbal products because they are easily available, have low cost and some people are of the view that these are safe. Some of the allopathic medicines, like insulin and hypoglycemic agents taken orally are mainly used, but they bear some complications. These problems are expected to be partially overcome by the use of medicinal herbs because of their adverse effects and more accessibility (Shruthi et al., 2012).

Recently many plant species have been studied for their antidiabetic effects*.* For example, *Cephalaria gigantean* (Mbhele et al., 2015), *Oncocalyx glabratus* (Ahmed et al., 2015) and *Pistacia lentiscus* (Saad Ur Rehman et al., 2015) showed to possess antidiabetic activity.

Gaultheria trichophylla Royle (Ericaceae) is also known as Himalayan snowberry. The plant of genus Gaultheria is employed in the management of arthritis in traditional medicine (Liu et al., 2013), as anti-inflammatory agents (Zhang et al., 2011), and antibacterial activity (Cybulska et al., 2011). The author has explored this plant for its cytotoxic potential on cancer cell lines (Alam et al., 2015).

There are a paucity of information on the *Gaultheria* plants for their antidiabetic potential. For example, study has been carried out on the *Gaultheria hispidula* (Harbilas et al., 2009). Keeping in view the utilization of natural products as safe and effective agents for diabetes treatment, it is necessary to further explore natural sources. This study was, therefore, conducted to evaluate the antidiabetic potential of the methanol extract of *G. trichophylla* for *in vivo* and *in vitro* models.

Materials and Methods

Chemicals

Chemicals used were of analytical with high purity grade procured from the standard commercial sources.

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Methanol, diethyl ether, ethanol, ethyl acetate, chloroform, *n*-hexane were purchased from the Merck (Germany); Glibenclamide and alloxan monohydrate from the Sigma Aldrich; DMSO from the Fluka Chemicals; Glucose 5%, normal saline 0.9% from the Shahzeb Pharma, Pakistan; Cholesterol kit, triglycerides kit, acarbose and a-glucosidase from the Sigma-Aldrich Co., USA.

Plant material

G. trichophylla plant (5 kg) was collected in November 2013 from the Kaghan valley (Latitude: 34° 54' 14.99" N, Longitude: 73° 38' 30.59" E, altitude of 8,202 feet), District Mansehra, KPK, Pakistan. The plant was identified by the second author. It was placed at the herbarium of the institute labeled with a voucher specimen number (CTPHM-GT01, 13). The plant material was washed, shade dried and pulverized to powder. The powder material was kept in closed container before the process of extraction with organic solvent.

Preparation of the plant extract

The plant material in powder form (100 g) was extracted using soxhlet extractor for 20 hours each with organic solvents i.e. methanol, chloroform and *n*hexane. It was preceded with the filtration process using Whatman grade 1 filter paper. In the next step, the filtrate was reduced to concentrated residue under reduced temperature (40°C) and pressure on vacuum rotary apparatus. The extraction yield was measured. The respective percentage yield for methanol, chloroform and *n*-hexane extracts were 21.9%, 16.4% and 7.0%. The desiccator was used to remove the remaining moisture, and finally the extracts were stored in the air tight containers at 4°C for further use.

Experimental animals

Healthy adult albino mice (26-30 g) of either sex were selected for the study. The animals were obtained from the National Institute of Health (NIH) and then bred in the Animal house of CIIT Abbottabad. Mice were kept in polypropylene cages (47×34×20 cm) lined with husk (changed every 24 hours). They were given a standard diet and water *ad libitum*. The pellets consisted of protein (23%), lipids (5%), crude fiber (4%), ash (8%), calcium (1%) , phosphorus (0.6%) , glucose (3.4%) , and carbohydrates (5%).

