

BJP

Bangladesh Journal of Pharmacology

Research Article

Anti-snake venom potential of *Clerodendrum serratum* extracts on *Bungarus caeruleus* and *Daboia russelii* venom

Anti-snake venom potential of *Clerodendrum serratum* extracts on *Bungarus caeruleus* and *Daboia russelii* venom

Shwetha Vasudev¹, Sunil Shivajirao More¹, Gautham Santhekadur Annappa¹ and Veena Sunil More²

¹School of Basic and Applied Sciences, Dayananda Sagar University, Bangalore, Karnataka, India; ²Department of Biotechnology, Saphthagiri College of Engineering, Bangalore, Karnataka, India.

Article Info

Received: 7 May 2018
Accepted: 4 June 2018
Available Online: 14 June 2018
DOI: 10.3329/bjp.v13i2.36597

Cite this article:

Vasudev S, More SS, Annappa GS, More VS. Anti-snake venom potential of *Clerodendrum serratum* extracts on *Bungarus caeruleus* and *Daboia russelii* venom. Bangladesh J Pharmacol. 2018; 13: 187-91.

Abstract

Anti-snake venom therapy is the only treatment for snake bite but leads to acute and chronic conditions which may be severe. The medicinal plants have gained importance over years to find an effective alternative to anti-snake venom. The present study focused on evaluating the potential of *Clerodendrum serratum* for the anti-snake venom activity. Phytochemicals were extracted from the *C. serratum* with different solvents. The ethyl acetate and methanolic extracts were found to neutralize the major enzyme toxins (phospholipase A₂, protease and hyaluronidase) of *Bungarus caeruleus* and *Daboia russelii* venom at a concentration of 100 µg/mL. The fibrinolytic activity of both the venoms were neutralized. The study proves that the plant *C. serratum* possesses certain compounds which inhibit the toxins present in the venom of *B. caeruleus* and *D. russelii*.

Introduction

Around 3,000 species of snakes are identified so far, of which only 13-14% are found to be potentially dangerous to humans. Indian cobra (*Naja naja*), common krait (*Bungarus caeruleus*), Russell's viper (*Daboia russelii*) and saw-scaled viper (*Echis carinatus*) are considered responsible for snake bite deaths in India (Mukharjee, 2012).

Presently, anti-snake venom obtained from the sera of horse is the only available treatment and has various limitations like cost, storage facility, anaphylactic reactions and serum sickness.

Numerous attempts have been made by researchers to develop snake venom antagonists from plants on the basis of the fact that these medicinal plants possess rich bioactive compound with potent pharmacological activity (Martz, 1992; Soares et al., 2005). The plants are *Eclipta sp.*, *Casearia sp.*, *Curcuma longa*, *Mimosa pudica* (Meenatchisundaram et al., 2009), *Musa paradisiaca*,

Mucuna pruriens, *Bauhinia forficata*, *Hibiscus esculentus*, *Annoma senegallensis*, *Mikania glomerate* (Floriano et al., 2009), *Piper sp.*, *Schizolobium parahyba* (Mendes et al., 2008) etc.

Clerodendrum serratum L. Moon is a medically important plant, belonging to family Lamiaceae. Traditionally it is said to possess medicinal value in treating respiratory disorders, fever, rheumatism inflammation, snake bite etc. Scientific studies on *C. serratum* has revealed its therapeutic properties for asthmatic, mast cell stabilization and anti-allergic effects, also includes pharmacological activities such as hepatoprotective nature, antioxidant, anti-inflammatory and anti-cancer potential (Patel et al., 2014).

Snake venom neutralization studies was performed in genus *Clerodendrum* (*C. viscosum*) against *Naja naja* venom and was found to be effective which justifies the traditional use of *Clerodendrum viscosum* against snake bite (Richard et al., 2006). In the present study, attempt was made to evaluate the potential of *C. serratum*, for its



ability to neutralize the venom of *B. caeruleus* and *D. russelii*, two medically important snakes of India by *in vitro* and pharmacological methods.

Materials and Methods

Lyophilized venoms of *B. caeruleus* and *D. russelii* were obtained from Irula Snake Catcher's Cooperative Society, Kancheepuram, Chennai. Venom of known concentration was dissolved in physiological saline and centrifuged at 2000 x g for 10 min. Supernatant was used for further analysis and stored at 4°C. Protein content was estimated according to the method of Lowry et al., (1951).

