

Available online at www.banglajol.info

Bangladesh J. Sci. Ind. Res. 44(2), 215-220, 2009

BANGLADESH JOURNAL OF SCIENTIFIC AND INDUSTRIAL RESEARCH

E-mail: bjsir07@gmail.com

## Study of Mutagenic Effect of Coccinea cordifolia (Linn.) Cogn. Leaf Extract on Neurospora crassa Fungus

### M. Nazrul Islam Bhuiyan<sup>a\*</sup>, T. I. M. A.Mozmader<sup>b</sup>, M. Amirul Hoque<sup>c</sup>, Sumi Akter<sup>d</sup> and M. Rafiqual Islam<sup>e</sup>

<sup>a</sup>BCSIR Laboratories, Chittagong-4220, <sup>b</sup>Department of Botany, University of Dhaka, Dhaka-1000, <sup>c</sup>BCSIR Laboratories, Dhaka-1205, <sup>d</sup>.icddr, b, Dhaka and <sup>e</sup>Asiatic Society of Bangladesh.

### Abstract

*Coccinea cordifolia* (Linn.) Cogn. leaf extract showed significant mutagenic effect on *Neurospora crassa fungus*. Observation after 24 hours, control showed 2.9 cm linear vegetative growth of the mycelia where *C. cordifolia leaf* with *N. crassa* showed 0.25 cm growth and it is 12 times less than the control. Besides, 0.5, 1.0, 1.5, 2.0, 3.0 and 4.0 ml extract showed gradual decrease of growth of mycelia. This result indicates that this plant was suitable for induction of mutation in *N. crassa*. After mutation of *N. crassa* Ema (5297) conidia with *C. cordifolia* leaf extract, 6 types of mutants were found namely- *dirty, ropy, albino, mat, cauliflower, conidial brand. Albino* mutant showed highest frequency (35%) where *ropy* showed lowest frequency (5%).

Key words: Neurospora crassa, Coccinea cordifolia, Mutants, Dirty, Ropy, Albino, Mat, Cauliflower and Conidial brand.

### Introduction

*Neurospora crassa* is a type of red bread mold of the phylum Ascomycota. The genus name, meaning, "nerve spore" refers to the characteristic striations on the spores that resemble axons. *N. crassa* is used as a model organism because it is easy to grow and has a haploid life cycle that makes genetic analysis simple since recessive traits will show up in the off-spring.

Neurospora was used by Beadle and Tatum (1941) in their experiments for which they won the Nobel Prize in Physiology or Medicine in 1958. Beadle and Tatum (1941) exposed N. crassa to x-rays, causing mutations. Their extremely short life cycle and capacity to grow in minimal medium help the scientists of this branch to perform extensive work within a period of short time, which is one of the most essential factors of genitival study (Watson, 1970). Beadle and Tatum (1941) first used the N. crassa in genitival and at the same time for biochemical study. N. crassa was originally selected for studying biochemical genetics because it is an organism in which both formal genetic analysis and investigation of nutritional mutants appeared possible, (Beadle and Tatum, 1941). Plant extracts play an important role to check the growth of various fungi. Scientists are interested in evaluating the antifungal activities of plant extracts against plant pathogenic fungi (Ahmed and Sultana, 1984; Bashar and Rai, 1991; Anwar et. al., 1994). Haque and Shamsi (1996) observed that the leaf

\* Author for correspondence: Nazrul119@yahoo.com

extracts of Neem (Azadirachta indica) have antifungal properties and it decreased the radial growth of fungus but none of them studied the mutagenic effect of the plant extracts. By the application of artificial mutation many mutants can be obtained which fail to grow unless some specific substances, such as amino acid or vitamin are added to the culture medium. The actual mutation is supposed to take place in the level of DNA structure. Perhaps Westergaard and Mitchel (1947) has made the most extensive and systemic surveys of the action of biochemical mutagens on fungi on N. crassa (Fincham et al., 1979). Therefore, a plan was made to induce mutation morphological change by using leaf extracts of C. cordifolia. Few selected and interesting mutants were studied for there morphologically changes by studying the linear growth, mycelial weight, time required for germination, fertility, mating type, heterocaryosis in comparison with the wild type Ema. Chemical mutagens have the ability to penetrate the cells and to change the DNA. Presently, scientists are interested to evaluate the mutagenic properties of chemicals on N. crassa. So the present study was undertaken to treat C. cordifolia leaf aqueous extract by N. crassa to produce morphological mutants. If morphological mutants formed, then all of them were isolated and characterized separately to determine the mutagenic and growth inhibitory effects of C. cordifolia present in the leaves.