α-Glucosidase inhibitory assay

The assay was carried out according to the method described elsewhere with slight modification (Matsui et al., 1996) . All the samples were dissolved in DMSO. An enzyme solution consisting of α -glucosidase (0.8 units/ ml) in 50 mM phosphate buffer (pH 7), containing 100 mM NaCl was prepared fresh. The solution was maintained cool throughout the test. The substrate,

pNP-G (0.7 mM) in phosphate buffer, was prepared fresh earlier to use. The test solution $(20 \mu L)$ and enzyme solution (80 μ L) was pre-incubated at 37 \degree C for 5 min. The reaction was initiated with 1.9 mL of substrate solution and incubated for 15 min at 37°C. The reaction was stopped by adding 2.0 mL (0.5M) aqueous Tris solution, and the absorbance of PNP released from the PNP-G was noted at 400 nm. DMSO (20 µL) was kept as blank (without addition of the test solution). Acarbose was used as a positive control. Analysis was carried out in triplicates, and the results were calculated as ± SEM.

The a-glucosidase inhibitory activity was presented as the %age, calculated as

$$
= 100 - (AB-AS)/AB
$$

Where $AB =$ absorbance of blank, $AS =$ absorbance of the sample

Oral glucose tolerance test

The oral glucose tolerance test was conducted before the induction of diabetes in 18 hours fasted normal mice as per (Weir and Bonner-Weir, 2004). Healthy mice were randomly selected and distributed into five groups (n=6). Glucose 2 g/kg was fed. Blood glucose levels were measured from the vein of mice tail at 0, 60, 90, 120 and 150 min of glucose administration.

Induction of diabetes and experimental design

Antidiabetic activity was carried out on selected healthy albino mice (Ou et al. 2013). Diabetes was induced in mice using freshly prepared solution of alloxan in 0.9% w/v of NaCl. For inducing diabetes, the mice were kept on fasting for 12 hours and were given a single i.p. injection of alloxan monohydrate (150 mg/ kg). To prevent fatal hypoglycemia initially due to massive pancreatic insulin release, the mice were provided with 5% glucose solution after six hours supplied in water bottles in their cages for next 24 hours. Animal were kept at room temperature (27 ± 2° C), humidity (55 \pm 5%), and a 12 hours' cycle of light and dark.

The glucose level of the fasting animals was measured after 72 hours. After acclimatization, the animals were separated into following the groups (six mice in each group); Groups A, Normal control treated with saline; B, Diabetic control; C, Diabetic mice treated with 500 mg/kg body weight of the extract; D, Normal mice given 500 mg/kg of methanol extract and E, reference control treated with glibenclamide (10 mg⁄kg). An identification mark was given to the mice of each group on the tail with a permanent marker. Each of mice was weighed and the dose was calculated accordingly. The extract was given orally. All the groups were given respective treatments daily for 15 days. To check the effect of the extracts on the weight of animals, weight of

the mice was recorded prior to the administration of the extracts and at the end of the study as well i.e. on the day 15.

The blood samples were collected in glass test tubes and allowed to clot for 1 hour at 37°C. The blood was collected using capillary tubes into the Eppendorf Tubes® containing heparin for analysis of plasma profile. With a glass Pasteur, carefully, the clot was loosened from the sides of the tube. The serum was centrifuged at 5,000 rpm for 5 min at 4° C. A micropipette was used gently to separate the serum from the clot. The serum was labeled with the animal number and the estimations were made (Phuong et al., 2004).

Biochemical analysis

The blood sugar level was measured using Accu-Chek Active test meter by withdrawing the blood on the test strips from the vein of mice tail. Total cholesterol (TC) and triglycerides (TG) were assayed using the protocol described elsewhere (Burtis et al., 2012). The level of serum urea and creatinine were assayed using the protocol described elsewhere (Thomas et al., 1998). High density lipoprotein (HDL) and low density lipoprotein (LDL) were measured by the protocol described elsewhere (Burstein et al., 1970).

