Bangladesh J. Sci. Ind. Res. 42(3), 269-280, 2007

# Some Nutritional Composition and Functional Properties of *Prosopis Africana*

# M. O. Aremu, A. Olonisakin, B. O. Atolaye and C. F. Ogbu

Department of Chemistry, Nasarawa State University, P. M. B. 1022, Keffi, Nigeria.

#### Abstract

The nutritional composition and functional properties of *Prosopis Africana* were studied using standard analytical techniques. The results gave proximate composition as follows: moisture, total ash, ether extract, crude protein, crude fibre and carbohydrate were 1.9, 4.4, 12.8, 23.6, 3.3 and 54.0g % respectively. The predominant mineral was K (617.5mg/100g sample) followed in the highest rank by Mg (420.1mg/10g sample). Other minerals such as Na, Ca, P, Mn, Cu, Zn, Fe and Cr were 110.7, 362.5, 196.4, 36.2, 46.2, 22.4, 15.5 and 0.2mg/100g sample respectively while Cd and Pb were not detected. Amino acid analysis revealed that the sample contained nutritionally useful quantities of most of the essential amino acids. The first limiting amino acid was Thr and predicted isoelectric point was 5.4. Functional properties results were: foaming capacity, 3.9 %, foaming stability 96.2 %; water absorption capacity, 340.0; oil absorption capacity, 120.0 %; emulsion capacity 30.0 mLg<sup>-1</sup>; emulsion stability 38.4 mLg<sup>-1</sup>; least gelation capacity, 16.0 % and bulk density 0.5268 mLg<sup>-1</sup>. The protein solubility of the *P. Africana* was found to have minimum solubility at pH 4.5.

Key words : Nutritional composition, functional properties, Prosopis Africana

### Introduction

*Prosopis Africana (P. Africana)* is a *Leguminosae* belonging to the family of Fabaceae. It reaches 4-20m in height; has an open crown and slightly rounded buttresses; bark is very dark, scaly, slash, orange to redbrown with white streaks. It was once said that *P. Africana* is the only tropical *Africana Prosopis* species, occurring from Senegal to Ethiopia in the zone between the Sahel and savannah forests. Due to extensive overex

ploitation, it has disappeared from extensive parts of the Southern Sahel and the adjavent Sudan savannahs (Von May Dell *et al.*, 1986 and Vogt, 1995). The trees of P. Africana are common in the Middle belt and Northern parts of Nigeria. The local names are "Kiriya" and "Okpehe" in Hausa and Idoma/Tiv languages in Nigeria respectively. In many areas where the trees are grown/available, the fermented seeds of *P*. *Africana* are used as a food condiment. Other traditional processing of *P. Africana* are well known but very little on the nutritional evaluation and functional properties as an ingredient in foods formulation. Therefore, the present study aims at finding the nutritive values as well as functional properties of *P. Africana*.

## **Materials and Methods**

### **Collection and treatment of sample**

Seeds of *P. Africana* were purchased from Markurdi market in Benue State, Nigeria. The seeds were sorted to remove stones and bad ones. Boiling water was added to the seeds, left for seven days and the seeds dehulled, dried in the oven at  $40^{\circ}$  C and freely ground with Kenwood food blender. The flour was kept in a refrigerator at  $4^{\circ}$  C prior to use.

### **Proximate analysis**

Proximate analysis of the sample flour for moisture, ash, fat and crude fiber were carried out in at least triplicate using the methods described by Pearson (Pearson, 1976). Nitrogen was determined by the micro-Kjeldahl method described by AOAC (OAC, 1990), and the percentage nitrogen was converted to crude protein by multiplying with the factor - 6.25. Carbohydrate was determined by calculating the difference.

#### Mineral analysis

The minerals were analyzed from solutions obtained by fitst dry-ashing the seed flour at  $550^{\circ}$  C. The ash obtained was boiled with 15ml 15ml of 20 % hydrochloric acid in a beaker, filtered into a 100ml standard flask and made up to the mark with distilled water. Sodium and potassium were determined by using a flame photometer (Pearson, 1976); phosphorus by using colorimeter (Aremu, *et al.*, 2005) and other metals AAS (Techtron, 1975). All determinations were done in triplicate.

