

MOLECULAR CHARACTERIZATION OF *CANNA INDICA* L. BASED ON RANDOM AMPLIFIED POLYMORPHIC DNA MARKERS

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Abstract

Random amplified polymorphic DNA (RAPD) markers were employed for characterization, assessment of genetic variation and inferring relationships among six variants of *Canna indica* L. A total of 198 RAPD bands ranging from 200 bp to 3 kbp were generated by all the six variants. Among them, most of the bands were found to be polymorphic, four bands were unique of which two bands (OPA02₂₀₀₀ and OPA04₃₀₀₀) were observed in the variant 2 (small red) and the other two (OPA01₃₀₀₀ and OPA05₃₀₀₀) were noticed in the variant 4 (orange), and the remaining bands were found to be monomorphic. The pair-wise genetic distance was determined among the six variants that ranged from 0.1446 to 0.6554. A dendrogram was constructed based on the RAPD profiling to infer the relationship among the six variants of *C. indica* that resulted in two major clusters: the first one contained two variants, viz. variant 1 (local red) and variant 2 (small red), while the second cluster composed of the remaining four variants. The results as revealed from the RAPD analysis were found congruent with those of morphological and anatomical investigation of the species.

Introduction

The family Cannaceae, comprising the single genus *Canna* L. is widely distributed throughout the tropical regions. Cannas are worthy garden perennials because of their ornamental value, and the flowering perennial carries an exotic beauty to garden sites with its showy flowers and sometimes with very colourful leaves. In global context, Cannas are one of the popular garden plants, and a large horticultural industry depends on this plant. The commonly cultivated garden Cannas are mostly of hybrid origin, with *Canna indica* as the principal parent (Cronquist, 1981). The genus *Canna* is composed of only 8-10 wild species, and over 1,000 hybrids which are used as garden ornamentals in Europe, North America and many tropical countries (Patra *et al.*, 2008). *Canna* is considered to be native in Mexico, Central America, the Caribbean and tropical South America, West Indies and Central America (Heywood, 1993). In Bangladesh, *Canna* is represented by a single species, *Canna indica*, and is found in almost all over the country as well as planted in many gardens.

The genus *Canna* testifies economical, horticultural and medicinal values. The rhizome of Cannas is rich in starch with multifarious uses in agriculture. Rootstock of *Canna indica* is diaphoretic, diuretic and demulcent, and decoction of root is used in fevers, dropsy and dyspepsia. Seed extract is administered for relieving earache (Ghani, 2003). Young shoots are eaten as green vegetables. The leaves are suitable for wrapping and as plates; both the leaves and the rhizomes

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are used as fodder. Fumigated stems and leaves are used as insecticide (Ong and Siemonsma, 1996). A pure dye is obtained from seeds. Fibre is obtained from the stem which is used as a jute substitute.

RAPDs (Random Amplified Polymorphic DNAs) are widely used molecular markers where DNA fragments are amplified by the Polymerase Chain Reaction (PCR) using short oligonucleotide primers (Williams *et al.*, 1990). RAPD markers are found to be useful in molecular characterization (Islam *et al.*, 2020), DNA fingerprinting (Hossain *et al.*, 2002), assessment of genetic diversity (Karande *et al.*, 2017), cultivar identification (Venkatachalam *et al.*, 2008), taxonomic problems (Vilatersana *et al.*, 2005), systematic relationships (Rahman, 2010), phylogeny reconstruction (Poczail *et al.*, 2008), population genetic structure (Sales *et al.*, 2001), species hybridization (Caraway *et al.*, 2001) and linkage mapping (Atienza *et al.*, 2002).

In spite of economical, horticultural and medicinal value, a very few systematic studies were carried out to detect variation in *Canna indica*. Very recently, Sultana *et al.* (2019) detected six variants of *Canna indica* based on morphological and anatomical investigation. However, molecular studies to assess genetic variation in *Canna indica* are lacking in Bangladesh. Because of universality and reproducibility the RAPD markers were employed to detect genetic diversity of *Canna indica*. The present study aimed at assessing genetic variation and relationships among the six variants of *Canna indica* occurring in Bangladesh for the first time.

Materials and Methods

Plant material:

Plant specimens of *Canna indica* were collected from different parts of Bangladesh and were maintained under the controlled climatic condition, and planted in the Botanical Garden of Jagannath University. These were supplemented by the herbarium specimens examined at the Bangladesh National Herbarium (DACB) and Dhaka University Salarkhan Herbarium (DUSH).

Isolation of Genomic DNA:

DNA was isolated from the leaf tissue ranging from 1.0 to 1.5 g using CTAB method (Doyle and Doyle 1987). The isolated DNA was dissolved in TE buffer and stored at -20°C until further use.