### **Materials and Methods**

*N*.*crassa* Ema (5297) was the experimental material. The wild type strain was received from Fungal Genetic Stock Centre, Department of Microbiology; University of Kansas Medical School, Kansas, U.S.A. Strain Ema (5297) was used. Vogel's minimal medium (VM) (Vogel, 1956) was used for maintaining the cultures. Solid VM was used for obtaining and measuring linear growth of conidia (Ryan *et. al.*, 1943). Different concentrations of aqueous extracts of the leaves of *C. cordifolia* were used in the experiments. The extraction procedure is given below:

#### **Aqueous extract**

Mature fresh green leaves of *C. cordifolia* was washed with sterilized distilled water and then air dried and 100 g of clean leaves was ground with mortar and pestle. The paste was filtered to get extracts. The filtered extract was centrifuged for 5 minutes in 300 rpm at  $25^{\circ}$  C. The supernatant was used for this experiment.

# Preparation of solution at different concentration of *C. cordifolia*

Experiments were made to find out the mutagenic effect of C. cordifolia extracts on N. crassa. Leaf extracts of different density were prepared with the help of sterilized distilled water. Leaves of different weights were taken separately, washed 3-4 times with tap water and 3 times with distilled water. After air-drying, the leaves were ground in an electric grinder with distilled water and without distilled water. The paste was filtered with a sterilized cotton cloth and finally filtered with filtre paper to get clean solution. Different extracts of known strengths were prepared using leaves and sterilized distilled water. 20 g leaves were pasted without distilled water and it was as  $N_{\text{P}}$ . The next solution of 20 g leaves and 10 ml distilled water (N1), 10 g leaves and 10 ml distilled water (N2), 10 g leaves and 18 ml distilled water (N<sub>3</sub>), 10 g leaves and 20 ml distilled water (N<sub>4</sub>), and 10 ml distilled water and 4 g leaves (N<sub>5</sub>) were prepared which were denoted as N1, N2, N3, N4 and N5 (Table I). All extracts namely Np, N1, N2, N3, N4 and N5 were preserved separately in sterilized test tubes in a refrigerator. Some of the extracts were also sterilized in autoclave.

#### Preparation of different extracts of *C. cordifolia* leaves:

Different concentrations of the leaf extracts of *C. cordifolia* with sterilized distilled water were prepared.

Amount of leaf	Amount of
	sterilized distilled water
20g	-
20g	10ml
10g	10ml
10g	18ml
10g	20ml
4 g	10ml
	20g 20g 10g 10g 10g

#### Table I. Different doses of C. cordifolia Solution

# Effect of water extracts of *C. cordifolia* on the growth of *Neurospora crassa*

For testing the effect of water extracts of *C. cordifolia* on the growth on *N. crassa*, different concentrations of it were prepared. Different solutions were taken separately on the petri dishes at the rate of 1 drop, 2 drops, 3 drops, 4 drops, 8 drops, 0.5 ml, 1ml, 1.5ml, 2ml, 3 ml and 4ml. Ten ml of molten SM medium was added in each Petri dish and rotated gently for uniform mixing of solution with the medium. When the medium became solid, the centre of the Petri dish was marked and fresh culture of Ema was inoculated at that point with a sterilized needle. All the Petri dishes were kept in an incubator at  $25^{\circ}$ C after 16, 24, 40 and 48 hours, the radial growth of Ema of each petri dish was measured in cm.

# Treating of Ema with desired extracts of *C. cordifolia* leaves

#### a) Obtaining fresh culture

To obtain fresh culture, Ema was cultured 4 times at 4 days intervals in each case. Five days old culture was used for treating conidia.