### Amino acid analysis

About 2.0g sample flour was defatted with chloroform/methanol mixture Between 30-35mg of defatted sample was put in glass ampoule. Seven milliliter of 6M HCI was added and oxygen expelled by passing nitrogen into the ampoule. The sealed ampoule was put in an oven at  $105 + 5^{\circ}$  C for 22h and later allowed to cool before the content was filtered. The filtrate was then evaporated to dryness at 40° C under vacuum in a rotary evaporator. Residue was dissolved with 5ml acetate buffer (pH=2.0). Amino acid analysis was done by Ion Exchange chromatography (IEC) (Pearson, 1976) using the Technicon Sequential Multisample Amino Acid Analyzer (TSM) (Technicon Instruments Corporation, New York). The amino acid score was determined by the following formula (FAO/WHO, 1973) -

Amino acid score =

mg of amino acid per g of test - protein mg of amino acid per g protein in reference pattern

Predicted iso-electric point (p1) was estimated by applying the equation (Olaofe and Akintayo, 2000) -

р

Where pIm is the isoelectric point of the mixture of amino acids, Pli, is the iso-electric point of the i<sup>th</sup> amino acid in the mixture and  $X_i$  is the mass or mole fraction of the i<sup>th</sup> amion acid in the mixture and there are n amino acids in the mixture (Graham Solomons, 1984).

### **Functional properties**

The variation of protein solubility with pH was determined as described by Olaofe, (1994) and Akintayo *et al.*(2002). The protein of the supernatant was determined using the micro-Kjeldahl method (OAC, 1990) and exoressed as mg protein per mikkiliter, the water and oil absorption; and emulsion capacities were determined as reported by Beuchat (1977) using Executive Chef Oil with density of 0.98g/ml. The foaming capacity and stability and least gelation concentration were determined using the procedure of Coffman and Garcia (1977). Bulk density of the seed flour was determined

using the procedure of Wand and Kinsella (1976) modified by Akpapunam and Markakis (1981); and Narayana and Narasinga Rao (1984). All chemicals used were of analytical (Analar) grade.

### **Results and Discussion**

### **Proximate composition**

Table I presents the proximate composition of *P. Africana* flour. The percentage of moisture  $(1.9\pm0.3\%)$  was low but comparable to rear cowpea  $(1.8\pm01.25\%)$ , cranberry bean  $(1.7\pm0.51\%)$  and kersting's groundnut  $(1.7\pm0.12\%)$  as reported by Aremu *et al.* (2006); and this value is within the range expected to be present in most legumes (Olaofe and Sanni, 1988; and Oyenuga, 1968). The low moisture content will afford a long shelf for *P. Africana.* The value of crude fat  $(12.8\pm1.2\%)$  was higher as compared with

 Table I.
 Proximate composition (g %) of

 P. Africana flour

, j	0
Parameter	Concentration <sup>a</sup>
Moisture	$1.9 \pm 0.3$
Total ash	$4.4 \pm 0.1$
Other extrat	12.8 <u>+</u> 1.2
Crude protein	$23.6 \pm 1.5$
Crude fibre	$3.3 \pm 0.0$
Carbohydrate by difference	$54.0 \pm 0.8$
Energy (KJ/100g) <sup>b</sup>	1792.8

<sup>a</sup>Values are mean  $\pm$  standard deviation of triplicate determinations

<sup>b</sup>calculated metbolisble energy (KJ.100g) : (protein  $x17 \pm fat x 37 \pm carbohydrate x 17$ )