RAPD amplification:

A total of 10 decamer oligonucleotide primers were examined, and based on reproducibility the following five primers were finally chosen for RAPD analysis: OPA01 (5'-CAGGCCCTTC-3'), OPA02 (5'-TGCCGAGCTG-3'), OPA04 (5'-AATCGGGCTG-3'), OPA05 (5'-AGGGG TCTTG-3') and OPA10 (5'-GTGATCGCAG-3'). Each PCR included 2.0 μl of 25ng genomic DNA, 1.0 μl primer, 2.5 μl 10X Taq buffer, 0.5 μl of dNTP mixture, 0.2 μl Taq polymerase enzyme, and 18.8 μl sterile, deionised distilled water up to final volume of 25 μl . PCR reaction was performed in an oil-free thermal cycler (Biometra, UNO II) as per following temperature profile: initial denaturation at 94°C for 5 min, denaturation at 94°C for 45s, annealing at 32°C (for 60% GC rich content primer) and 34°C (for 70% GC rich content primer) for 30s, extension at 72°C for 3 min followed by 55 cycles. A final 7 min extension at 72°C ensured full extension of all amplified fragments.

Gel electrophoresis:

Amplified PCR products were separated on 1% agarose gel and stained with ethidium bromide solution. The size of the amplicons was determined using standard 1Kb ladder. DNA bands were visualized under UV-transilluminator and photographed.

Data analysis:

RAPD bands were recorded in a binary data matrix scored as presence (1) or absence (0) for each sample. Similarity Matrix coefficient was used for measuring genetic relationship among the variants analyzed. UPGMA (Unweighted pair group method with arithmetic average) tree was generated by clustering the distance matrix. Data were analyzed using POPGENE32 (Nei, 1972).

Results and Discussion

RAPD fingerprints:

The present study revealed a total of 198 RAPD fingerprints generated by five oligonucleotide primers ranging from 200 bp to 3 kbp in six variants of *Canna indica*. The studied five primers generated reproducible bands in all the variants investigated. The number of bands generated by the primers varied within the investigated variants and showed polymorphisms among them.

The primer OPA01 (5'-CAGGCCCTTC-3') generated a total of 50 RAPD bands in the six variants of *Canna indica*. All the variants presented 4 monomorphic bands each at the same locus (OPA01₂₅₀₀, OPA01₁₅₅₀, OPA01₉₀₀ and OPA01₇₅₀). The highest number of bands (11) was generated by the variant 6 (yellow with red spots) of which 4 bands (36.36%) were found to be monomorphic and 7 bands (63.64%) were polymorphic. The variant 1 (local red) produced the lowest number of bands (6) showing 85.71% similarities with the variant 2 (small red), and among the 6 bands produced, 4 were monomorphic, while the other 2 were polymorphic. The variant 2 produced 7 bands of which 4 were monomorphic and 3 were polymorphic. The variant 3 (pink) and variant 4 (orange) produced 9 bands each, among them 4 bands were monomorphic and 5 were polymorphic. The variant 5 (yellow) produced 8 RAPD bands at the same locus position of the variant 6 and showed 72.72% similarities with it. Out of 9 bands generated by the variant 4 one unique band was detected by the primer OPA01₃₀₀₀ in this variant (Fig. 1a).

The primer OPA02 (5'-TGCCGAGCT G-3') generated the highest number of bands (62) in all the six variants. Among all the variants, the highest number of bands (12) was observed in the variant 6, of which 7 bands (63.64%) were monomorphic and 4 (36.36%) were polymorphic. The lowest number of bands (9) was found in the variant 1, and among them 7 bands (77.78%) were monomorphic and 2 (22.22%) were polymorphic. The variants 2, 3 and 4 generated 10 bands each, among them the variants 3 and 4 showed the same banding pattern (100% similarities), whereas, the variant 2 displayed 80% similarities with the variant 4. In the variant 5, a total of 11 bands were found of which 7 bands were monomorphic and 4 polymorphic. The variant 5 showed 91.67% affinity with the variant 6. One unique band was found in the variant 2 at the OPA02₂₀₀₀ position (Fig. 1b).

The OPA04 (5'-AATCGGGCTG-3') primer displayed a total of 38 bands among the six variants. The highest number of bands (11) was observed in the variant 2 of which 10 bands (90.91%) were polymorphic and 1 was monomorphic. The lowest number of bands (3) was found in the variant 6 and among them 2 bands (66.67%) were polymorphic and 1 was monomorphic. The variant 1 presented 9 bands of which 8 bands (88.89%) were polymorphic and 1 was monomorphic. The variant 3 displayed 7 bands and among them 6 bands (85.71%) were polymorphic and 1 was found to be common. The variant 4 and the variant 5 both exhibited 4 bands, of which 1 was monomorphic. No unique polymorphic band was generated by the primer OPA04 (Fig. 1c).

