

## Pesticidal effect of naphthalene and different botanicals against anguimous grain moth (*Sitotroga cerealella*)

T. K. Roy<sup>1\*</sup>, A. Sannal<sup>2</sup>, S. Akter<sup>1</sup>, S. M. M. S. Tonmoy<sup>3</sup>, T. Chakrobarty<sup>4</sup>, M. R. Hasan<sup>5</sup> and M. N. Bari<sup>1</sup>

<sup>1</sup>Entomology Division, Bangladesh Rice Research Institute, BRRI

<sup>2</sup>Plant Pathology Division, Bangabandhu Sheikh Mujibur Rahman Agricultural University, BSMRAU

<sup>3</sup>Farm Management Division, Bangladesh Rice Research Institute, BRRI

<sup>4</sup>Rice Farming System, Bangladesh Rice Research Institute, BRRI

<sup>5</sup>Plant Breeding Division, Bangladesh Rice Research Institute, BRRI

### ARTICLE INFO

Received: 13 June 2023

Revised: 14 August 2023

Accepted: 17 August 2023

eISSN 2224-7157/© 2023 The Author(s).  
Published by Bangladesh Council of  
Scientific and Industrial Research  
(BCSIR).

This is an open access article under the  
terms of the Creative Commons Non  
Commercial License (CC BY-NC)  
(<https://creativecommons.org/licenses/by-nc/4.0/>)

DOI: <https://doi.org/10.3329/bjsir.v58i3.66797>

### ABSTRACT

Anguimous grain moth is a strong seed destroyer causing economic losses to the marginal farmers. Treatments of seeds with different chemicals as remediation of this problem risks environment and creates health hazards. Pesticidal effects of some botanicals and naphthalene on anguimous grain moth inhabiting grains have been tested in this experiment. A total of eight treatments were performed against Anguimous grain moth with one control (T<sub>1</sub>). The treatments contained: T<sub>2</sub>=5 whole dried neem leaf, T<sub>3</sub>=0.25 ml neem oil, T<sub>4</sub>=0.25 g ginger powder, T<sub>5</sub>=0.25 g neem leaf powder, T<sub>6</sub>= 0.25 g mixture of neem leaf powder and ginger powder, T<sub>7</sub>=0.25 g naphthalene, T<sub>8</sub>= 5 whole tulsi leaf and T<sub>9</sub>= 0.25 g neem seed powder. Results indicated that T<sub>7</sub> (100%) showed better performance in case of insect mortality and no F<sub>1</sub> emergence followed by T<sub>3</sub> (53.33%), T<sub>9</sub> (43.66%) and T<sub>6</sub> (40%). Moderate performance was found T<sub>5</sub> and T<sub>2</sub> in case of insect mortality and number of F<sub>1</sub> emergence. But in case of seed germination T<sub>5</sub> (92.00%) showed better performance followed by T<sub>7</sub> (88.00%), T<sub>9</sub> (87.00%), T<sub>6</sub> (85.33%) and T<sub>3</sub> (80.33%). So, naphthalene, neem oil, neem seed powder and neem leaf powder considered as the best treatment among nine treatments for the management of the most destructive Anguimous grain moth.

**Keywords:** Anguimous grain moth (*Sitotroga cerealella*); Neem oil; Naphthalene; Neem leaf powder; Neem seed powder; Storage pest.

### Introduction

Rice (*Oryza sativa* L., Family: Poaceae) is one the most important cultivated food crops in the world as well as in Bangladesh. In the rice cultivation, insect infestation which may be field or storage borne affects production. During cultivation of rice, insects are always present in the field as well as presence in storage condition. Rice needs a long storage period after harvesting until further use for consumption, seed purpose etc. Under storage, rice is infested by Rice weevil (*Sitophilus oryzae*), Anguimous grain moth (*Sitotroga cerealella*), saw-toothed grain beetle (*Oryzaephilus surinamensis*), Red flour Beetle (*Tribolium castaneum*) and Indian Meal Moth (*Plodia interpunctella*) etc and cause extensive economic loss in the

production. Anguimous grain moth (*Sitotroga cerealella*) is a prevalent insect species and often placed at the top of the list as major insect pest of stored rice. Tadesse (2020) reported that in order to ensure that a country has enough food, postharvest storage for proper grain is more crucial than intense and widespread farming. But there are many reasons for rice storage losses. Qu *et al.* (2021) demonstrated that lack of knowledge, inappropriate farming techniques, poor infrastructure and improper harvest management procedures were major causes of rice harvest losses. Reducing postharvest losses of food crops is essential to enhancing agricultural productivity in a sustainable manner (Stathers *et al.* 2020).