42(3) 2007

the values reported for two varieties of bambara groundnut (4.10 and 6.72%) (Aremu, 2006), and pear millet and quinoa (7.6 and 6.3%) (Oshodi, et al. 1999) but lower than soybean (23.5%) (Paul and Southgate, 1985), C. vulgaris (47.7%) (Ige et al. 1984), pumkin (49.2%) (Fagbemi and Oshodi, 1991) and T. occidentalis (54.4) (Olaofe et al. 1994). This is an indication that P. Africana cannot be grouped as part of oil-rich legume seed. The quantity of crude protein  $(23.6\pm1.5\%)$  was high compared with crude proteins in protein-rich foods such as soybeans, cowpeas and pigeon pea (Olofe, et al. 1993) and Kersting's groundnut (Aremu, et al., 2006). The crude fibre  $(3.3\pm0.0\%)$  is comparable with most legumes such as pigeon pea and cowpea (Aletor, et al. 1989); P. coccineus (Aremu et al. 2006) and oil seeds (Olaofe, et al. 1994). There is evidence that dietary fibre has a number of beneficial effects related to its indigestibility in the small intestine (Asp, 1996). The calculated metabolisable energy value of 1792.8KJ/100g sample shows that P. Africana is favourably comparable to cereals (Brain and Allan, 1977) in terms of its energy values.

#### **Mineral composition**

Mineral content of *P. Africana* is presented in Table II. Potassium was found to be the most abundant mineral. This is in close agreement with the observation of Olaofe and Sanni (1988) that potassium was the most predominant mineral in Nigerian agricultural

<i>Africana</i> flour	
Mieneral	Concentration <sup>a</sup>
Na	110.7 <u>+</u> 0.1
Κ	617.5 <u>+</u> 0.1
Mg	1420.1 <u>+</u> 0.1
Ca	362.5 <u>+</u> 0.2
Р	196.4 <u>+</u> 0.1
Mn	36.2 <u>+</u> 0.4
Cd	ND
Cu	46.2 <u>+</u> 0.7
Zn	22.4 <u>+</u> 0.0
Pb	ND
Fe	15.5 <u>+</u> 0.4
Cr	0.2 <u>+</u> 0.5
Na/K	0.18
Ca/P	3.76
Ca/Mg	0.86

Table II.Mineral composition (mg/100g) of P.

Values are mean  $\pm$  standard deviation of triplicate determinations

ND = not detected

products. Magnesium was found to be the next highest mineral component amounting to about  $420.1\pm0$ . 1mg/100g sample. It has been reported that magnesium is an activator of many enzyme systems and maintains the electrical protential in nerves (Ferrao, *et al.* 1987). The seed was rich in potassium, magnesium, calcium and phosphorus while the values of copper, zinc, irpn and manganese were low. It is to be noted that cadmium and lead were not in the detectable range. The mineral content in the flour is favourably comparable with that of soybean and cowpea (Olaofe and Sanni, 1988), African yam bean

(Adeyeye, 1989) and Bilphia sapida (Akintayo, et al. 2002) and Triticum durum (Adeyeye and Aye, 2005). This indicates that P. Africana is good for feed supplement. The ratio of sodium to potassium, Na/K, in the body is of great concern for prevention of high blood pressure. Na/K ratio less than one is recommended (Nieman, et al. 1992). The Na/K value of 0.18 (Table II) which is less than one indicates that P. Africana would probably save the capacity to reduce high blood pressure. The Ca/P and Ca/Mg ratios were 3.76 and 0.86 respectively. The value of Ca/Mg is very close the recommended value of 1.0 (National Research Council, 1989). Food is considered 'good' if the Ca/P ratio in the present study (3.76), provides an indication P. Africana would serve as good source of minerals for bone formation.

### Amino acid composition

The result of amino acid composition is shown in Table III. Asp and Glu were found to be the most abundant as expected to be present in legumes. Asp and Glu together make upto 23.3mg/100g protein with a percentage of 31.3%. This value is in close agreement with the report of Olaofe and Akintayo (2000); Kuri *et al.* (1991); Oshodi *el. al* (1998); Aremu *et al.* (2006) and Adeyeye (2004) who had observed that Asp and Glu were the most abundant in legumes and nuts. Leu was also found to be in abundance with value of 6.4mg/100g protein. Table IV depicts the essential, non-essential

protein) of 1. Alfredata nour		
Amino acid	Concentration	
Lysine (Lys) <sup>a</sup>	4.2	
Histidine (His) <sup>a</sup>	2.5	
Arginine (Arg) <sup>a</sup>	5.0	
Aspartic acid (Asp)	10.0	
Threonine (Thr) <sup>a</sup>	2.2	
Serine (Ser)	3.2	
Glutamic acid (Glu)	13.3	
Proline (Pro)	3.0	
Glycine (Gly)	3.3	
Alanine (Ala)	2.8	
Cyctine (Cys)	1.3	
Valine (Val) <sup>a</sup>	4.2	
Methionine (Met) <sup>a</sup>	1.4	
Isoleucine (Ile) <sup>a</sup>	3.7	
Leucine (Leu) <sup>a</sup>	6.4	
Tyrosine (Tyr)	3.2	
Phenylanine (Phe) <sup>a</sup>	4.7	
lso-electric point (pl)	5.4	
Phenylanine (Phe) <sup>a</sup>	4.7	