Statistical analysis

All the data obtained from the body weight, fasting blood sugar, and other biochemical assessments were represented as mean ± standard deviation and analyzed for ANOVA–Dunnet's test. The differences between groups were considered significant at p<0.001 and p< 0.05 levels. The normal control was compared with the normal extract treated groups while diabetic control was compared with the diabetic extract treated and glibenclamide treated groups.

Results

α-Glucosidase inhibitory activity

The methanol extract of *G. trichophylla* demonstrated high activity against α -glucosidase (98.4 ± 2.5% inhibition and with $IC_{50} = 17.5 \pm 0.1 \text{ µg/mL}$, the chloroform and hexane extracts showed no prominent effect (49.5 ± 2.0 and $33.1 \pm 3.2\%$ inhibition respectively) (Table I). The standard drug acarbose was used as control (92.2 ± 0.1% (0.5 mM) inhibition and with IC₅₀ = 38.2 ± 0.1 μg/ mL. Therefore, it was decided to test the methanol extract for *in vivo* activity on animal model.

Effect on body weight

The effect of extract of *G. trichophylla* on body weight of mice showed that the change in the body weight of the normal groups $(27.7 \pm 2.3 \text{ g})$ (untreated) increased (34.3 g) ± 5.3 g) after 15 days (Table II). Similarly, the body

Data represented as mean \pm S.D values of 6 animals each ^{a}p <0.001, ^bp<0.05 (One-way NOVA, Dunnet's t-test, GraphPad Prism software)

weight of the normal groups $(29.6 \pm 2.3 \text{ g})$ treated with the methanol extract also increased $(34.0 \pm 1.8 \text{ g})$. The body weight of the diabetic control groups $(32 \pm 1.8 \text{ g})$, as expected, decreased $(25 \pm 3.8 \text{ g})$ significantly (p<0.05) after 15 days. The body weights of diabetic mice treated with methanol extract (33.0 ± 2.1 to 30.0 ± 3.1 g) of *G. trichophylla* and the standard (glibenclamide) (30 ± 1.8 to 29.3 ± 1.8 g) did not reduced to extent as observed in diabetic mice group and this difference between the groups was also significant (p<0.05).

Data represented as mean \pm S.D values of 6 animals each ^{a}p < 0.001, ^bp<0.05 (One-way NOVA, Dunnet's t-test, GraphPad Prism software)

Effect on blood glucose level

The alloxan-induced diabetes had caused significant initial increase in the blood glucose levels (fasting) of all animal groups (Table III). The diabetic control group showed significant increase in blood glucose level throughout the study period as compared with the normal control group (p<0.001). However, the methanol extract treated and the glibenclamide-treated groups showed significant (p<0.001) reduction in the fasting blood glucose as compared to diabetic control.

The effect was more pronounced in the standard treated group, which showed significance decrease in the blood glucose level $(p<0.001)$ from day 3 to day 15

Data represented as mean ± SD values of 6 animals each ap<0.001, bp<0.05 (One-way ANOVA, Dunnet's t-test, GraphPad Prism software). Normal control was compared with normal control and extract treated. The diabetic control was compared with diabetic extract/standard treated groups

of the experiment. The methanol extract of *G. trichophylla* caused a significance (p<0.05) decrease in the glucose level on day 12-15.

Effect on TC, TG, HDL and LDL

There was no significance difference in TC and TG levels when normal control $(81.7 \pm 2.1 \text{ and } 75.3 \pm 4.3 \text{)}$ mg/dL) was compared with the normal extract treated

group (81.0 ± 1.9 and 73.7 ± 3.9 mg/dL) (Figure 1). The diabetic control (163.2 \pm 10.1 and 183.4 \pm 7.2 mg/dL) showed hyperlipidemia compared with normal control as indicated by increased level of TC and TG in diabetic mice. The extract (113.7 \pm 2.8 and 138.7 \pm 4.5 mg/dL) and standard (122.6 \pm 1.9 and 140 \pm 3.9 mg/dL)-treated diabetic groups showed a significant decreased in the serum level of cholesterol and triglycerides compared