Plant collection and extraction

Leaves of *C. serratum* was collected from sanjeevani vatika, Department of Horticulture, GKVK, Bangalore, Karnataka and was authenticated (Authentication no 44) by Dr. M. Vasundhara, at GKVK.

Leaves were thoroughly washed, shade dried, ground into fine powder and sieved to get uniform particle size. Powdered material was then subjected to continuous hot extraction (60-80°C) in soxhlet apparatus using solvents such as petroleum ether, chloroform, ethyl acetate and methanol. The extraction was carried out until the loop in the siphon tube was colourless. The extract obtained was concentrated using rotary vacuum evaporator [model: Biobase RE 100-pro] and dried at 40°C. The residue obtained was expressed as in terms of dry weight (g), which was used for further analysis.

Qualitative analysis of phytochemicals

The qualitative phytochemical analysis of the extracts was performed as per the method described by Kemp, (1986) and Sazada et al., (2009), to detect the presence of different phytoconstituents such as alkaloids, terpenoids, flavonoids, phenols, saponins, carbohydrates and other metabolites.

Enzyme inhibition studies

For the inhibition studies, the venom was pre-incubated with the extracts for 30 min at 37°C and the assays was performed accordingly in triplicates.

Hyaluronidase assay

Hyaluronidase assay was determined turbidometrically (Pukrittayakamee et al., 1988). The assay mixture contained Tris-HCl buffer (pH 8.0), 50 µg hyaluronic acid (in buffer) and enzyme in a final volume of 1.0 mL. The mixture was incubated for 15 min at 37°C, the reaction was stopped by the addition of 2 mL of 2.5% (w/v) cetyltrimethylammonium bromide in 2% (w/v) sodium hydroxide, and absorbance was read at 400 nm (within 10 min). Blank contained 1 mL of same buffer and 2 mL of 2.5% (w/v) cetyltrimethylammonium

bromide in 2% (w/v) sodium hydroxide. Turbidity-reducing activity was expressed as percentage of the remaining hyaluronic acid, taking the absorbance of a tube in which, no enzyme was added as 100%. One unit was defined as the amount of enzyme causing 50% turbidity reduction as 1.0 unit of international standard preparation. Specific activity is turbidity-reducing units per mg of protein per min. Inhibition studies was performed as mentioned earlier.

Phospholipase A₂ assay

Phospholipase A₂ activity was measured by the method of Marinetti (1965), with suitable modifications. Briefly, egg yolk was suspended in Tris-HCl buffer pH 8.0 and adjusted to an initial absorbance of 1.0 at 740 nm, venom sample (10 µL of 0.1%) was added to 5 mL of this suspension and the readings were recorded. Phospholipase activity was assessed as decrease in optical density over an incubation period of 1 min. One unit of phospholipase activity corresponds to the decrease of 0.001 absorbance per min. Activity was expressed as U/mg of venom. Inhibition studies was performed as mentioned earlier.

Protease assay

Reaction mixture for protease assay was composed of 0.5% casein, 1.0 mL Tris- HCl buffer (pH 8.0), and 0.5 mL 0.25% crude venom. The reaction mixture was incubated for 4 hours at 37°C. At the end of 4 hours, the reaction was topped by adding 4.0 mL of trichloroacetic acid (TCA) and mixture was filtered using Whatman No. 1 filter paper (Greenberg, 1955); and 1.0 mL of the filtrate was used for protein estimation using L-tyrosine as a standard (Lowry et al., 1951). In the above investigation, one unit of enzyme activity was defined as that which liberated 0.02 µmole tyrosine/hour under the experimental conditions described.

Pharmacological studies

Fibrinogenolytic assay

Fibrinogenolytic activity of venom was assayed as per the method described by Ouyang and Teng (1976). Reaction mixture composed of venom and Bovine fibrinogen (50 µg), incubated for 1 hour at 37°C. Reaction was terminated by the addition of 20 µL denaturing buffer (1M urea, 4% SDS and 4% β-mercaptoethanol). The hydrolysed product was analysed with 12% SDS and stained with Coomassie brilliant blue for visualizing the protein band. Venom to plant extract of 1:10 concentration was pre-incubated for 30 min at 37°C for inhibition studies.