\*Corresponding author's e-mail: [tapon.entom@brii.gov.bd](mailto:tapon.entom@brii.gov.bd); [taponroy.brii@gmail.com](mailto:taponroy.brii@gmail.com)

So, management of storage pest is important for reducing postharvest losses. Since 1950s, synthetic insecticides were used to manage storage pests of agricultural crops. However, synthetic chemicals for pest control effects

human, wild and aquatic life and the environment at large (Köhler and Triebkorn, 2013; Muñoz-Quezada *et al.* 2013; Baltazar *et al.* 2014; Yuan *et al.* 2014; Meyer-Baron *et al.* 2015; Guyton *et al.* 2015). Besides, the use of adulterated and expired ineffective pesticides produces rapid evolution of pesticide resistance pathogens in the ecosystem (Stevenson, 2014). Therefore, alternate methods need for the management of rice storage pest which is non-toxic, environment friendly and also free from human health hazard is an urgent research issue. In Bangladesh, poor and marginal farmers store small quantities of rice for consumption and for use as seed and they cannot practice expensive control measures for storage pest. Botanicals containing bio-pesticidal properties in their bioactive components play vital role for the management of rice storage. Some plants are known to contain bioactive metabolites, which show anti-feedant, repellent and toxic effects on a wide range of insect pests (Stevenson, 2014). Examples of these mostly used important bioactive plants are catnip, pale persicaria, holy basil, artemisia, borage, dahlia, ginger, hyssop, chrysanthemum, lime, black pepper, clove, mahogany, neem and ginger etc. Plant extracts in powder or essential oil form from different bioactive plants are known to be effective repellents against different economic storage pests of grains, even for stored cereals (Khan and Gumbs, 2003). Anguimous grain moth cause economic loss, nutritional damage and reduce germination percentage on rice by breeding on grain and feeding of grain. Therefore our present study was undertaken to assess the effectiveness of neem, tulsi and ginger and naphthalene for the management of Anguimous grain moth (*Sitotroga cerealella*) of rice.

### Materials and methods

The present study was conducted at Bangladesh Rice Research Institute (BRRI), Regional Station Rangpur laboratory during the period from June to October 2021. The experiment was carried out following complete randomized design (CRD). A total of eight treatments were performed against one control (T<sub>1</sub>) with three replications. The treatments were:

Treatments	Status
T1	Control (Untreated)
T2	Whole neem leaves, 5 in number
T3	0.25 ml neem oil
T4	0.25 g ginger powder
T5	0.25 g neem leaf powder
T6	0.25 g mixture of neem leaf powder
T7	0.25 g naphthalene
T8	Whole tulsi leaf, 5 in number
T9	0.25 g neem seed powder