The primer OPA05 (5'-AGGGGTCTTG-3') produced a total of 16 bands in six variants. The highest number of bands (7) was found in the variant 4, among them 1 (OPA05₃₀₀₀) was unique band and 6 were polymorphic (85.71%). No bands were found in the variant 6. The variants 3 and

5 produced only 1 polymorphic band at the same position (OPA05₂₅₀₀). The variant 1 showed 5 bands, and all of them were found to be polymorphic and showed 71.43% similarities with the variant 4. Two polymorphic bands were observed in the variant 2.

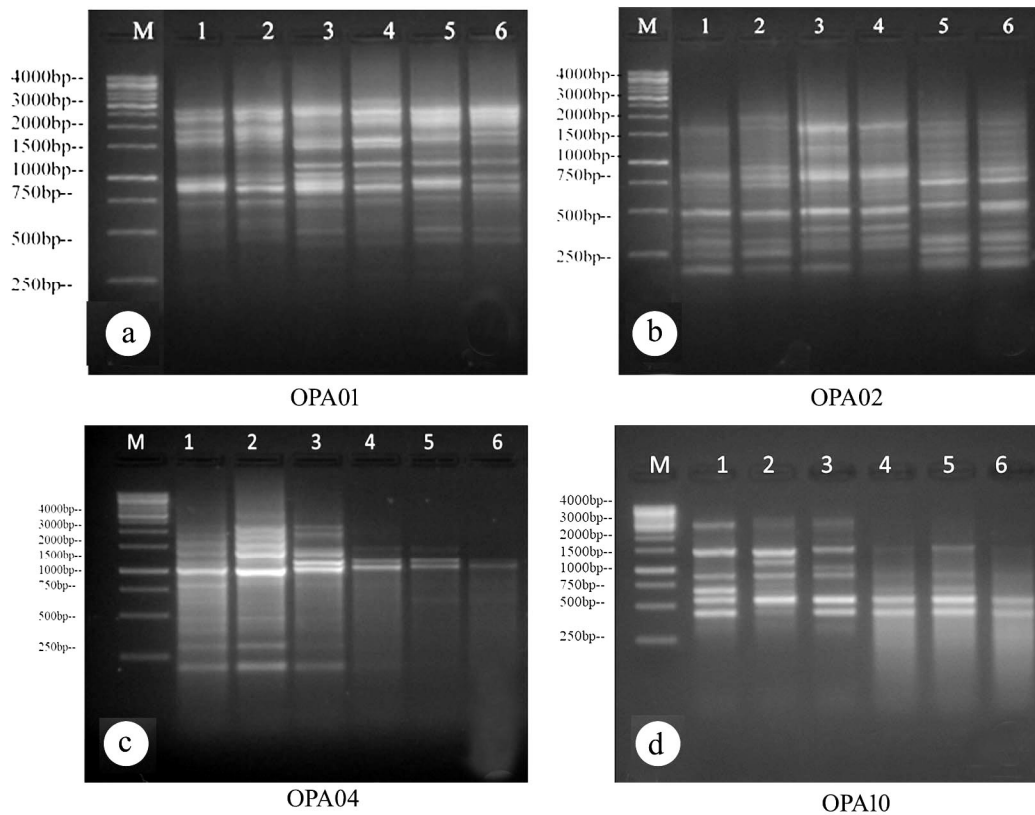


Fig. 1. RAPD fingerprints in six variants of *Canna indica* L.: a. OPA01; b. OPA02; c. OPA04; d. OPA10. M: Molecular marker (1Kb); 1. Local red; 2. Small red; 3. Pink; 4. Orange; 5. Yellow; 6. Yellow with red spots.

The primer OPA10 (5'-GTGATCGCAG-3') generated 32 bands in all the six variants. The highest number of bands (6) was found in the variants 1, 2 and 3, among which 4 bands were found to be common (66.67%) and 2 were polymorphic. The similar banding pattern was identified in the variant 2 and variant 3. The variants 4 and 5 produced 5 bands each, and among them 4 were common and 1 was polymorphic (OPA10₆₀₀). The variant 6 presented 4 bands and all of them were found to be monomorphic. No unique band was observed in any of the variants (Fig. 1d).

RAPD polymorphism, genetic diversity and molecular relationships

The present study demonstrated a total of 198 bands generated by 5 primers in all the six variants of *Canna indica* with an average of 39.6 RAPD loci per primer. The highest polymorphism (55.56%) was detected in the variant 2, while the lowest (44.83%) was found in the

variant 5. The other variants showed relatively high level of polymorphism (Table 1). The average polymorphism was found to be 51.19%.

Table 1. RAPD fingerprints and polymorphism in six variants of *Canna indica*.