Results

Extracts obtained from soxhlet extraction were dried using rotary vacuum evaporator and the weight was

expressed in terms of dry weight (w/w) where the yield per 100 g was as follows: Petroleum ether 0.16 g, chloroform 0.31 g, ethyl acetate 0.21 g and methanol 1.04 g. Enzyme inhibition studies was performed in triplicates for all the extracts.

The qualitative analysis of phytochemicals revealed the presence of alkaloids and flavonoids in chloroform extract, tannins, steroids, flavonoids and glycosides in ethyl acetate extract and saponins in methanolic extract. Phenols, resins and proteins were absent in the extracts.

Enzyme inhibition studies with crude extracts (100 µg/

Table I		
Activity of <i>B. Caeruleus</i> and <i>D. russelii</i> venom on enzymes		
Enzymes	Specific activity	
	<i>B. caeruleus</i>	<i>D. russelii</i>
Protease (µM/hour/mg)	1.2 ± 0.1	1.4 ± 0.1
PLA ₂ (U/mg)	634.2 ± 42.4	260.0 ± 35.3
Hyaluronidase (TRU/min/mg)	1.3 ± 0.8	0.5 ± 0.0

mL) was performed, all the extracts of the plant were found to inhibit hyaluronidase of both the venom (Table I), except petroleum ether did not show any inhibition on *D. russelii*. The PLA₂ activity was found to be inhibited by all the extracts of *C. serratum* however, the percentage inhibition was less than 50% at 100 µg concentration of plant extract for *B. caeruleus* venom where as PLA₂ inhibition was minimal in case of *D. russelii*. Similarly, protease of *B. caeruleus* was inhibited by all the extracts, highest inhibition was observed in E. acetate and methanol. In case of *D. russelii* only E. acetate extract had inhibited protease by 28% (Figure 1). Thus, it was found that, the plant exhibited good inhibitions on *B. caeruleus* venom compared to *D. russelii*.

Venom-induced fibrinogenolytic activity was analysed using bovine fibrinogen on 12% gel for both the venom (Figure 2A). Minimum fibrinogenolytic activity of *B. caeruleus* venom was observed at a concentration of 5 µg where Aα band was degraded and same was observed at concentration of 10 and 15 µg, whereas *D. russelii*-induced degradation of both Aα and Bβ band at a minimum concentration of 5 µg, further increase in venom concentration up to 10 and 15 µg showed similar

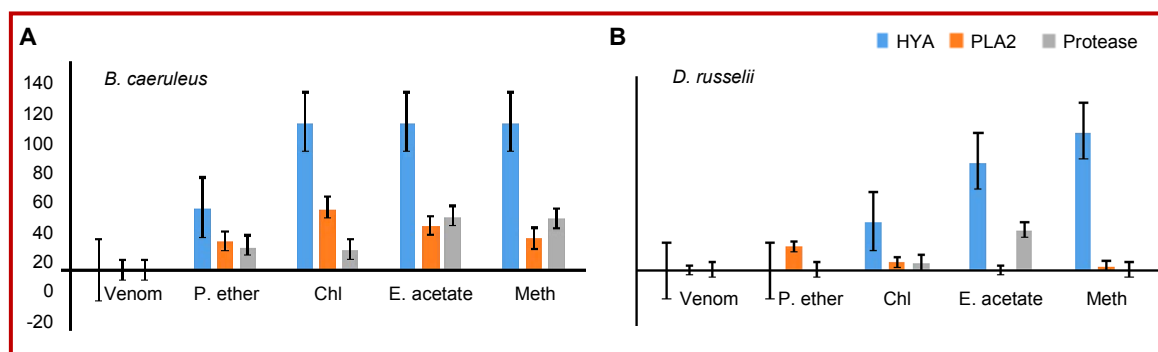


Figure 1: *In vitro* inhibition studies of *C. serratum* plant extracts against the MET's of *B. caeruleus* venom (A) and *D. russelii* venom (B). Extract are petroleum ether (P. ether), chloroform (Chl), ethyl acetate (E. acetate) and methanol (Meth)