ginger bulb (*Zingiber officinale*), fresh neem leaves (*Azadirachta indica*), ripe neem kernel and fresh tulsi (*Ocimum tenuiflorum*) leaf were collected and dried in the oven at 50°C to constant weight. Part of the ginger bulb, fresh dried neem leaf and ripe neem kernel were powdered by electric grinder machine then sieved (mesh number 25) to obtain fine and uniform material and the powdered materials were stored in airtight polythene bag until further treatment. Equal amount of neem leaf powder and ginger powder was mixed together to make mixture of neem leaf and ginger powder. Neem oil (pressed organic neem oil) and naphthalene (99% purity) were collected from the market. At the beginning of the experiment *Sitotroga cerealella* population was collected from the BRRI regional station Rangpur store and reared on rice (BRRI dhan87) in laboratory. The female and male moths were distinguished by visual observation of their abdominal tergites and the size of the body with the help of an electron microscope (Olympus Model SZ2-STU2, Tokyo, Japan and scanning objective lens was zoomed at 4×). Fresh and de-infected BRRI dhan87 were collected from BRRI Regional Station seed store house and fully sundried for 3 days. Then seeds were taken in 27 Petri dishes @ 25 g/Petri dishes. A weighing balance (KERN ABJ220-4NM, Balingen, Germany) was used to measure the powder extracts. After the application of treatments, 5 pairs of fresh adults were introduced in each replication and they were kept for 5 days for oviposition at 65% RH and 30°C. Number of dead insect were counted at 24 h after treatment, 48 h after treatment and 72 h after treatment and dead insect were removed. The Petri dishes were kept in regular observation for F<sub>1</sub> adult emergence. Adults were removed and data collection was continued until no F<sub>1</sub> moth was found. Insect mortality data were calculated by using the square root transformation formula. All data were subjected to statistical

analysis separately by using analysis of variance technique by R software (versions 4.2.1, 2022). Percent insect mortality

$$\text{Insect mortality (\%)} = \frac{\text{Number of dead insect at treated treatment}}{\text{Total number of insect treated}} \times 100$$

was calculated by using the following formula.

#### Germination percentage

One hundred rice seeds were taken randomly in a Petri dish from each replication of each treatment. Rice seeds were taken separately for each replication. The seeds were placed on moist filter papers in the Petri dish and maintained at room temperature. The number of germinated seed were counted

$$\text{Germination (\%)} = \frac{\text{Number of germinated seeds}}{\text{Total number of tested seeds}} \times 100$$

and recorded after completion of germination. The percentage germination was computed using the following formula.

## Results and discussion

#### Number of dead insects

The number of cumulative dead insects at 24 h after treatment and 48 h after treatment showed statistically significant results. Data presented in Table I showed that after 24 h of treatment the highest number of dead insects was recorded in T<sub>7</sub> (2.97) treatment. The second most dead

insect was observed in T<sub>3</sub> (1.95) treatment. Statistically similar results were found in T<sub>6</sub> (1.58) and T<sub>9</sub> (1.58) treatment and T<sub>2</sub> (1.34) and T<sub>5</sub> (1.46) treatments. The lowest dead insect was found in T<sub>1</sub> (0.71) treatment. After 48 h of treatment the highest number of dead insect was found in T<sub>9</sub> (1.46) and T<sub>6</sub> (1.46) treatment followed by T<sub>2</sub> (1.34), T<sub>3</sub> (1.34) and T<sub>5</sub> (1.34) treatment and which were statistically similar. The lowest number of dead insects was found in T<sub>1</sub> treatment. After 72 h of treatment, there were statistically no significant difference among the treatments but numerically different. Numerically highest dead insect were found in T<sub>2</sub>, T<sub>3</sub>, T<sub>5</sub>, T<sub>7</sub> and T<sub>8</sub> treatment and lowest were found in T<sub>1</sub> and T<sub>4</sub> treatment. From the result, it might be concluded that highest number of dead insect was found at T<sub>7</sub>, i.e naphthalene followed by T<sub>3</sub> (neem oil), T<sub>9</sub> (neem seed powder) and T<sub>6</sub> (mixture of neem leaf and ginger powder). Highest dead insects were found at naphthalene due to its solid form that turns into toxic gas and due to the exposure of toxic gas it caused huge inhalation and its exposure did cause dermal contact, hemolytic anemia and neurological damage as well. Azadirachtin bioactive ingredients present in neem plant that are responsible to kill insect. Facknath (2006) reported that neem (*Azadirachta indica* A. Juss.) can