 Table III. Amino acid composition (g/100g protein) of P. Africana flour

<sup>a</sup>Essential amino acids

acidic, neutral and sulphur containing amino acids. The total essential amino acids (TEAA) (with His) was 34.3mg/100g protein which represents 46.1%. This is comparable with values obtained for selected oil seeds (Olaofe, 1994), varieties of V. subterranean and *P. coccineus* (Aremu, *et al.* 2006). Pellet and young (1977) reported that the nutritive value of a protein depends primatily in its capacity to satisfy the needs for nitrogen and essential amino acid. Therefore *P. Africana* can be considered as good diets that can provide the required essential amino acid. The

Classification	Concentration
Total amino acid (TAA)	74.4
Total non-essential amino acid (TNEAA)	40.1
% TNEAA	53.9
Total essentoal amino acid (TEAA)	
-with histidine	34.3
-without histidine	31.8
% TEAA	
-with histidine	46.1
-Without histidine	42.2
Essential alphatic amino acid (EAAA)	16.5
Essential aromatic amino acid (EArAA)	4.7
Total neutral amino acid (TNAA)	39.4
% TNAA	53.0
Total acidic amino acid (TAAA)	23.3
% TAAA	31.3
Total basic amino acid (TBAA)	11.7
%TBAA	15.7
Total sulphur amino acid (TSAA)	2.7
% cystine in TSAA	48.1

Table IV. Classification of amino acid composition (g/100g protein) of P. Africana flour

observed value 16.5g/100g protein for essential aliphatic amino acid (EAAA) which constitute the hydrophobic region of protein indicates that *P. Africana* may have better emulsification properties. The total acidic amino acid (TEAA) (31.3g%) was found to be greater than the total basic amino acid (TBAA) (15.7g%). This indicates that its protein is probably acidic in nature. The scoring table (Table V) reveals that first and second limiting amino acids were Thr (0.55) and Lys (0.76) respectively. Apart from Lys, other EAA that have been described as the limiting AA are Met (and/or Cys) and Try (Bender, 1992). Try was not determined. The predicted iso-electric point (p1) of 5.4 (Table III) is a good starting point in predicting the pl for *P. Africana* protein in order to enhance a quick precipitation of protein isolate from biological samples (Olaofe and Akintayo, 2000).

### **Functional properties**

Fig. 1 shows the *P. Africana* protein solubility as a function of pH. This Provides a good index of the potential or limitation of a protein as a functional ingredient. The minimum

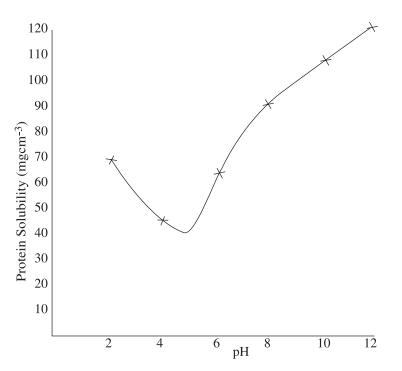


Fig. 1. Variation of protein solubility of P. Africana flour

AAC	PAAESP <sup>a</sup> (g/100g protein)	EAAC	AAS
Ile	4.0	3.7	0.93
Leu	7.0	6.4	0.91
Lus	5.5	4.2	0.76
Met + Cys (TSAA)	3.5	2.7	0.77
Phe + Tyr	6.0	7.9	1.32
Thr	4.0	2.2	0.55
Try	1.0	nd	na
Val	5.0	4.2	0.84
Total	36.0	31.3	6.08

