SUBCLINICAL MASTITIS IN LACTATING COWS: COMPARISON OF FOUR SCREENING TESTS AND EFFECT OF ANIMAL FACTORS ON ITS OCCURRENCE

M. Badiuzzaman¹, M. A. Samad¹, S. H. M. F. Siddiki³*, M. T. Islam¹ and S. Saha²

¹Department of Medicine, ²Department of Microbiology and Hygiene, Bangladesh Agricultural University, Mymensingh-2202, Bangladesh; ³Department of Medicine, Bangabandhu Sheikh Mujibur Rahman Agricultural University, Gazipur-1706, Bangladesh

ABSTRACT

A total of 444 quarter milk samples from 111 crossbred dairy cows were subjected to California mastitis test (CMT), somatic cell count (SCC) test, white side test (WST) and surf field mastitis (SFMT) test to quantify their efficacy in detecting sub clinical mastitis in dairy cows of Bangladesh during the period from 2010 to 2011. Milk samples positive by CMT, SCC, WST and SFMT were 265,230, 225 and 222 respectively. All samples were subjected to cultural isolation (gold standard test for comparison of indirect mastitis tests). Of these, 261 samples were positive by cultural isolation. The sensitivity of the CMT, SCC, WST and SFMT were 80.08%, 86.60%, 60.54% and 57.47%; specificity 69.40%, 97.81%, 63.38% and 60.66%; percentage accuracy 75.68%, 91.22%, 61.71% and 58.78%; positive predictive value 78.87%, 98.26%, 70.22% and 67.57%; and negative predictive value 70.95%, 83.64%, 52.97% and 50% respectively. Quarter-wise sub clinical mastitis (SCM) was detected in 59.68%, 51.80%, 50.68% and 50% samples by CMT, SCC, WST and SFMT, respectively, while animal-wise SCM was recorded in 72.07%, 66.67%, 64.86% and 61.26% samples by CMT, SCC, WST and SFMT, respectively. The right hind quarters were most significantly (p<0.001) susceptible to SCM than others. Kappa value of SCC was higher than that of CMT. CMT was concluded to be the most accurate test after cultural isolation and SCC. Unlike laboratory tests as cultural isolation and SCC that require adequate laboratory facilities and skilled personnel, CMT is a reliable diagnostic method in field conditions.

Keywords: Sub clinical mastitis, risk factors, bacteriological culture, California mastitis test, somatic cell count

INTRODUCTION

Bovine mastitis is the most costly disease of dairy cattle due to economic losses from reduced milk production, treatment costs, increased labour, milk withheld following treatment, death and premature culling (Miller *et al.*, 1993). It is now a well known fact that the sub clinical mastitis (SCM) is more serious and is responsible for much greater loss to the dairy industry in Bangladesh. The annual economic losses occur due to reduced milk production alone caused by SCM in Bangladesh have been estimated to be Taka 122.6 (US \$ 2.11) million (Kader *et al.*, 2003). Early detection of mastitis cow is important for most dairy farmers to reduce production losses and to enhance prospects of recovery. The diagnosis of subclinical mastitis is more problematic than clinical mastitis since the milk appears normal but usually has an elevated somatic cell count. Various methods, based on physical and chemical changes of milk and cultural isolation of organisms, are used for diagnosis of subclinical mastitis (Batra and Mcallister, 1984). The diagnosis of mastitis according to the International Dairy Federation (IDF) recommendations is based on the somatic cell counts (SCC) and microbiological status of the quarter. Bacteriological culture of milk samples is the standard method for identifying mastitis. However, the logistic and financial considerations involved with sampling all fresh cows have precluded this technique from being widely adopted. Bacteriological culture of milk identifies the presence of mastitis pathogens but does not provide a measure of the degree of inflammation associated with the infection.

Among the simplest tests, which do not require any complex laboratory equipment, are solutions containing detergents (Sargeant *et al.*, 2001). The California mastitis test (CMT), white side test (WST) and surf field mastitis test (SFMT) are arguably the only reliable cow side screening tests for sub clinical mastitis that can easily be applied. The CMT is a simple, inexpensive, rapid screening test for sub clinical mastitis, based upon the amount of cellular nuclear protein present in the milk sample. Modern mastitis tests allow for indirect determination of the number of somatic cells in milk (Greiner *et al.*, 2000). As the inflammatory process

^{*}Corresponding e-mail address: ufs.vet@gmail.com Copyright © 2015 Bangladesh Society for Veterinary Medicine

develops in udder tissue, the number of these cells (particularly leukocytes) in milk sharply increases. SCC, CMT and intra-mammary infection are associated significantly; therefore these parameters provide the necessary information to evaluate udder health status in cows.