#### b) Sterilization

All the media, essential elements and instruments were sterilized in an autoclave at 120°C under 15 Ib pressure for 20 minutes. The inoculation chamber, needle, centrifuge machine etc. were also sterilized with rectified spirit.

#### c) Centrifugation

10 ml *C. cordifolia* leaf extracts of the concentration  $N_P$ ,  $N_1$ ,  $N_2$ ,  $N_5$  were taken into 4 centrifuge tubes. One loop of conidia (about 10,000 conidia) of Ema was taken into each tube and was shaken for homogenous solution. The solution was centrifuged with the help of a centrifuge machine for 20 minutes in 300 rpm at 25<sup>o</sup> C.

#### d) Filtration

After centrifugation, the solution above conidia was poured out from the centrifuge tube. 10 ml of sterilized distilled water was added to the centrifuge tube and centrifuged for 3 minutes. Then the distilled water was poured out: The same procedure was repeated twice.

#### e) Preparation of suspension with treated conidia

10 ml of distilled water was added to the treated conidia remaining at the bottom of the centrifuge tube and the tube was shaken well.

#### f) Plating of treated conidia

The sterilized Petri dishes were marked as  $N_P$ ,  $N_1$ ,  $N_2$ ,  $N_5$ and 1 drop of the suspension of each was taken. Accordingly, 10 ml of molten SM medium (Sorbose and Infanger, 1964) was added to a Petri dish and were shaken gently to mix with the suspension and media. The plates were kept inside the incubator at 25°C for maximum growth of conidia.

#### g) Isolation of single conidial colony

A number of well-separated colonies were isolated by cutting agar blocks from the conidial colony with an arrow shaped isolating needle and were inoculated into small tubes containing VM media (Vogel, 1956). Precautions were taken so that separated growing conidial colonies can be isolated only.

#### h) Classification

After 5 days, all the cultures were observed and classified by comparing their characters with wild type Ema (Table II). The conidial cultures with any morphological variation were subcultured several times in small tubes and checked carefully whether any permanent morphological change occurs.

# Growth test of different morphological mutants on SM, VM (sucrose) and VM (glucose)

Growth tests were made in sterilized Petri dishes containing sorbose minimal medium, Vogel's minimal medium (sucrose) and vogel's minimal medium (glucose was used in lieu of sucrose). The plates were divided into 24 compartments by glass marking pencil. Then conidia from a particular mutant were put in one compartment, 23 compartments contained conidia from different mutants and one contained wild type (Ema) as control.

# Study of mutagenic effect of extracts of *C. cordifolia* on *Neurospora crassa*

Mature fresh green leaves of C. cordifolia were washed with sterilized distilled water and then air-dried. 50 gm of clean leaves were ground with mortar and pestle. The paste was filtered through extracts. The filtered extract was centrifuged for 5 minutes. The supernatant was used for this experiment. 6 sets of experiments were set taking 0.5 ml, 1 ml, 1.5 ml, 2 ml, 3 ml and 4 ml of extracts. Conidia of Ema (5297) of N. crassa were treated for 3-4 hours in 100% concentration of the extracts. It was centrifuged and the supernatant was discarded, treated conidia were washed twice with sterilized distilled water by pouring 1 ml sterilized distilled water in each centrifuge tube. Final suspension was made with sterilized distilled water. Then, 1 drop, 2 drops, 3 drops, 4 drops and 8 drops of conidial suspensions were taken in each of the 30 Petri dishes {Table III (i) and (ii)} and radial growths of Ema on VM containing C. cordifolia extracts were observed. Vogel's minimal medium (VM) was poured in each of the 30 Petri dishes. Plates were incubated for the formation of conidial colonies for 3 days. Observation was made daily for the appearance of the colony. The colonies were isolated in the test tubes containing Vogel's minimal medium (VM) and the isolates were incubated for growth at 25°C. After 4 days, the

Table II.	Characteristics of	f the mutants of N.	crassa obtained	by t	the induction	with C.	cordifolia	(Different dose	s).
-----------	--------------------	---------------------	-----------------	------	---------------	---------	------------	-----------------	-----