Table V. Amino acid scores of P. Africana flour

<sup>a</sup>Source: Belshant *et al*<sup>53</sup> (1975)

nd = Not determined and na = Not available

AAC = Amino acid composition;

PAAESP = Provisional amino acid (Egg) scoring pattern;

EAAC = Essential amino acid composition (See Table III) and

AAS = Amino acid scores.

solubility occurred at 4.5. The observed value is in excellent agreement with the melon seed protein's minimum solubility at pH 4.5 reported by Ige *et al.* (1984). The enhanced solubility of the flour protein in acid region indicates that the protein may be useful in the formulation of acid foods such as protein rich carbonated beverages.

The results of foaming capacity and stability are presented in Table VI. *P. Africana* produced thin foams with medium size air cells and its foaming capacity (FC) of  $3.92\pm0.7$  % was very low when compared with benni seed (18.0 %), pearl millet and quinoa (19.0 %) reported by Oshodi *et al.* (1999); selected sea foods (6 - 14 %) (Ogunlade, *et al.* 2005); varieties of legume seeds (7.9 - 155%) (Aremu, *et al.* 2006); soybean (66 %) (Lin, *et al.* 1974); great Northern bean (32 %) (Sathe, *et al.* 1982); and varieties of African yam

 Table VI. Functional properties of P. Africana

 Flour<sup>a</sup>

rioui	
Property	Value
Foaming capacity (FC) %	$3.9 \pm 0.7$
Foaming stability (FS) %	96.2 <u>+</u> 0.5
Water absorption capacity (WAC) %	$340.0\pm0.0$
Oil absorption capacity (OAC) %	$120.0\pm1.2$
Emulsion capacity (EC) mLg <sup>-1</sup>	30.0 <u>+</u> 1.4
Emulsion stability (ES) mLg <sup>-1</sup>	38.4 <u>+</u> 2.0
Least gelation concentration (LGC) %	16.0 <u>+</u> 0.0
Bulk density (BD)gmL <sup>-1</sup>	$0.5268 \pm 0.3$

<sup>a</sup>Values are mean  $\pm$  standard deviation of triplicate determinations

bean (54.0-55.0 %) (Oshodi, et al. 1997). Foaming stability recorded after 12hrs was  $96.2\pm0.5\%$ . Water absorption capacity, WAC (340 + 0.0%) (Table VI) is higher than that of sun flower (107 %) and soy bean (130 %) reported by Lin et al (1974); Z. variegatus (127.5 %) (Olaofe et al. 1998) and various liman bean (130 - 142 %) (Oshodi and Experigin, 1989). The high WAC recorded in this report suggested that P. Africana may be used in the formulation of some foods such as soups or baked products (Olaofe et al. 1998). The oil absorption capacity (OAC) is also presented in Table VI. The value is lower than those of varieties of legume seeds (127.8-172.0 %) (Aremu, et al. 2006) and cowpeas (281-310 %) (Olofe, et al. 1993); Z. variegatus (46.7 %) (Olaofe, et al. 1998), pigeon pea (89.7 %) (Oshodi and Ekperigin, 1989). OAC is important as oil acts as a flavour retainer and improves the mouth feel of foods. P. Africana will be good in these respects. For emulsifying activity; P. Africana has a low percentage of emulsion capacity, 30.0±1.4mLg-1 (Table VI) in comparison with benni seed, pear millet and quinoa (63.0, 89.0 and 104.0 %) (Lin, et al. 1974) but higher than values reported for soybean (18%) (Lin, et al. 1974) and pigeon peas (7-11%) (Oshodi and Ekperigin, 1989). This indicates that P. Africana might be useful in the production of sausages, soups and cakes (Kinsella, 1979). Table VI still depicts the emulsion stability after 24hrs, which is the volume of water separated. The value obtained for *P. Africana*  $(38.4 + 20 \text{mLg}^{-1})$  is close to pear millet (34.0%) (Oshodi, et al. 1999), The least gelation concentration (16 %) which is also shown in Table VI is higher than the values reported for some legumes e.g. pigeon pea (10 %) (Oshodi and Ekperigin, 1989), cowpea (10 %) (Attschul and Wileke, 1985) and lupin seed (14 %). The ability of protein to form gels and provide a structural matrix for holding water, flavours, sugars and food ingredients is useful in food application and in new product development, thereby providing an added dimension protein functionality to (Padmashree and Shashilala, 1987). The bulk density (BD) was 0.5268+0.3 gmL<sup>-1</sup> (Table VI). This value is higher than the values reported for various samples of extrusion texturized soya products with varied protein and soluble sugar contents  $(0.2382-0.4460 \text{gmL}^{-1})$  (Adeyeye and Adamu, 2003).