Figure 1: Effect of extract and standard drug on total cholesterol (A), triglycerides (B), LDL (C) and HDL (D) levels of normal and alloxan-induced diabetic mice. Data represented as mean \pm SD values of 6 animals each ^ap<0.001, ^bp<0.05 (One-way ANOVA,

Figure 2: Effect of extract and standard drug on serum urea (A) and creatinine (B) levels of normal and alloxan-induced diabetic mice. Data represented as mean ± S.D values of 6 animals each ap<0.001, bp<0.05 (One-way ANOVA, Dunnet's t-test, GraphPad

to the diabetic control group (p<0.001). The results of hypolipidemic potential of the plant extract was comparable to standard drug.

Alloxan rendered diabetic groups showed a significant (p <0.001) decrease in HDL level (20.7 \pm 1.0 mg/dL) compared to the normal control $(42.5 \pm 2.5 \text{ mg/dL})$. The extract (41.4 \pm 1.4 mg/dL) and standard drug (37.7 \pm 2.2 mg/dL) in the course of treatment for 15 days showed a significance improvement of HDL level in diabetic mice compared to the diabetic control ((p<0.001).

In contrast to HLD level, the LDL level was significantly increased in diabetic control (96.9 ± 3.0) mg/dL) compared to normal control $(24.0 \pm 1.0 \text{ mg})$ dL). The extract (65.8 \pm 4.6 mg/dL) and standard drug $(63.4 \pm 3.8 \text{ mg/dL})$ treated groups showed to reduce the LDL level significantly (p<0.001) compared to the diabetic control after 15 days of treatment.

Effect on serum urea and creatinine level

Alloxan-treated diabetic mice showed a significance (p<0.001) increase in the serum urea level (17.7 \pm 0.8 mmol/L) (Figure 2). Normal mice treated with the methanol extract for 15 days showed no prominent change in the urea level $(4.6 \pm 0.4 \text{ mmol/L})$ when compared with the normal control $(5.1 \pm 0.4 \text{ mmol/L}).$ When compared with the diabetic control, the extracttreated groups after 15 days of treatment showed a significance (p<0.001) improvement (decrease) of urea level (11.9 \pm 1.5 mmol/L). The standard drug (15.6 \pm 0.6 mmol/L) also showed similar results.

When compared with the normal group (28.2 ± 1.8) mmol/L) the diabetic control group displayed a significant (p<0.05) increase in serum creatinine level $(37.3 \pm 0.8 \text{ mmol/L})$. The normal mice groups treated with methanol extract showed no prominent effect on the serum creatinine level (29.5 \pm 1.0 mmol/L). A significance (p<0.05) decrease in the serum creatinine level (33.7 ± 0.8 mmol/L) was observed with methanol

extract, and (p<0.001) standard drug glibenclamide $(32.8 \pm 2.3 \text{ mmol/L})$ when compared with the diabetic control.

Discussion

ǂ-Glucosidase is the key enzyme in the digestion of carbohydrates and it is present in the surface membranes of intestine. The α -glucosidase inhibitors suppress the postprandial hyperglycemia by retarding the liberation of glucose of oligosaccharides and disaccharides from complex carbohydrates in diet and therefore, delay the absorption of glucose (Gao et al., 2008). There is need to explore more natural sources which are safe and effective α -glucosidase inhibitors as potential agents against diabetes. Therefore, the methanol extract seemed to be the ideal inhibitor of this enzyme. In this study, the methanol extract of exhibited the strongest inhibition of α-glucosidase which was comparable with acarbose used as a positive control. This is depicted by the lowest IC_{50} generated with respect to the other extract and standard tested. This is in accordance with previous studies that an ideal antidiabetic agent should have a strong inhibitory activity against a-glucosidases (Bairy et al., 2016). The inhibition of this enzyme slows down the breakdown of disaccharides to monosaccharides such as glucose. This results in reduced quantity of glucose absorbed into the blood and thus ameliorating hyperglycemia (Kazeem and Ashafa, 2015).