**Table I. Effect of botanicals and chemicals against anguimous grain moth after different duration of treatments**

Treatment	No of dead insects after different duration of treatments		
	24 hours	48 hours	72 hours
T <sub>1</sub> (Control)	0.71 f	0.71 d	0.71
T <sub>2</sub> (5 whole dried neem leaf)	1.34 cd	1.22 ab	1.05
T <sub>3</sub> (0.25 ml neem oil)	1.95 b	1.34 ab	1.05
T <sub>4</sub> (0.25 g ginger powder)	1.05 e	0.88 cd	0.88
T <sub>5</sub> (0.25 g neem leaf powder)	1.46 cd	1.34 ab	1.05
T <sub>6</sub> (0.25 g mixture of neem leaf & ginger powder)	1.58 c	1.46 a	0.88
T <sub>7</sub> (0.25 g naphthalene)	2.97 a	1.22 ab	1.05
T <sub>8</sub> (5 whole dried tulsi leaf)	1.22 de	1.34 ab	1.05
T <sub>9</sub> (0.25 g neem seed powder)	1.58 c	1.46 a	1.22
Level of significance	***	**	Ns
LSD	0.2583719	0.337307	
CV%	9.768086	16.72475	26.49772

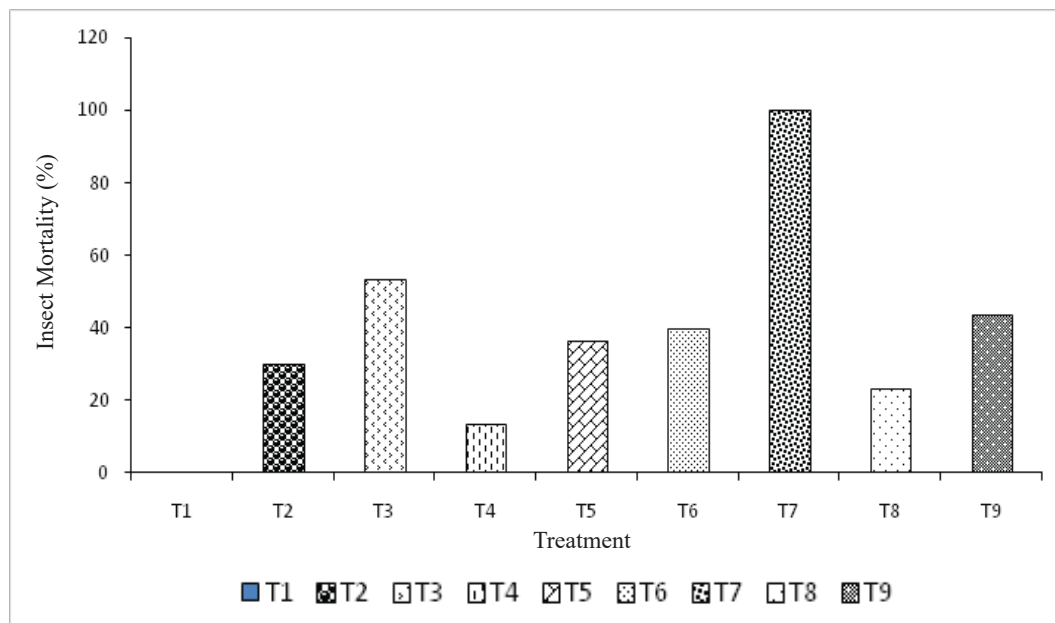
\*, \*\*, \*\*\* indicate significant at 5%, 1% and 0.1% level of probability, respectively and Ns indicates not significant. Values followed by the different letter(s) are significantly different from each other.

reduce insect populations infesting stored products through its toxic effects on the pests by disrupting the insect growth.

#### Percent insect mortality of anguimous grain moth

Percent insect mortality showed statistically significant

*al.* (2015) reported that naphthalene-induced respiratory toxicity is related to lipid peroxidation, disruptions of membrane components, and imbalanced energy supply. The study of Obeng-Ofori *et al.* (1998) have shown that, the contact toxicity of naphthalene (concentrations between 100 µg and 100 mg) on stored product insect pest with mortalities above



**Fig. 1. Insect (Anguimous grain moth) mortality (%)**

The treatments were:

T<sub>1</sub>=Control, T<sub>2</sub>=5 whole dried neem leaf, T<sub>3</sub>=0.25 ml neem oil, T<sub>4</sub>=0.25 g ginger powder, T<sub>5</sub>=0.25 g neem leaf powder, T<sub>6</sub>= 0.25 g Mixture of neem leaf powder and ginger powder, T<sub>7</sub>=0.25 g Naphthalene, T<sub>8</sub>= 5 whole tulsi leaf and T<sub>9</sub>= 0.25 g neem seed powder.