### Acknowledgements

We are very grateful to the Management of Nasarawa State University, Jeffi for providing some of the facilities used; and technical assistance rendered by Mr. Bernard Ashikaa (Laboratory Technologist) of Chemistry Department, Nasarawa State University, Keffi is highly acknowledged.

#### References

- Adeyeye, E.I. and Adamu, A.S. (2003) Biosciences Biotech. Res. Asia. **3**(2): 265 -272.
- Adeyeye, E.I. and aye, P.A. (2005) *Pakistan J. of Nutrition*, **4**(3) : 187-196.
- Adeyeye, E.I. (1996) *Bangladesh J. Sci and Res.* **31**(3): 1711.
- Adeyeye, E. I. (2004) Oriental J. of Chemistry, 20(3), 471-473.
- Akintayo, E.T., Adebayo, E.A. and Arogundade, L.A. (2002) *Food Chemistry*, **77** : 333-336.
- Akpapunam, M.a. and Makakis, P. (1981) *J. Food Science* **46** : 972 - 973.
- Aletor, V.A. and Aladedun, O.O. (1989) *Die Nahrung*, **10** : 999-1007.
- Aremu, M.O. (2006) *J. of Applied Sciences* **6**(9) : 1900-1903.
- Aremu, M.O., Olaofe, O. and Akintayo, E.T. (2006) J. of Food Agriculture and Environment, 4(2): 39-43.
- Aremu, M.O., Olaofe, O. and Akintayo, E.T. (2006) *J. of Food Technology*, **4**(3) (1n press).

- Aremu, M.O., Olaofe, O. and Akintayo, E.T. (2006) *Pak. Journal of Nutrition* **5**(1) : 34-38.
- Aremu, M.O., Olaofe, Olaofe, O. and Akintayo,
  E.T. (2006) *Int. J. of Chemistry*, 16(1): 57-64.
- Aremu, M.O., Olonisakin, A., Otene, I.W. and Atolaye, B.O. (2005) Oriental J. of Chemistry, 2(3): 419 - 426.
- Asp. N.G. (1996) J. Food Chemistry, 7(1): 9 14.
- Attschul, A.M. and and Wileke, H.L. (1985) New protein foods Vol. 5 seed storage protein, London: Academic press 107-129.
- Belshant, A.A., Lyon, C.K. and Kohler, G.O. (1975) In: 'Food protein sources' (N. W. Pirieed). Cambridge University press, Cambridge, U. K..
- Bender, A. (1992) Meat and meat products in human nutrition in developing countries, FAO food and Nutrition paper, 53, FAP, Rome, Rome.
- Beuchat L.R. (1977) Functional and elctrophoretic characteristics of succinylated peanut flour protein. *J. Agric. Food chem.* **25** : 258-261.
- Brain, A.F. and Allan, G.C. (1977) Food Science -A Chemical Approach, 3rd Ed. Ellis Horwood Chichester, England, pp. 214.