However, the prevalence and different potential risk factors of bovine mastitis are reported by many researchers in Bangladesh (Rahman *et al.*, 1997, 2009, 2010, 2014; Rabbani and Samad, 2010; Islam *et al.*, 2011; Sarker *et al.*, 2013; Barua *et al.*, 2014) but there is a lack of reports on comparison of the indirect screening tests in detection of bovine subclinical mastitis along with the risk factors in Bangladesh. The objectives of the present study were to compare the four screening tests for the diagnosis of subclinical mastitis and to find out the relationship between animal risk factors and the occurrence of SCM in lactating cows.

MATERIALS AND METHODS

Animals

The present study was carried out on Rajshahi Dairy and Breed Development Farm, Rajabari, Rajshahi, Bangladesh during the period from 2010 to 2011. A total of 111 cross-bred dairy cows (Sahiwal × Deshi) of different age groups (3 to 11 years), parities (1 to 5) and stage of lactation (early, mid and late) were included in the study group. All cows were housed in tie stall barns and milked twice daily by hand. The regular practices of washing of udder before milking with potassium permanganate solution were used by milkers.

Sample collection

Milk samples (n=444) were collected after proper disinfection of teat surface with 70% ethanol. Sixty ml of milk from all four quarters were collected aseptically in separate sterile polyethylene screw-capped, wide mouth vials after squirting few streams (Buswell, 1995). CMT was conducted at the spot before milk sample collection. The milk samples were placed in an ice box and carried to the laboratory of Department of Medicine, Faculty of Veterinary Science, Bangladesh Agricultural University, where they were kept at 4 to 10°C in a refrigerator for further laboratory investigation.

Diagnostic tests

The milk samples were subjected to following diagnostic tests: California mastitis test (CMT), somatic cell count (SCC) test, white side test (WST) and surf field mastitis (SFMT) test. The procedure of CMT was followed in this study as per manufacturer's instruction on quarter fore-milk samples. In brief, 2 ml of milk sample was taken in the CMT paddle and equal quantity of CMT Kit (Leucocytest[®], Synbiotics Corporation-2, Lyon, France and marketed by Advance Chemical Co. Bangladesh Ltd.) was added in each cup, rotated for few seconds and the result was recorded within 30 s as 0 (negative), T (trace), 1+, 2++, or 3+++ (Ikram, 1997).

SFMT was performed and scored following the method described by Muhammad *et al.* (1995) in brief, about 2 ml milk was drawn from bottle into test cup and an estimated 2 ml reagent (Surf excel 3% solution in distilled water) was squirted from a polyethylene wash bottle. Mixing was accomplished by gentle circular motion of the paddle in a horizontal plane for few seconds. The reaction developed almost immediately with milk containing a high concentration of somatic cells. The peak of reaction was obtained within 30 seconds and immediately scored as 1+, 2+ and 3+.

The WST was performed as per procedure described by Kahir *et al.* (2008), in brief, after thorough mixing avoiding violent shaking, 50 µl (five drops) of milk were placed on a glass slide with a dark background by micropipette. Subsequently 20 µl of WST reagent (4% NaOH) were added to the milk sample and the mixture was stirred rapidly with a toothpick for 20-25 seconds. A breaking up of milk in flakes, shreds and viscid mass was indicative of positive reaction. On the other hand, milky and opaque and entirely free of precipitant was indicative of negative reaction. The SCC test was performed according to Schalm *et al.* (1971).