Name of the variants	Total no. of bands	No. of polymorphic bands	% of polymorphism	Average % of polymorphism
1. <i>Canna indica</i> (local red)	35	19	54.29	
2. <i>C. indica</i> (small red)	36	20	55.56	
3. <i>C. indica</i> . (pink)	33	17	51.53	
4. <i>C. indica</i> (orange)	35	19	54.29	51.19
5. <i>C. indica</i> (yellow)	29	13	44.83	
6. <i>C. indica</i> L (yellow with red spots)	30	14	46.67	

The highly reproducible bands ranging from 200 to 3000 bp were scored for assessment of genetic variation among the six variants of *Canna indica*. Among the five oligonucleotide primers employed in the present study, 50 bands were generated by the primer OPA01, 62 by OPA02, 38 by OPA04, 16 by OPA05 and 32 bands by OPA10 primer. The highest genetic distance (0.6554) was found between the variants 1 and 6, and between the variants 2 and 4 followed by the genetic distance as observed between the variants 2 and 6 (0.6190). The same genetic distance (0.4249) was found between the variants 1 and 4, and 4 and 5. Among all the variants, the lowest genetic distance (0.1446) was observed between the variants 5 and 6 indicating a close affinity between these two variants (Table 2).

Table 2. Genetic distance among six variants of *Canna indica*.

Name of the variants	Var.1- local red	Var. 2- small red	Var. 3- pink	Var. 4- orange	Var. 5- Yellow	Var.6 -yellow with red spot
Var. 1- local red	0					
Var. 2 - small red	0.2377	0				
Var. 3 - pink	0.4855	0.3403	0			
Var. 4 - orange	0.4249	0.6554	0.4855	0		
Var. 5 - yellow	0.5500	0.5839	0.3677	0.4249	0	
Var. 6 - yellow with red spot	0.6554	0.6190	0.3403	0.3959	0.1446	0

UPGMA tree constructed from RAPD fingerprints showed the inter-relationships among the six variants of *Canna indica* (Fig. 2). The UPGMA analysis in *C. indica* resulted in two major clusters: cluster 1 contained two variants, viz. variant 1 (local red) and variant 2 (small red), while the cluster 2 composed of the remaining four variants, viz. variant 3 (pink), variant 5 (yellow), variant 6 (yellow with red spot) and variant 4 (orange).

In the present investigation, RAPD markers were employed to characterize the six variants of *Canna indica* alongside with assessments of genetic variation and to infer molecular relationships among the variants. Out of 10 RAPD primers investigated, 5 showed significant amplifications in PCR analysis and altogether produced 198 bands, and the size of the bands ranged from 200 bp to 3.0 kbp. Considerable genetic variability existed in the variants of *Canna indica*. The present

study revealed that the variants 5 and 6 joined together indicating a close relationships between them, and genetic distance between these two variants was found to be 0.1446 (Fig. 2, Table 2). The study also exposed a close affinity between the variants 1 and 2 where the genetic distance was reported to be 0.2377 (Table 2). The variant 4 was found to be distantly related with other variants. Out of 198 bands generated by five primers in six variants of *Canna indica*, 102 bands were found to be polymorphic and 96 as monomorphic.

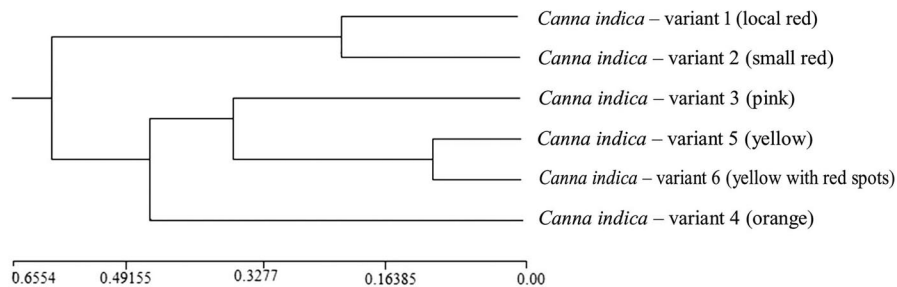


Fig. 2. UPGMA dendrogram showing genetic relationships among the six variants of *Canna indica* as revealed by RAPD markers.

The study exhibited four unique bands, *viz.* OPA02₂₀₀₀ and OPA04₃₀₀₀ as found in the variant 2 and OPA01₃₀₀₀ and OPA05₃₀₀₀ as noticed in the variant 4. The present investigation demonstrated that the average polymorphism within all the variants was found to be 51.195%. Results obtained from the present study were found to be consistent with those of morphological and anatomical investigation among these six variants of *Canna indica* (Sultana *et al.*, 2019). In order to have better understanding on genetic variation and molecular relationships large number of taxa should be employed with additional molecular markers, such as AFLP, ISSR and microsatellites which will through more light on the phylogeny of *Canna indica*.

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