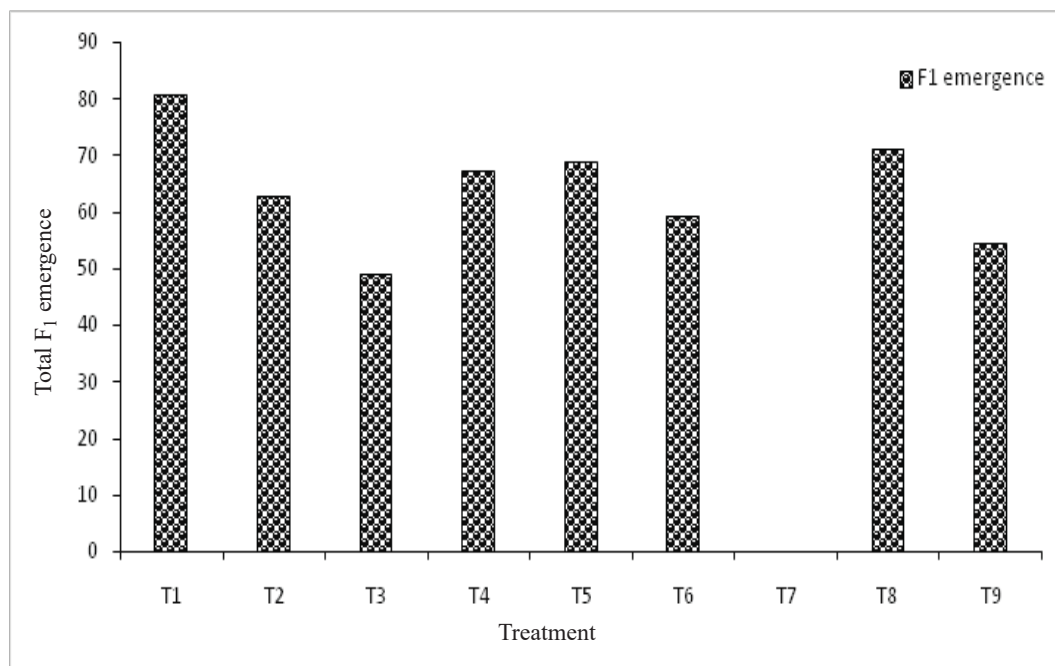
variation for commonly used different botanicals and chemical used as experimental treatments for the management of anguimous grain moth in stored rice grain (Figure 1). The highest (%) insect mortality was demonstrated in T<sub>7</sub> (100%) and which was statistically different from others. The second highest (%) insect mortality was observed in T<sub>3</sub> (53.33%) treatments which was also different from other treatments followed by T<sub>9</sub> (43.66%) and T<sub>6</sub> (36.33%) treatment and they were statistically significant. No insect mortality was recorded in T<sub>1</sub> (control) treatment. From the figure I it might be concluded that naphthalene (T<sub>7</sub>) was found as 100% mortality in case of anguimous grain moth due to its emission of toxic gas and pungent smell. Naphthalene acts by the activation of cytochrome P450 enzyme in neuronal channel which triggers acute toxicity of neural cells (Li *et al.* 2011). Lin *et*

70% in maize weevil, groundnut weevil, red flour beetle, and large grain borers. On the other hand, neem as a botanical pesticide has many excellent attributes including its broad-spectrum in insect growth regulatory effects, systemic action in some plants, minimal effects on natural enemies and pollinators, rapid degradation in the environment, and no toxicity to vertebrates. It is also reported that the lethality of neem oil is confirmed depending on the concentration applied and can be compared to neurotoxic insecticides and growth inhibitors, as a potent natural insecticide (Isman, 2006; Mourão, 2016). Jibrin and Mohammed (2020) reported that active ingredients in neem tree that make the leaves bitter and azadirachtin bioactive ingredients in neem are useful for insect repellent and mortality role.

