# ACHENE MICROMORPHOLOGY OF SEVEN TAXA OF ACHILLEA L. (ASTERACEAE) FROM TURKEY

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Keywords: Achene micromorphology; Achillea; Slime cells; Turkey.

### Abstract

Micromorphological characters of achenes in seven taxa of Turkish Achillea L. (Asteraceae) were investigated using stereomicroscope and scanning electron microscope (SEM). Some morphological descriptions of achenes were given for each species. A.biserrata Bieb. has the biggest  $(0.69\pm0.092 \times 2.01\pm0.252 \text{ mm})$  and A. grandiflora Friv. has the smallest  $(0.30\pm0.018 \times 1.12\pm0.058 \text{ mm})$  achenes. The achenes are oblong-lanceolate in A.biserrata and A. teretifolia Willd. and they are oblong in the remaining taxa. In surface sculpturing, the ornamentation and slime cell distribution varied among the taxa. However, A. biebersteinii Afan. has distinct slime cells forming groups scattered over the achene surface. Mature achenes are ribbed and glabrous in all studied taxa. A. biserrata has distinct carpopodium structure.

## Introduction

The genus Achillea L. (Asteraceae) includes about 140 species distributed in South-west Asia and South-eastern Europe (Akyalcın *et al.*, 2011). According to recent studies, the genus Achillea is represented in Turkey by 48 species (54 taxa), 24 of which are endemic for Anatolia (Akyalcın *et al.*, 2011). Achillea L. is classified into five sections, namely sect. Othantus (Hoffmanns. & Link) Ehrend. & Y. P. Guo (one species), sect. Babounya (DC.) O. Hoffm. (30 species), sect. Ptarmica (Mill.) W.D.J. Koch (2 species), sect. Anthemoideae (DC.) Heimerl (2 species) and sect. Achillea (13 species) (Huber-Morath, 1975; Duman, 2000; Arabacı and Yıldız, 2006; Arabacı and Budak, 2009).

Achene micromorphological characters have been found useful in systematics of the family Asteraceae (Abid and Qaiser, 2007a, b; Shekhar *et al.*, 2011). Cypselar external morphology and anatomy in members of different tribes of Asteraceae are found important for delimitation of genera (Garg and Sharma, 2007; Pandey and Kumari, 2007). Abid and Qaiser (2002) studied cypselar morphology of *Dittrichia* Greuter, *Duhaldea* DC., *Inula* L., *Iphiona* Cass. and *Pentanema* Cass. (Asteraceae) from Pakistan and Kashmir and concluded that two distinct groups of taxa can be recognized. It was confirmed that most of the species of *Anaphalis* DC. (Asteraceae) were delimited due to their distinct micromorphological characters of cypsela (Abid and Qaiser, 2007a). Zhu *et al.* (2006) concluded that achene wall anatomy and surface sculpturing of *Lactuca* L. (Asteraceae) and related genera displayed variation within genera.

Slime cells are widespread in higher plants, especially in fruits and/or seeds in different families, *viz.*, Brassicaceae, Euphorbiaceae, Plantaginaceae, Linaceae, Malvaceae and Lamiaceae (Huang *et al.*, 2000; Western *et al.*, 2000). Slime cells are usually rectangular and they form ladder-like columns that are elongated in a parallel form to the long axis of the achene. Within the Asteraceae, slime cells have been reported in *Achillea* (Kreitschitz and Valles, 2007; Akcin and

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Akcin, 2010), Anthemis L., Chrysanthemum L. (Grubert, 1974) and Artemisia L. (Huang et al., 2000; Yakovleva et al., 2002). It is also suggested that slime plays an important role in controlling of germination, mostly in plants that grow in the condition of water deficiency in arid and semiarid environments (Kreitschitz and Valles, 2007). It can also help fruit or seed dispersal and defence against pathogens (Huang and Gutterman,1999; Huang et al., 2000). Despite several studies on achene micromorphology of Achillea were carried out recently (Abid and Qaiser, 2009; Akcin and Akcin, 2010), however, there is very little information on the Turkish taxa. The main aim of the present work is to study the detailed achene morphology of some Turkish taxa belonging to the genus Achillea and to find out how useful these characters are in the systematics of the genus Achillea.