- Coffmann, C.W. and Garcia, V.C. (1977) Functional properties and amino acid content of a protein isolate from mung bean flour. *J. Food Technology*, **12** : 473 - 482.
- Fagbemi, J.N. and Oshodi, A.A. (1991) *Nigeria* Food Journal **9**: 26-32.
- FAO/WHO. (1973) Energy and protein requirement. Technical report series No. 52, WHO Geneva, Switzerland, 1-118.
- Ferrao, J.E.M., Ferrao, A.M.B.C. and Anatures, A.M.G. (1987) Garcia deorta, serieda estudes Agronomicos, 14(1 - 2), 35 - 39.
- Graham Solomons, T.W. (1984) Organic Chemistry, 3rd Ed. John wiley & Sons, 1021-1022.
- Ige, M.M., Ogunsua, A.O. and Oke, O.L.J. (1984) Agric. Food Chemistry, **32** : 822 - 825.
- Kinsella, J.E. (1979) J. of the American Oil Chemists Society 56: 242 - 258.
- Kuri, Y.E., Sundav, R.K., Kahuwi, C., James, G. P. and Rwett, D.E.J. (1991) Agric. Food Chemistry 39 : 1702.
- Lin, M.J.Y., Humbert, E. S. and Sozulike, F.W.J. (1974) *Food Science*, **39** : 368 - 370.
- Narayana, K. and Narasinga Rao, M.S.J. (1984) *Food Science*, **49** : 944 - 947.
- National Research Council, Recommended Dietary Allowances, 10th Ed. National

Academy press, Washington, D. C. USA (1989). 36. Olaofe, O. and Akintayo, E. T. *The J. of Technoscience*, **4** : 49 - 53 (2000).

- Nieman, D.C., Butterworth, D.E. and Nieman, C.N. (1992) Nutrition Winc Brown Publishers Dubugue, USA. Pp. 237 - 312.
- OAC official methods of analysis, 15th edn. Washington, D. C. Association of official analytical Chemists (1990).
- Ogunlade, I. Olaofe, O and Fadare, T.J. (2005) Food Agric and Environment, **3**(2) : 130 - 133.
- Olaofe, O. and Akintayo, E.T. (2000) *The Journal* of *Technoscience*, **4** : 49.
- Olaofe, O. and Sanni, C.O. (1988) *Food Chemistry* **30** : 73 - 77.
- Olaofe, O., Adeyemi, F.O. and Adediran, (1994) J. of Agric Food Chemistry, **42** : 878 - 8811.
- Olaofe, O., Arogundade, L.A., Adeyeye, E.I. and Falusi, O.M. (1998) *Tropical Science* **38** : 233 - 237.
- Olofe, O. Umar, Y.O. and Adediran, G.O. (1993) Food Chemistry, **46** : 337 - 342.
- Oshodi, A.A. and Ekperigin, M.N. (1989) *Food Chemistry* **34** : 187 - 191.
- Oshodi, A.A., Esuruoso K.O. and Akintayo, E.T. (1998) La Rivisita Italiana Delle Sostanze

grasse. Vol. LXXV, Agosto.

- Oshodi, A.A., Ipinomoroti, K.O. and Adeyeye, E. I. (1997) *Int. J. Food Sci. and nutri.*, **48** : 243 - 252.
- Oshodi, A.A., Ogungbele, H.N. and Oladimeji. Int. (1999) J. of Food science, 50 : 325 -331.
- Oyenuge, V.A. (1968) Nigeria's Foods and Feeding Stuff, 3rd Ed. Ibadan University Press. Pp. 36 - 40.
- Padmashree, T.S. and Shashilala, P. (1987) J. Food Sci. Sci and Tech. 24 : 221.
- Paul, A.A. and Southgate, D.A.T., McCance and Widdowsen's (1985) The Corporation of Foods Royal Society of Chemistry. London.
- Pearson, D. (1976) Chemical analysis of foods.7th edn. London Church Hill Livingstone, 37 - 39.
- Sahni, K.C. (1968) Important trees of the Northern Sudan. United Nations and FAO.
- Sathe, S.K., Deshpande, S.S. and Salunkhe, D.K. (1982) *J. Food Science*, **47** : 491 - 497.
- Techtron, V. (1975) Basic Atomic Absorption Spectroscopy. A modern introduction, Dominican press, Victoria Australia, pp. 104 - 106.

- Vogt, K. A. (1995) Field guide to the identification, propagation and uses of common trees and shrubs of dry land Sudan. SOS, SAHEL International.
- Von May dell, H.J. (1986) Trees and Shrubs of the Sahel - their characteristics and uses. GTZ 6MBH, Esuborn.

Wand, J.C. and Kinsella, J.F. (1976) *J. Food Science*, **41** : 286 - 289.

Received : August 31, 2006; Accepted : July 25, 2007