Name of	Percent of	Characteristics of the mutants
the mutants	mutants	
Ropy	5	The mycelia look like beautiful ropes, conidia pinkish a orange in colour.
		Growth is less than wild type.
Albino	35	Less growth of mycelia, conidia are very scanty in number.
		Mycelia and conidia are completely colourless.
Mat	30	Mycelia have a characteristic frayed appearance like mat
Dirty	10	Small conidial lump scattered here and there in the tube.
Conidial brand.	12	Dense conidial growths form a band shaped structure at the top.
Cauliflower	8	It produces cauliflower-like buttons of growth.

isolates were examined and classified (Table II). Different concentrations of *C. cordifolia* extracts were used to test the mutagenic effect on *N. crassa*. Fixed concentrations had appropriate mutagenic effects on *N. crassa*.

#### **Results and Discussion**

C. cordifolia leaf extract effects of chemical mutagens on frequency and specificity of chemical mutations in N. crassa. Suspensions of N. crassa conidia were inactivated by C. cordifolia leaf extract. Chemical substances, which showed mutagenic activities, became longer in every year. The authors tested the mutagenic effect of leaf extracts of C. cordifolia on the fungus, N. crassa. The plant aerial parts contain protein, fat, vitamin C, sterols, ß-sitosterol, phenolic compounds, triterpenoids, bitter glycosidic constituents, alkaloids, cephalandrine A and B, alcohol, cephalandrol, tritriacontane, heptacosane. (Ghani, 1998). To determine the efficacy of C. cordifolia on the radial growth of N. crassa different doses of leaf extracts (sterilized and non-sterilized) were used. It was evident from Table III (i, ii) that the radial growth of N. crassa was proportional to the doses of C. cordifolia extracts used. By increasing the doses of leaf extracts of C. cordifolia radial growth of Ema was reduced in comparison to control. Np doses were found to be more effective than  $N_1$ ,  $N_2$ ,  $N_3$  and  $N_4$  doses. An extensive study was conducted on different doses, so that *N. crassa* may tolerate little checked growth and to determine the extract amount to kill the fungus *N. crassa*.

From the Table III (i and ii) it was noted that 4 ml of doses decreased the radial growth extensively and the colony became very compressed and checked as compared to the control and 4 ml of dose killed the fungus N. crassa. There was no notable difference between sterilized and non-sterilized doses on the growth of *N. crassa* {Table III (i and ii)}. Aqueous extracts of the leaves of C. cordifolia showed significant inhibition of growth and mutagenesis on N. crassa. Result showed that 1 ml extract reacted with the test organism of N. crassa effectively. During the period of 24 hours, it produced only 2.9 cm linear vegetative growth of the mycelia was observed. The linear growth decreased with the increase of the doses of the extracts. Conidia of Ema (5297) of N. crassa were treated for 3-4 hours in different concentrations of the aqueous leaf extracts. One hundred percent extracts produced seven types of mutants, namely- dirty, ropy, albino, mat, cauliflower and conidial brand (Table II and Fig.1) and albino showed highest frequency (35%)

Table III. (i) Effect of C. cordifolia extracts on the radial growth of N. crassa Ema