### **Materials and Methods**

Achenes of *Achillea* L. taxa were studied by stereomicroscope (Leica T1A) and Scanning Electron Microscope (SEM) (JEOL-Neoscope JCM-5000). The studied plant materials were collected from different populations in Turkey. A list of specimens examined is given in Table 1. The specimens have been deposited at the Herbarium of the Department of Biology, University of Ondokuz Mayıs, Turkey (OMUB).

Table 1. List of Achillea L. taxa examined along with their vouchers	Table 1	l. List (	of Achillea L.	. taxa examined	l along with	their vouchers.
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Taxon	Vouchers
Sect. Achillea s. lat.	
1. Achillea biebersteinii Afan.	A5 Amasya: Vicinity of Amasya Education Faculty, road side, 550 m, 11.6.2010, T. Akcin, OMUB 6446
2. A. coarctata Poir.	B5 Kayseri: Kayseri to Avanos, road side, 1100 m, 10.6. 2008, T. Akcin, OMUB 6447
3. A. grandifolia Friv.	A6 Samsun: Kocadağ, 1200 m, 20.7.2008, A. Akcin, OMUB 6448
4. A. millefolium L. subsp. millefolium	A5 Kırşehir: Mucur, 900 m, 21.6.2009, A. Akcin, OMUB 6444
5. A. millefolium L. subsp. pannonica (Scheele) Hayek	A5 Amasya: Yemişen village, 800 m, 28.7.2010, A. Akcin, OMUB 6445
Sect. Babounya (DC.) O. Hoffm.	
6. A. teretifolia Willd. (endemic)	A5 Amasya: Yemişen village, road side, 750 m, 28.7.2010, T. Akçin, OMUB 6442
Sect. Ptarmica (Mill.) W.D.J. Koch	
7. A. biserrata Bieb.	A6 Samsun: Kocadağ, 1200 m, 14.7.2009, T. Akcin, OMUB 6443

The achenes were examined using a stereomicroscope to ensure size, shape, colour and maturity. In order to determine the average achene sizes, 25 mature achenes were measured. For SEM observations, the mature achenes were placed on stubs using double-sided adhesive tape. Following that, they were observed and photographed with a JEOL-Neoscope JCM-5000 scanning electron microscope. The terminology of achene surface patterns adopted is mainly from Barthlott (1981, 1984), Barthlott *et al.* (1998) and Johnson *et al.* (2004).

#### **Results and Discussion**

In the present study, achene morphology of seven taxa belonging to the genus *Achillea* were reported for the first time. Within the examined taxa, *A. teretifolia* is endemic to Turkey (Ekim *et al.*, 2000). Achene shapes were oblong-lanceolate in *A. biserrata* and *A. teretifolia*, oblong in *A. millefolium* subsp. *millefolium*, *A. millefolium* subsp. *pannonica*, *A. coarctata*, *A. biebersteinii* and *A. grandifolia* (Table 2). When achene width and length were considered, different groups were obtained (P < 0.001, Table 2). The widest ( $0.69\pm0.092$  mm) and longest ( $2.01\pm0.252$  mm) achenes were seen in *A. biserrata* (Table 2). Mature achenes ribbed and glabrous in all studied taxa. Achene colours vary from brown to light brown and yellowish-brown.

The position of carpopodium is almost similar (basal) in all taxa studied. However, carpopodium is with narrow circular ring in *A. teretifolia* (Fig. 1a), *A. millefolium* subsp. *millefolium* (Fig. 1e), *A. millefolium* subsp. *pannonica* (Fig.1h), *A. coarctata* (Fig. 2m), *A. biebersteinii* (Fig. 2j) and *A. grandifolia* (Fig. 2o). On the other hand, *A. biserrata* could be separated from other taxa due to its carpopodium shape (Fig. 1c). Abid and Qaiser (2009) reported that cypselar features may also be utilized for specific delimitation as the species of

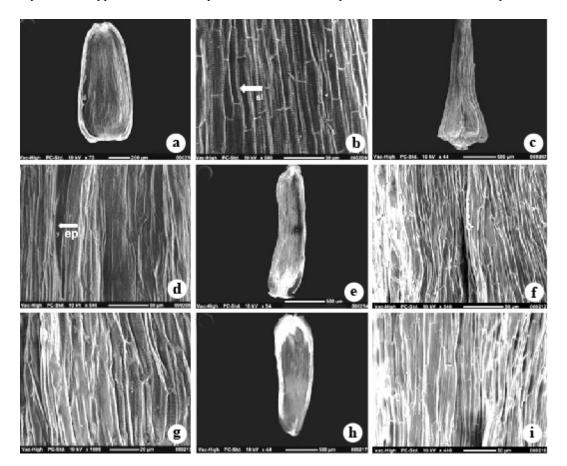


Fig. 1. SEM micrographs of achenes in *Achillea* L. a-b) *A. teretifolia*; c-d) *A. biserrata*; e-g) *A. millefolium* subsp. *millefolium*; h-i) *A. millefolium* subsp. *pannonica*, ep = epidermal cells; sl = slime cells.