The milk sample was mixed thoroughly and the cream was dispersed throughout the specimens. A measured drop $(0.01\text{ml}/10\mu\text{l})$ by means of micro-pipette was then spread evenly over an area of one square centimeter on a microscopic glass slide. The milk film was allowed to air dry. Then the fat was removed from dried film in the following way: The slides were dipped in Xylene for 1 to 2 minutes. Later they were dried and immersed in 95% ethyl alcohol for 2 to 5 minutes and drain dry. Then immerse in Broadhurst-Paley stain for 5 seconds, or longer if necessary, to obtain proper intensity of staining. Then briefly rinse the slide in water and drain dry. Ten microscopic fields were counted by using oil immersion objective $(10\times100\text{mm})$ for the determination of the number of the leukocyte cells present. The required number of fields is counted by moving the slide horizontally from one edge of the film through the center to the opposite edge. The second count is made vertically through

the film in the same manner. The total number of the leukocyte cells counted is multiplied by the Working Factor (WF) of the particular microscope to obtain the no. of cells/ml of milk.

According to Schalm *et al.* (1971), the relationship of Working Factor (WF) with the number of fields to be counted using an oil immersion objective having a field diameter of 0.016 cm given below:

The following criteria are used in making the cell count:

- 1. Count all nucleated somatic cells within a field including those at the periphery with more than 50% of the cell body in view.
- 2. Free nuclei representing more than 50% of the nuclear material are counted.
- A cytoplasmic mass without nucleus and small cell fragments with little nuclear material are not counted.

Animals were considered positive for mastitis when CMT, WST and SFMT score was $\ge 1+$ and SCC value was $\ge 2 \times 10^5$ /ml of milk (threshold value). The samples (n=444) were also subjected to bacteriological culture as per Quinn *et al.* (1994).

The following diagnostic test characteristics were determined using the milk bacteriological culture result as a gold standard control.

Where: TP =True Positive, FP =False Positive, TN =True Negative, FN =False Negative.

Statistical analysis

The percentage accuracy of the tests and sensitivity, specificity, and the predictive values of the CMT, SCC, WST and SFMT results, compared to culture results, were calculated using standard two-by-two contingency tables. Correlations between the dependent variables were calculated using Pearson's correlation. Data were also analyzed by Chi-square test to observe the significant influence of parity, lactation stage on subclinical mastitis of cows.

RESULTS AND DISCUSSION

Percentage accuracy, agreement and correlation of various indirect tests

Among the 444 quarter milk samples tested for SCM in lactating cows, 265 milk samples were CMT positive and 179 samples were CMT negative. Milk samples positive by SCC, WST and SFMT were 230, 225 and 222 respectively. The accuracy of CMT, SCC, WST and SFMT was 75.68%, 91.22%, 61.71% and 58.78% (Table 1). The sensitivity, specificity, positive and negative predictive values and likelihood ratios are given in Table 2. In the present investigation, the sensitivity and specificity of the SCC were higher than those of CMT, WST and SFMT. The sensitivity and specificity of SFMT were lower than those of other. SCC showed the highest correlation (r=0.831) with bacterial culture test, followed by CMT, WST and SFMT. It is accepted that Kappa values <0.4 indicate poor agreement, values between 0.4 and 0.75: fair to good agreement and values >0.75:

excellent agreement. Thus, SCC exhibited excellent agreement with bacterial culture test while CMT had fair to good agreement (Table 2). In the present study it was apparent that SCC was the most reliable test and closest

Table 1. Percentage accuracy of various indirect tests used for the diagnosis of mastitis

Tests	Samples examined	Positive samples	TP	FP	TN	FN	Accuracy
CMT	444	265 (59.68%)	209 (78.87%)	56 (21.13%)	127 (70.95%)	52 (29.05%)	75.68%
SCC	444	230 (52%)	226 (98.26%)	4 (1.74%)	179 (83.64%)	35 (16.36%)	91.22%
WST	444	225 (50.68%)	158 (70.22%)	67 (29.77%)	116 (52.97%)	103 (47.03%)	61.71%
SFMT	444	222 (50%)	150 (67.57%)	72 (32.43%)	111 (50%)	111 (50%)	58.78%

TP=True Positive, FP=False Positive, TN=True Negative, FN=False Negative.