Doses of the solution										cm				
			0 drop (Control)	1 Drop	2 drops	3 drops	4 drops	8 drops	0.5 ml.	0.1 ml.	1.5 ml.	2 ml.	3 ml.	4
		16	4.80	3.80	2.90	1.90	1.70	1.60	4.20	4.00	3.50	2.90	2.30	0
		24	4.70	3.80	2.90	2.60	2.30	1.80	2.90	2.50	2.10	1.90	0.80	0
10 g leaves +	Sterilized	40	over	5.10	5.00	4.70	3.80	3.90	3.70	2.60	1.40	0.90	0.30	0
20 ml distilled		48	over	over	5.20	5.00	4.90	4.50	3.90	3.80	3.50	3.50	0.50	0
water (N <sub>4</sub> )		16	over	1.90	1.80	1.70	1.60	1.40	4.45	3.20	2.10	1.90	0.60	0
water (N <sub>4</sub> )	Unsterilized	24	over	over	4.20	4.10	3.10	2.80	2.90	2.20	1.50	1.20	0.70	0
		40	over	5.00	4.90	4.00	3.80	3.70	3.60	3.20	3.00	2.60	0.40	0.10
		48	over	over	5.10	5.00	4.50	4.10	3.10	2.90	1.70	1.60	0.50	0.20
		16	4.90	4.70	3.80	3.30	2.80	2.20	5.00	4.90	4.10	3.90	0.20	0
		24	4.70	4.50	3.90	3.10	2.50	1.80	2.90	2.60	2.20	1.10	0.90	0
	Sterilized _	40	over	5.20	5.10	5.00	4.90	4.60	3.40	3.30	3.20	2.90	0.50	0
10 g lagyag I		48	over	over	5.20	5.30	5.20	5.00	4.90	3.80	3.50	3.10	0.60	0
10 g leaves + 20 ml distilled		16	3.70	3.60	3.50	2.40	2.10	1.90	4.90	4.70	3.60	2.40	0.80	0
water $(N_3)$		24	4.50	3.90	3.60	2.60	2.20	1.80	2.90	2.20	2.10	2.00	0.90	0
		40	over	over	5.10	5.00	4.80	3.70	3.40	3.30	3.20	2.90	1.70	0.10
	Unsterilized	48	over	over	over	5.10	5.20	4.90	4.70	4.60	3.90	2.50	1.90	0.20

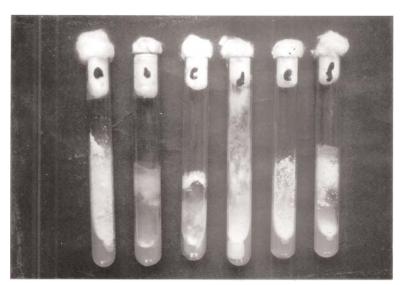


Fig. 1. Growth pattern of different mutants of N. crassa with wild type Ema.

Doses of the solution	Treatment time (hr.)		Amount of solution taken in a petri dish and growth obtained in cm										
		0 drop (Control)	1 Drop	2 drops	3 drops	4 drops	8 drops	0.5 ml.	0.1 ml.	1.5 ml.	2 ml.	3 ml.	4
10 g leaves +	16	2.30	1.90	1.80	1.40	1.30	1.20	2.90	2.70	2.50	1.30	0.10	0
10 g leaves + 10 ml distilled water $(N_2)$	24	2.50	2.40	2.35	2.30	2.20	1.90	2.90	2.80	2.40	2.30	1.20	0
	40	4.00	3.50	3.40	3.30	3.20	3.00	2.90	2.70	2.20	1.50	0.30	0
	48	over	4.20	3.90	3.80	3.70	3.50	3.40	3.10	2.80	1.70	0.40	0
20 g leaves +	16	3.60	2.20	1.90	1.80	1.60	1.10	3.20	3.00	2.80	1.00	0.40	0
10 ml distilled	24	4.50	4.10	3.40	2.60	2.20	1.90	2.90	2.70	2.50	2.10	1.50	0
water $(N_1)$	40	over	4.70	3.90	3.80	3.70	3.10	2.90	2.60	2.30	1.20	1.00	0
	48	over	over	4.90	3.70	3.60	3.50	3.10	2.80	2.30	1.30	1.10	0
20g leaves	16	4.90	4.70	4.20	3.50	3.30	3.20	3.40	3.10	2.60	1.30	1.20	0
(N <sub>p</sub> )	24	4.80	4.50	4.30	3.70	3.50	3.30	2.90	2.60	2.40	2.10	1.20	0
	40	over	5.10	4.80	4.20	4.10	3.90	3.80	3.50	2.40	1.60	1.40	0.20
	48	over	over	5.00	4.70	4.40	3.50	4.10	3.60	3.10	1.90	1.60	0.30

Table III. (ii) Contd C. cordifolia extracts on the radial growth of N. crassa Ema.