			Achene				Carpopodium	
	Shape	Surface	Colour	Width (mm)	Lenght (mm)	Diameter of Carpopodium (µm)	Diameter of Foramen (µm)	Shape
1. Achillea biebersteinii	Oblong	Ribbed, glabrous, slime cells forming	Brown	$0.54 \pm 0.096b$	$0.54 \pm 0.096b  1.24 \pm 0.092cd$	$120.68 \pm 4.088c$	$99.04 \pm 7.778$	With narrow circular ring
2. A.biserrata	Oblong	groups Ribbed, glabrous, slime cells cover	Yellowish- brown	$0.69\pm0.092a\ \ 2.01\pm0.252a$	2.01 ± 0.252a	215.93 ± 41.751a	$108.66 \pm 17.605$	Ring lobed
3. A.coarctata	Oblong	the entire surface Ribbed, glabrous, slime cells cover	Yellowish- brown	$0.54\pm0.048b\ 1.30\pm0.038c$	$1.30\pm0.038c$	$139.07 \pm 7.742c$	$133.81 \pm 27.195$	With narrow circular ring
4. A.grandifolia	Oblong- lanceolate	the entire surface Indefinite ribbed, glabrous, slime cells alternate with	Light brown	$0.30 \pm 0.018c$ 1.12 $\pm 0.058d$	$1.12 \pm 0.058d$	$215.91 \pm 9.703a$	$96.40 \pm 9.713$	With narrow circular ring
5. A. millefolium subsp. millefolium	Oblong	the epidermal cells Ribbed, glabrous, slime cells alternate	Light brown	$0.33\pm 0.045c\ 1.63\pm 0.077b$	$1.63 \pm 0.077b$	$170.13 \pm 11.783b$	$119.54 \pm 4.590$	With narrow circular ring
6. A. millefolium subsp. pannonica	Oblong	with the epidermatic cells Ribbed, glabrous, slime cells alternate with the epidermal	Light brown	$0.53 \pm 0.048b$ $1.74 \pm 0.089b$	$1.74 \pm 0.089b$	196.35 ±1 8.799a	122.49 ± 24.227	With narrow circular ring
7. A. teretifolia	Oblong- lanceolate	cells Ribbed, glabrous, slime cells alternate with the epidermal	Light brown	$0.54 \pm 0.024b$ $1.37 \pm 0.063c$	$1.37 \pm 0.063c$	134.57 ± 12.769b	<b>86.27 ± 11.000</b>	With narrow circular ring

Table 2. Achene features of *Achillea* taxa [means in the column the same letter are not significantly different using Tukey's test (P < 0.001), data are given as mean  $\pm$  soll.

Achillea are grouped on the basis of ribbed and non-ribbed cypsela. A. millefolium was observed to have 10-12 ribbed yellowish-brown cypsela, while non-ribbed cypselas are characteristics for A. wilhemsii (Abid and Qaiser, 2009). Similarly, A. millefolium as indicated by Abid and Qaiser (2009) seperated from A. wilhemsii by the presence of circular ring shaped carpopodium, while A. wilhemsii has carpopodium with 4-6 lobed ring. The present investigation also support the observation of Abid and Qaiser (2009). The achenes of A. millefolium subsp. millefolium and A. millefolium subsp. pannonica have a circular ring shaped carpopodium and the ribbed surface of achenes (Figs 1 e-i). However, carpopodium diameters of the achenes differ in the taxa (P< 0.001, Table 2). The widest carpopodium diameter was present in A. biserrata (215.93  $\pm$  41.751 µm) followed by A. grandifolia (215.91  $\pm$  9.703 µm), while it was narrow in A. biebersteinii (120.68  $\pm$ 4.088 µm). The diameters of foramen of carpopodium among the studied taxa were not significant (P < 0.001, Table 2).