Antidiabetic potential of other *Gaultheria* species is also evident by the earlier studies (Harbilas et al., 2009; Karki et al., 2014; Nachar et al., 1990; Suman, 2014). The methanol extract of *G. trichophylla* at a concentration of 500 mg/kg displayed significant effect on the glucose tolerance of mice and the extracts also exhibited reduction in the fasting blood glucose levels of the normoglycemic mice, thus revealing the hypoglycemic

effect of the extract. The hyperglycemic mice were treated for 15 days with the extracts and levels of glucose were measured. There was a gradual reduction of blood glucose levels in mice treated with the extract throughout the period of experiment. The results of the extract treated groups were compared statistically to glibenclamide and was observed as significant. Alloxan caused the destruction of β -cells of islets of Langerhans and stops the production of Insulin and results in induction of diabetes. Therefore, in this case the extracts might have produced the hypoglycemic effect by a mechanism not involving insulin. The other possible mechanism can be endogenous glucose generation inhibition or by the reduction of absorption of intestinal glucose and additionally enzyme inhibition mechanism cannot be over ruled as discussed earlier (Eddouks et al., 2004).

It is a common observation that the level of serum lipids is usually high in diabetes mellitus. This elevation can induce coronary heart disease. The hyperlipidemia that characterizes the diabetic conditions may be regarded as a result of the uninhibited actions of lipolytic hormones on the fat depots. Therefore, a drug therapy or a dietary provision can reduced the risk of vascular ailments by lowering the serum lipid concentration (Sikarwar and Patil, 2010). The decreased levels of TG and other cholesterol in diabetic animals that received glibenclamide revealed that these drugs do ameliorate dyslipidemia as described previously (Trivedi et al., 2004). The study revealed that the dose of 500 mg/kg of methanol extract recovered the level of serum TC and TG in a significant manner (p<0.001) compared to the diabetic control. The level of LDL, over a period of 15 days was significantly reduced (p<0.001) towards normal as compared with diabetic control. However, the level of cardioprotective lipid HDL was improved significantly by the extract in diabetic mice. The effect of extracts on HDL level of normal mice was not prominent. This shows significant hypolipidemic effect of the extract.

It has been observed that phenolic compounds could be responsible for the antidiabetic effect by preventing the destruction of β -cells through peroxidation chain reaction inhibition (Patel and Mishra, 2011). Previously, we have also evaluated the plant extract and have found to contain high phenolic contents (Alam et al., 2017). It is known fact that the alloxan inducted diabetes results in loss of body weight as indicated by diabetic control group. The results also revealed that extract and glibenclamide-treated groups did not show the loss in body weight as was observed in the diabetic control group. Increased serum levels of urea and creatinine are indicators of impaired renal function (Gerbes et al., 2002). Diabetic control mice showed an increased level of creatinine and urea and this level remained elevated throughout the period of study. The

normal and diabetic groups treated with extract showed a little improvement (reduction) in both the urea and creatinine levels as compared with the diabetic control group over the 15 days of treatment.

Conclusion

To the best of our knowledge, the present study is the first to report on the antidiabetic properties of *Gaultheria trichophylla*. The results show that oral administration of *G. trichophylla* extract can control increased glucose levels in diabetes and may be good source of a-glucosidase inhibitor and may be employed for the therapy of diabetes and its complication.

Ethical Issue

The approval of the Research, Ethical Committee (REC), department of Pharmacy, CIIT, Abbottabad was taken before the animal studies were conducted. The collection of plants from the Kaghan valley was made in such a way that the species' abundance was not disturbed. Care and handling of animals followed the internationally accepted procedures according to the Institute for Laboratory Animal Research´s Guide for the Care and Use of Laboratory Animals.

Conflict of Interest

All authors have completed the ICMJE uniform disclosure form and declare no support from any organization for the submitted work.

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