### Number of $F_1$ emergence

Number of  $F_1$  emergence showed statistically significant results for the application of experimental treatment for the management of Anguimous grain moth in storage rice grain (Figure 2). Highest number of  $F_1$  emerged in  $T_1$

can be found in fruits and leaves. Alexander *et al.* (2023) reported that neem oils contain insecticidal, anti-ovipositant and ovicidal properties that reduce  $F_1$  emergence. Neem oil is a feeding inhibitor, delaying development and growth, reducing fecundity and fertility, changing behavior and causing anomalies in eggs, larvae and adults of



**Fig. 2.  $F_1$  emergence of Anguimous grain moth**

The treatments were:

$T_1$ =Control,  $T_2$ =5 whole dried neem leaf,  $T_3$ =0.25 ml neem oil,  $T_4$ =0.25 g ginger powder,  $T_5$ =0.25 g neem leaf powder,  $T_6$ =0.25 g Mixture of neem leaf powder and ginger powder,  $T_7$ =0.25 g Naphthalene,  $T_8$ = 5 whole tulsi leaf and  $T_9$ = 0.25 g neem seed powder.

(80.67) control untreated treatment which was followed by  $T_8$  (71.00),  $T_5$  (68.67),  $T_4$  (67.00),  $T_2$  (62.67),  $T_6$  (59.00),  $T_9$  (54.33) and  $T_3$  (48.67) respectively. No  $F_1$  was emerged in  $T_7$  treatment. Due to the highest mortality and toxic gas emission of naphthalene and thus acted as poison for anguimous grain moth responsible for lowest or no  $F_1$  emergence. Neem oil contain bioactive ingredient azadirachtin that was higher toxic by feeding and thus causes insect mortality, delaying insect growth, curtailing fertility and emergence of  $F_1$  population. Azadirachtin bioactive ingredient found different parts of neem plant. Bruce *et al.* (2004) reported that azadirachtin is the main compound of the neem oil with insecticidal activity and

insects or mites reported by Masood *et al.* (2006).

### Germination of seed

Germination (%) of treated seed showed statistically significant variation for the application of experimental treatments of anguimous grain moth management in stored rice grain (Table II). The highest germination was found in  $T_5$  (92.00%) and which was statistically differ from other treatments.  $T_7$  (88.00%),  $T_9$  (87.00%) and  $T_6$  (85.33%) treatment showed statistically similar result for seed germination. The lowest seed germination was found in  $T_1$  (73.00%) untreated control treatment due to highest  $F_1$  emergence, lowest insect mortality and damage by breed-

**Table II. Effect of botanicals and chemicals against Anguimous grain moth on germination (%) of seed**

Treatment	Seed germination (%)	% Germination seed over control
T <sub>1</sub> (Control)	73.00 e	--
T <sub>2</sub> (5 whole dried neem leaf)	78.00 cd	6.85
T <sub>3</sub> (0.25 ml neem oil)	80.33 c	10.05
T <sub>4</sub> (0.25 g ginger powder)	78.00 cd	6.85
T <sub>5</sub> (0.25 g neem leaf powder)	92.00 a	26.03
T <sub>6</sub> (0.25 g mixture of neem leaf & ginger powder)	85.33 b	16.90
T <sub>7</sub> (0.25 g naphthalene)	88.00 b	20.55
T <sub>8</sub> (5 whole dried tulsi leaf)	77.00 d	5.48
T <sub>9</sub> (0.25 g neem seed powder)	87.00 b	19.18
Level of significance	***	--
LSD	2.839869	--
CV%	2.017103	--

\*, \*\*, \*\*\* indicate significant at 5%, 1% and 0.1% level of probability, respectively. Values followed by the different letter(s) are significantly different from each other.

ing and feeding on rice grain. Similarly in case of percent seed germination over control was found highest in neem leaf powder (T<sub>5</sub>) followed by naphthalene (T<sub>7</sub>), neem seed powder (T<sub>9</sub>), T<sub>6</sub> (mixture of neem leaf powder and ginger powder) and T<sub>3</sub> (neem oil). Hossain *et al.* (2018) reported that, dried neem leaf powder showed the highest percent of seed germination over control. From our study, the trend of efficiency among different botanicals and chemical in terms of percent seed germination was T<sub>5</sub> (neem leaf powder) > T<sub>7</sub> (naphthalene) > T<sub>9</sub> (neem seed powder) > T<sub>6</sub> (mixture of neem leaf powder and ginger powder) > T<sub>3</sub> (neem oil) > T<sub>2</sub> (whole dried neem leaf) > T<sub>4</sub> (ginger powder) > T<sub>8</sub> (whole dried tulsi leaf) > T<sub>1</sub> (control).