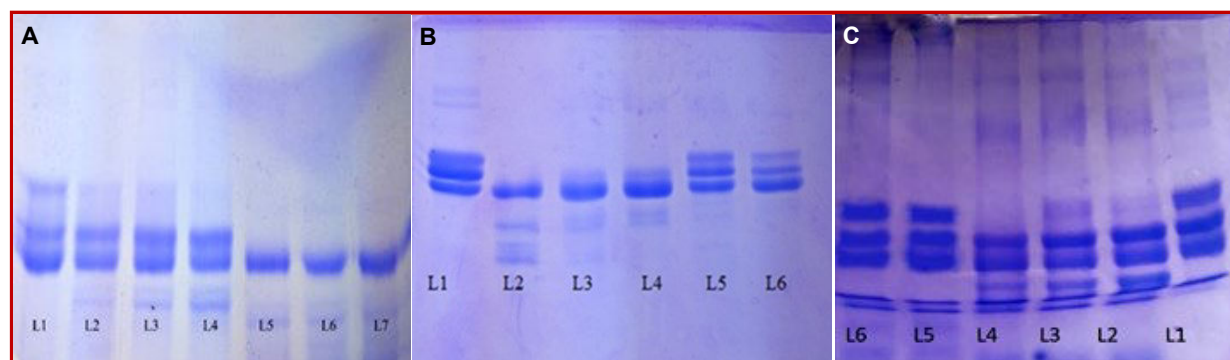


Figure 2: (A) Fibrinogenolysis induced by *B. caeruleus* and *D. russelii* venom at varying concentrations (5-15 µg). L1: Fibrinogen [F] (50 µg), L2-L4: F + *B. caeruleus* (5-15 µg) and L5-L7: F + *D. russelii* (5-15 µg); (B) Inhibition of fibrinogenolysis induced by *D. russelii* venom by different plant extract of *C. serratum*. L1: Fibrinogen [F] (50 µg), L2: F + DR + pet. ether, L3: F + DR + chloroform, L4: F + DR + ethyl acetate; L5: F + DR + methanol. DR= *D. russelii*; (C) Inhibition of fibrinogenolysis induced by *B. caeruleus* venom by different plant extract of *C. serratum*. L1: Fibrinogen [F] (50 µg), L2: F + BC + pet ether, L3: F + BC + chloroform, L4: F + BC + ethyl acetate; L5: F + BC + methanol. BC= *B. caeruleus*

results. The fibrinogenolytic activity of both the venom was neutralized by the ethyl acetate and methanolic plant extracts, where the fibrinogen structure remained intact at venom to extract at 1:10 (Figure 2B and 2C).

Discussion

The genus *Clerodendrum* has found to contain flavonoids as the predominant phytochemicals (Sinha et al., 1981), along with neo-clerodane diterpenes (Pandey et al., 2005), triterpenes, often phenolic compounds (Ganapaty and Rao, 1985) has also been reported, with some species showing the presence of macrocyclic alkaloids (Lumbu and Hootele, 1993) and cyanogenic glycosides (Miller et al., 2006), this is in agreement with the phytochemicals observed in the present study, absence of phenols may be due to phytochemical variation exhibited by the plants depending on its environmental conditions.

The potential of therapeutic properties evaluated for *C. serratum* has been credited for presence of phytochemicals like flavonoids, phenolics and saponins in the drug (Patel et al., 2014). It is interesting to note that in the present study the plant extract that showed inhibitions in ethyl acetate and methanol extract has the presence of those phytochemicals which has been shown to possess therapeutic properties in the previous studies.

In the present study, inhibition was performed on three enzymes of snake venom namely PLA₂, protease and hyaluronidase, also known as major enzyme toxins (METs) which if inhibited will be efficient in neutralizing the toxic effects of snake venom, although all extracts of *C. serratum* has shown inhibitions, on krait venom, the plant extracts were not able to efficiently inhibit *D. russelii* venom enzymes. This differential inhibition may be due to the structural differences exhibited by the venom belonging to two different classes, elapid and viperid. In a study reported by Mukherjee et al. (2008) a compound isolated from methanolic leaf extracts of *Azadirachta indica* exhibited significantly higher inhibition on *Naja naja* venom than on *D. russelii*. Hyaluronidase was found to be completely neutralized by ethyl acetate and methanolic extract of plant at a concentration of 100 µg/mL for both the venoms, hyaluronidase also known as spreading factor which helps in facilitating the easy diffusion of toxins, thus aiding the fast action of venom components. Inhibition of hyaluronidase facilitates in increasing the survival time by slowing the spreading of toxic components (Girish et al., 2004).