Slime envelope formation is known in several plant families, including the Asteraceae (Kreitschitz and Valles, 2007). Within the *Achillea* genus, slime has been reported previously (Grubert, 1974; Akçin and Akçin, 2010). Our results confirmed the presence of slime in the investigated *Achillea* taxa. Slime cells are usually rectangular and they form ladder-like columns that are elongated in a parallel form to the long axis of the achene (Figs. 1b, 2k,l,n). Slime cells can either cover almost entire surface of the achene as in *A. biserrata* (Fig. 1d) and *A. coarctata* (Fig. 2n), or alternate with the epidermal cells in *A. millefolium* subsp. *millefolium* (Figs 1f-g), and *A. biebersteinii* (Figs 2k-l). A distinct pattern occurs in *A. biebersteinii* achenes (Figs 2k-l), where slime cells form groups scattered over the achene surface. Slime formation on the fruit and/or seed surface is known to be an ecological adaptation to limited availability of water (Huang and Gutterman, 1999; Huang *et al.*, 2000). Several authors showed that the presence of the slime envelope facilitates the adherence of achenes to the soil surface and makes germination easier (Huang *et al.*, 2000; Kreitschitz and Valles, 2007).

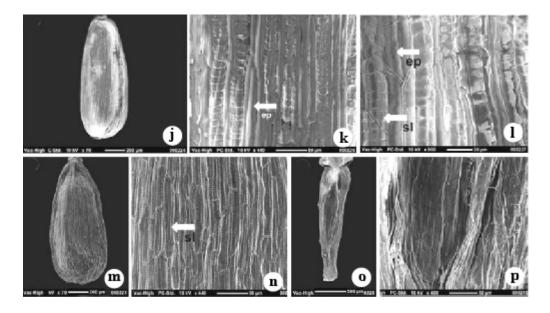


Fig. 2. SEM micrographs of achenes in *Achillea* L. j-l) *A. biebersteinii*; m-n) *A.coarctata*, o-p) *A. grandifolia*, ep = epidermal cells; sl = slime cells.

Taxa such as *A. millefolium* subsp. *millefolium*, *A. millefolium* subsp. *pannonica*, *A. teretifolia*, *A. coarctata*, *A. biebersteinii* occurring in less fertile and often arid location (Huber-Morath, 1975) are characterized by the presence of numerous strands of slime cells. Slime formation also plays an important role in the control of germination, mostly in plants that grow in the condition of water deficiency in arid and semiarid environments, thus faciliating imbibing and maintenance of the water (Kreitschitz and Valles, 2007).

It was also reported that slime cells in species growing in wet environment such as stream sides and meadows may not mature or function properly (Inceer et al., 2012). However, in A. biserrata and A. grandifolia, often growing around wet environment such as coniferous forest (Huber-Morath, 1975), the presence of a slime envelope may play an important role in dispersal of this species. Huang et al. (2000) reported that the slime can reduce the specific weight of diaspores thus facilitating their transportation with water current (hydrochory). In the present study, there are differences in distribution of the slime cells in the examined taxa. Slime cells in A. teretifolia similar to A. coarctata are usually ladder-like coloumns elongated parallelly to long axis of the achene (Figs 1b, 2n). However, A. biebersteinii has a distinct slime cells forming groups scattered over the achene surface (Figs 2k-l). For this reason, these species could be further separated by the sculpture of achene surface. A. grandifolia is separated from the remaining species by having indefinite individual ribs (Fig. 2p). It was also determined that slime cells alternate with epidermal cells in A. millefolium (Figs 1f,g,i), while it covers almost the entire surface of achene in A. biserrata (Fig. 1d) and A. coarctata (Fig. 2n). These results confirmed that slime cells on the fruit surface is associated with ecological adaptation. Akcin and Akcin (2010) reported that achenes of A. phrygia have myxogenic cells (slime-producing) almost throughout the entire surface but slime cells are more restricted in A. gypsicola. Such a relationship was reported for Lamiaceae (Mosquero et al., 2004), Artemisia (Asteraceae) (Kreitschitz and Valles, 2007) and Tripleurospermum (İnceer et al., 2012).

We can conclude that this work helps to characterize the taxa of *Achillea* L. Studies employing additional taxa of the genus are necessary which might further contribute to utilization of achene micromorphological characters as significant attributes in classification of the species.

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