### Conclusion

Findings of the study concluded that among the eight treatments against one control, highest insect mortality was found in T<sub>7</sub> (0.25 g naphthalene) treatment followed by T<sub>3</sub> (0.25 ml neem oil) in case of insect mortality. No F<sub>1</sub> emergence was found in T<sub>7</sub> (0.25 g naphthalene) and lowest F<sub>1</sub> emergence was observed in T<sub>3</sub> (0.25 ml neem oil) followed by T<sub>9</sub> (0.25 g neem seed powder). The highest seed germination was showed in T<sub>5</sub> (0.25 g neem leaf powder) followed by T<sub>7</sub> (0.25 g naphthalene) and T<sub>9</sub> (0.25 g neem seed powder) in case of percent seed germination.

### Acknowledgement

Authors are greatly thankful to principle scientific officer and head, Bangladesh Rice Research Institute (BRRI), regional station Rangpur for the fund and immense assistance for conducting the experiment.

### Reference

- Alexander MM, Mohammed MA, Emmanuel O and Ali S (2023), Toxicity of seed oils of neem (*Azadirachta indica*) and desert date (*Balanites aegyptiaca*) on the development of tropical warehouse moth (*Ephestia cautella*) in maize (*Zea mays*), *Discovery* **59**: e69d1255
- Baltazar MT, Dinis-Oliveira RJ, de Lourdes Bastos M, Tsatsakis AM, Duarte JA and Carvalho F (2014), Pesticides exposure as etiological factors of Parkinson's disease and other neurodegenerative diseases-Amechanistic approach, *Toxicology Letters* **230**: 85-103. DOI: 10.1016/j.toxlet.2014.01.039
- Bruce YA, Gounou S, Chabi-Olaye A, Smith H and Schulthess F (2004), The effect of neem (*Azadirachta indica* A. Juss) oil on oviposition, development and reproductive potentials of *Sesamia calamistis*