D. russelii and *B. caeruleus* venom possess several proteases that can lyse several important proteins, thus are said to be responsible for local and systemic effects of envenoming, extracts were able to inhibit the

protease of krait venom up to 40% at 100 µg/mL concentration whereas, there was no inhibitions observed for *D. russelii* venom. Further increasing the dose of the plant extract may help in evaluating the neutralization potency of the plant extracts. However, ethyl acetate and methanol extract of the plant was able to completely neutralize the fibrinogenolytic activity exhibited by both the venoms indicating its potency to neutralize snake venom serine protease.

It is interesting to note that many plant species belonging to Lamiaceae family has been reported to possess the ability to neutralize the venom components, both as per the literature, as well as through scientific studies for few plant species. For eg, Aqueous extract of *Ocimum sanctum* has been found to inhibit coagulation, fibrinolytic and PLA₂ activity of *Naja naja* venom at a concentration of 18 µg, 0.5 mg and 9 µg respectively (Kuriakose et al., 2012). In another study, *Rosmarinus officinalis* extract belonging to the same family has shown to completely inhibit the hyaluronidase activity of Egyptian desert horned viper *cerastes cerastes* (Wabhy et al., 2012). The study by Salama et al. (2017) has confirmed the efficiency of this plant both *in vitro* and *in vivo* proving its efficiency to treat lethal viper bites. Methanolic extracts of *Leucus asper* has shown to possess significant inhibitory effect on cobra venom (Kadiyala et al., 2014). Thus, plants belonging to lamiaceae has shown significant inhibitory effects on elapid and viper venoms, which shows that promising phytochemicals having snake venom antidote can be obtained.

Conclusion

The ethyl acetate and methanolic extracts of *C. serratum* has shown inhibitions on the venom enzymes *in vitro*, proving that the plant possesses the ability to neutralize the venom of *D. russelii* and *B. caeruleus*.

Acknowledgement

The authors would like to thank DST-SERB extramural grant (DST-SERB-EMR-2015/002086) for the project and also would like to acknowledge Ms. B. C. Spoorthi, Dr. Vineetha and Bhavya for help in video recording of the experiment.

Conflict of Interest

The authors declare that there is no conflict of interests regarding the publication of this article.