where *ropy* showed lowest frequency (5%). Others showed *dirty* (10%), *mat* (30%), *cauliflower* (8%) and *conidial brand* (12%). The types and frequency of mutants obtained with the leaf extracts of *C. cordifolia* were slightly different from that recorded earlier with leaf extracts of *Averrhoa carambola* (Bhuiyan *et al.*, 2007), *Azadirichta indica* (Keya, 1998), bulb extract of *Allium sativum* (Yesmin, 1998), leaf extracts of *Zingiber officinale* and *Andrographis paniculata* (Bhuiyan, 2003). So, we concluded that from leaf extracts of

*C. cordifolia* we found total six different types of mutants (*dirty, ropy, albino, mat, cauliflower* and *conidial brand*).

#### Acknowledgement

The authors gratefully acknowledge the receipt of financial grant received from the Ministry of Science & Information and Communication Technology, People's Republic of Bangladesh, for carrying out the present research work.

### References

- Ahmed N., and Sultana K. (1984) Fungitoxic effect of garlic on treatment of jute seed. *Bangladesh J. Bot.* **13**: 130-136.
- Anwar M. N., Singha P., Begum J., and Chowdhury J. U. (1994) Antifungal activities of some selected plant extracts on phytopathogenic fungi. *Bangladesh J. of life science*. 6: 23-26.
- Bashar, M. A. Bharat Rai. (1991) Antifungal activity of some plant extracts against *Fusarium oexosporium* f. sp. *ciccri. Bangladesh J. Bot.* **20** (2): 219-222.
- Beadle G. W., and Tatum E. L. (1941) Genetic control of biochemical reactions in *Neurospora* Proc. *Natl. Acad. Sci.*, 27: 499-506.
- Bhuiyan M. N. I. (2003) Induction of mutation in *Neurospora crassa* and characterization, genetical studies of some selected mutants. MS Thesis, Department of Botany, University of Dhaka, Pp. 25-63.
- Bhuiyan M. N. I., Mozmader T. I. M. A., and Rahman S. (2007) Microbial antagonism and induction of mutation in *Neurospora crassa* by crude leaves extract of *Averrhoa carambola.L."Bangladesh J. Sci. Ind. Res.* 42(2):503-508.
- Fincham J. R. S., Day P. R., and Radford A. (1979) Fungal. Genetics, 4th Ed., Blackwell. Scientific. Publications, Oxford, pp. 254-281.
- Ghani A. (1998) Medicinal Plants of Bangladesh: Chemical constituents and uses. Asiatic Society of Bangladesh, Old Nimtali, Dhaka. Pp. 136.

- Haque T., and Shamsi S. (1996) Activity of certain plant extracts against jute stem rot fungus *Macrophomina phaseolina*. *Dhaka Univ. J. Biol. Sci.* **5** (1):103-104.
- Keya S. U. (1998) Induction of mutants in *Neurospora* crassa with leaf extract of Neem (*Azadirachta indica A. Juss*), Diathane- M45, UV, EMS and their genetical studies M. Sc. Thesis, Department of Botany, University of Dhaka.Pp. 45-73.
- Ryan F. J., Beadle G. W., and Tatum E. L. (1943) The tube method of measuring the growth rate of *Neurospora crassa. Am. J. Bot.* **30:** 784-799.
- Sorbose A. M., and Infanger A. M. (1964) Formation of heterocaryonsby fusion of isolated hyphal tips on solid medium in petriplates *Neurospora*. *Newsl.* 6: 26.
- Vogel J. J. (1956) A convenient growth medium for Neurospora, Microb. Genet. Bull 13: 42-43.
- Watson J. D. (1970) Molecular Biology of the gene. 2nd ed. Benjamin, INC. Mento park, California, Pp. 2.
- Westergaard M., and Mitchel H. K. (1947) Neurospora crassa, V.a. Synthetic medium for favouring sexual reproduction. Am. J. Bot. 34: 573-574.
- Yesmin N. (1998) Induction of mutation in *Neurospora* crassa, characterization genetical and chromatographic studies of some selected mutants. M. Sc. Thesis, Department of Botany, University of Dhaka, Pp. 32-67.

Received : April, 24, 2008; Accepted : November 06, 2008