- Hampson (Lepidoptera: Noctuidae) and *Eldana saccharina* Walker (Lepidoptera: Pyralidae), *Agric Forest Entomol* **6**: 223-232. DOI:10.1111/j. 1461-9555.2004.00218.x
- Facknath S (2006), Combination of neem and physical disturbance for the control of four insect pests of stored products, *Int J Trop Insect Sci* **26**: 16-27. <https://doi.org/10.1079/IJT200698>
- Guyton K, Loomis D, Grosse Y, El Ghissassi F, Brenbrahim-Tallaa L, Guha N, Scoccianti C, Mattock H and Straif K (2015), Carcinogenicity of tetrachlorvinphos, parathion, malathion, diazinon, and glyphosate, *Lancet Oncology* **15**: 70134-70138 DOI: 10.1016/S1470-2045(15)70134-8
- Hossein MI, Ali MR, Akter MS, Akter MM and Roy SR (2018), Eco-friendly Management of Angoumois grain moth, *Sitotroga cerealella* Olivier using some botanicals on stored paddy, *Journal of Bioscience and Agriculture Research* **17**(02): 1422-1430. <https://doi.org/10.18801/jbar.170218.176>
- Isman MB (2006), Botanical insecticides, deterrents, and repellents in modern agriculture and an increasingly regulated world, *Annu Rev Entomol* **51**: 45-66. <https://doi.org/10.1146/annurev.ento.51.110104.151146>
- Jibrin MM and Mohammed AD (2020), Repellency Effect of Prepared Neem Tree Leaves Smoke against Mosquito, *Asian Journal of Research in Zoology*, **3**(3): 1-6. <https://doi.org/10.9734/ajriz/2020/v3i330090>
- Khan MA and Gumbs FA (2003), Repellent effect of Ackee (*Blighia sapida* Koenig) component fruit parts against stored product insect pests, *Journal of Tropical Agriculture* **80**: 19-27. <https://journals.sta.uwi.edu/ojs/index.php/ta/article/view/1261>
- Köhler HR and Triebkorn R (2013), Wildlife ecotoxicology of pesticides: can we track effects to the population level and beyond? *Science* **341**: 759-765. DOI: 10.1126/science.1237591
- Li L, Wei Y, van Winkle L, Zhang QY, Zhou X, Hu J, Xie F, Kluetzman K and Ding X (2011), Generation and characterization of a Cyp2f2-null mouse and studies on the role of CYP2F2 in naphthalene-induced toxicity in the lung and nasal olfactory mucosa, *J. Pharmacol. Exp. Ther.* **339**: 62–71. DOI: 10.1124/jpet.111.184671
- Lin CY, Huang FP, Ling YS, Liang HJ, Lee SH, Hu MY and Tsao PN (2015), Use of nuclear magnetic resonance-based metabolomics to characterize the biochemical effects of naphthalene on various organs of tolerant mice, *PLoS ONE*. **10**: e0120429. DOI: 10.1371/journal.pone.0120429.
- Masood KK, ur-Rashid M, Syed A, Hussain S and Islam T (2006), Comparative effect of neem (*Azadirachta indica* A. Juss) oil, neem seed water extract and baythroid against whitefly, jassids and thrips on cotton. *Pak Entomol* **28**, 31–37.
- Meyer-Baron M, Knapp G, Schäper M and van Thriel C (2015), Meta-analysis on occupational exposure to pesticides–Neurobehavioral impact and dose–response relationships, *Environmental Research* **136**: 234-245. <https://doi.org/10.1016/j.envres.2014.09.030>
- Mourão SA (2016), Toxic effects of the neem oil (*Azadirachta indica*) formulation on the stink bug predator, *Podisus nigrispinus* (Heteroptera: Pentatomidae), *Sci. Rep.* **6**: 30261. DOI: 10.1038/srep30261.
- Muñoz-Quezada MT, Lucero BA, Barr DB, Steenland K, Levy K, Ryan PB, Iglesias V, Alvarado S, Concha C, Rojas E and Vega C (2013), Neurodevelopmental effects in children associated with exposure to organophosphate pesticides: a systematic review. *Neurotoxicology* **39**: 158-168. DOI: 10.1016/j.neuro.2013.09.003
- Obeng-Ofori D, Reichmuth CH, Bekele AJ and Hasssan ali (1998), A. Toxicity and protectant potential of camphor, a major component of essential oil of *Ocimum kilimandscharicum*, against four stored product beetles, *Int. J. Pest Manag.* **44**: 203-209. <https://doi.org/10.1080/096708798228112>
- Qu X, Daizo K, Laping W and Mitsuyoshi A (2021), The losses in the rice harvest process: a review, *sustainability*. **13**(17): 9627. DOI: 10.3390/su13179627
- Stathers T, Holcroft D and Kitinoja L (2020), Ascoping review of interventions for crop postharvest loss reduction in sub-Saharan Africa and South Asia, *Nat Sustain.* **3**: 821–835. <https://doi.org/10.1038/s41893-020-00622-1>.

- Stevenson PC (2014), Using pesticidal plants for crop protection. Royal Botanic Gardens. Kew Publishers. <http://www.kew.org/discover/blogs/using-pesticidal-plants-crop-protection>
- Tadesse M (2020), Post-harvest loss of stored grain, its causes and reduction strategies, *Food Science and Quality Management*. **96**: 26-35. DOI: 10.7176/FSQM/96-04
- Yuan Y, Chen C, Zheng C, Wang X, Yang G, Wang Q and Zhang Z (2014), Residue of chlorpyrifos and cypermethrin in vegetables and probabilistic exposure assessment for consumers in Zhejiang Province, China, *Food Control* **36**: 63-68. DOI: 10.1016/j.food-cont.2013.08.008