References

Floriano RS, Nogueira RMB, Sakate M, Laposy CB, da Motta

- YP, Sangiorgio F, David HC, Nabas JM. Effect of Mikania glomerata (Asteraceae) leaf extract combined with anti-venom serum on experimental *Crotalus durissus* (Squamata: Viperidae) envenomation in rats. *Rev Biol Trop*. 2009; 57: 929-37.
- Ganapaty S, Rao DV. Triterpenoids of the stem bark of *Clerodendron nerifolium*. *Indian J Pharm Sci*. 1985; 47: 167-68.
- Girish KS, Shashidhara MR, Nagaraju S, Gowda TV, Kemparaju K. Isolation and characterization of hyaluronidase a spreading factor from Indian cobra (*Naja naja*) venom. *Biochimie* 2004; 86: 193-202.
- Greenberg DM. Plant proteolytic enzymes. In: *Methods in Enzymology*. Colowick SP, Kalpan NO (eds). New York, Academic Press Inc., 1955; 54-64.
- Kadiyala G, Kadali R, Gurunathan J. Inhibition of *Naja naja* venom enzymes by the methanolic extract of *Leucas aspera* and its chemical profile by GC-MS. *Toxicol Reports*. 2014; 1: 667-73.
- Kuriakose, Aleykutty NA, Nitha B. Evaluation of venom neutralising capacity of Indian medicinal plants by *in vitro* methods. *Asian J Pharm Hea Sci*. 2012; 2.
- Lowry OH, Rosebrough NJ, Farr AL, Randall RJ. Protein measurement with the Folin phenol reagent. *J Biol Chem*. 1951; 193: 265-75.
- Lumbu S and Hootele C. Buchnerine and N1-(Z)-p-methoxycinnamoyl-buchnerine, two new macrocyclic *Clerodendrum buchmeri*. *J Nat Prod*. 1993; 56: 1418-20.
- Marinetti GV. The action of phospholipase A₂ on lipoproteins. *Biochim Biophys Acta*. 1965; 98: 554-65.
- Martz W. Plants with a reputation against snakebites. *Toxicon* 1992; 30: 1131-42.
- Meenatchisundaram S, Priyagrace S, Vijayaraghavan R, Velmurugan A, Parameswari G, Michael A. Antitoxin activity of Mimosa pudica root extracts against *Naja naja* and *Bangarus caeruleus* venoms. *Bangladesh J Pharmacol*. 2009; 4: 105-09.
- Mendes MM, Oliveira CF, Lopes DS, Vale LH, Alcântara TM, Izidoro LF, Hamaguchi A, Homsí-Brandeburgo ML, Soares AM, Rodrigues VM. Anti-snake venom properties of *Schizolobium parahyba* (Caesalpinoideae) aqueous leaves extract. *Phytother Res*. 2008; 22: 859-66.
- Miller RE, McConville MJ, Woodrow IE. Cyanogenic glycol-sides from the rare Australian endemic rainforest tree *Clerodendrum grayi* (Lamiaceae). *Phytochemistry* 2006; 67: 43-51.
- Mukharjee AK. Green medicine as a harmonizing tool to antivenom therapy for the clinical management of snake-bite: The road ahead. *Indian J Med Res*. 2012; 136: 10-12.
- Mukherjee AK, Doley R, Saikia D. Isolation of a snake venom phospholipase A₂ (PLA₂) inhibitor (AIPLAI) from leaves of *Azadirachta indica* (Neem): Mechanism of PLA₂ inhibition by AIPLAI *in vitro* condition. *Toxicon* 2008; 51: 1548-53.
- Ouyang C, Teng CM. Fibrinogenolytic enzymes of *Trimersurus mucrosquamatus* venom. *Biochim Biophys Acta Protein Structure*. 1976; 420: 298-308.
- Pandey R, Verma RK, Gupta MM. Neo-clerodane diterpenoids from *Clerodendrum inerme*. *Phytochemistry* 2005; 66: 643-48.
- Patel JJ, Acharya SR, Acharya NS. *Clerodendrum serratum* (L.) Moon: A review on traditional uses, phytochemistry and pharmacological activities. *J Ethnopharmacol*. 2014; 154: 268-85.
- Pukrittayakamee S, Warrell DA, Desakorn V, McMichael AJ, White NJ, Bunnag D. The hyaluronidase activities of some Southeast Asian snake venoms. *Toxicon* 1988; 26: 629-37.
- Richard L, Punitha ISR, Rajendran K, Arun S, Annie S. Preliminary study on the anti-snake venom activity of alcoholic root extract of *Clerodendrum viscosum* (Vent.) in *Naja naja* venom. *Nat Prod Sci*. 2006; 12: 153-56.
- Salama WH, Abdel-Aty AM, Fahmy AS. Rosemary leaves extract: Anti-snake action against Egyptian *Cerastes cerastes* venom. *JTCM*. 2017; 1-11.
- Sinha NK, Seth KK, Pandey VB, Dasgupta B, Shah AH. Flavonoids from the flowers of *Clerodendron infortunatum*. *Planta Medica*. 1981; 42: 296-98.
- Soares AM, Ticli TK, Marcussi S, Lourenço MV, Januário AH, Sampaio SV, Giglio JR, Lomonte B, Pereira PS. Medicinal plants with inhibitory properties against snake venoms. *Curr Med Chem*. 2005; 12: 2625-41.
- Wahby AF, Mahdy E, EL-mezayen H, Salama WH, Ebrahim NM, Abdel-Aty AM, Fahmy AS. Role of hyaluronidase inhibitors in the neutralization of toxicity of Egyptian horned viper *Cerastes cerastes* venom. *J Genet Eng Biotechnol*. 2012; 10: 213-19.

Author Info

Sunil S. More (Principal contact)
e-mail: sunilacr@yahoo.co.in