Table 2. Agreement and correlation of various indirect tests used for the diagnosis of mastitis with bacteriological (cultural) examination

Test	Sensitivity %	Specificity %	PPV %	NPV %	PLR	NLR	κ	r %	p
CMT	80.08	69.40	78.87	70.95	2.617	0.287	0.496	49.6	0.00
SCC	86.60	97.81	98.26	83.64	39.543	0.137	0.823	83.1	0.00
WST	60.54	63.38	70.22	52.97	1.653	0.623	0.232	23.6	0.00
SFMT	57.47	60.66	67.57	50	1.461	0.701	0.176	17.8	

PPV=Positive Predictive Value, NPV=Negative Predictive Value, PLR=Positive Likelihood Ratio, NLR=Negative Likelihood Ratio, κ=Kappa (measure of agreement), r=Pearson's Correlation.

to the bacteriological results. Present findings are in agreement with Sharma et al. (2008). They reported that SCC was the most accurate test for the diagnosis of subclinical mastitis followed by the modified California mastitis test (MCMT) and the modified White side test (MWST). Patel et al. (2000) reported higher reliability of CMT (85.69%) followed by MWST (79.74%). Reddy et al. (1998) compared the specificity and sensitivity of CMT and SCC with standard cultural test and observed 100% predictive value with the cultural test of the milk, 84.84% specificity for SCC and 73.30% for CMT. Tanwar et al. (2001) also compared various diagnostic tests for detection of subclinical mastitis and indicated 100% sensitivity for SCC and 96% for CMT reaction. According to Goswami et al. (2003), animal-wise efficacy of indirect tests taking cultural examination as a standard was in the following descending order: SCC (97.46%), MCMT (69.62%) and MWST (63.29%). So, the present findings regarding accuracy and sensitivity of indirect tests for detection of subclinical mastitis support the earlier observations. From a study of SCC and CMT sensitivity and specificity for identifying intramammary infection in early lactation Sargeant et al. (2001) concluded that CMT could be used in dairy herd monitoring program as a screening test to detect fresh cows with intramammary infection caused by major pathogens. Barbosa et al. (2002) reported that the SCC and CMT are dependent and were highly correlated for diagnosis of subclinical mastitis. Sudhan et al. (2005) reported that the percentage of agreement of CMT, SLST, bromothymol blue card test and White side test with bacteriological examination were 57.89%, 62.07%, 64.46% and 68.65% respectively.

Prevalence of subclinical mastitis in lactating cows

In the present study, quarter-wise 59.68%, 51.80%, 50.68% and 50.00% and animal-wise as 72.07%, 66.67%, 64.86% and 61.26% of samples were found positive by CMT, SCC, WST and SFMT respectively (Table 3). The

quarters were found positive for SCM of different grades. Quarter side-wise prevalence of SCM was shown in Table 4. The highest prevalence of SCM was found in Right Hind (RH) quarter by all four tests that are 72.07%, 64.86%, 63.06% and 62.16% by CMT, SCC, WST and SFMT. In RH quarter, the prevalence of subclinical mastitis was significantly higher (p<0.001) than other quarter (quarter-wise). Lowest prevalence of SCM was found in Left Front (LF) quarter that represent as 45.05%, 39.64%, 36.04%

Table 3. Prevalence and severity of subclinical mastitis in lactating cows

~-	Test used	Types	Total			Negative	Level of		
Sl. No.			No. tested	1+	2+	3+	Total	— No. (%)	significance
1.	California Mastitis Test (CMT)	Cows	111	42 (52.25)	27 (33.75)	11 (13.75)	80 (72.07)	31 (27.93)	NS
	(CIII)	Quarters	444	146 (55.09)	82 (30.94)	37 (13.96)	265 (59.68)	179 (40.32)	**
2.	Somatic cell count (SCC)	Cows	111	40 (54.05)	27 (36.487)	7 (9.47)	74 (66.67)	37 (33.33)	NS
	(555)	Quarters	444	149 (64.78)	58 (25.22)	23 (10.00)	230 (51.80)	214 (48.20)	**
3.	White Side Test (WST)	Cows	111	39 (54.17)	26 (36.11)	7 (9.72)	72 (64.86)	39 (35.14)	NS
	, ,	Quarters	444	146 (64.87)	56 (24.87)	23 (10.22)	225 (50.68)	219 (49.32)	**
4.	Surf Field Mastitis Test (SFMT)	Cows	111	37 (54.41)	25 (36.76)	6 (8.82)	68 (61.26)	43 (38.74)	NS
	(51.11)	Quarters	444	144 (64.86)	55 (24.78)	23 (10.36)	222 (50.00)	222 (50.00)	**

^{**}Significant at 1% level (p<0.01)

and 35.14% by CMT, SCC, WST and SFMT respectively. The results are in agreement with Sudhan *et al.* (2005) who reported that the right hind quarter was the most affected (38.18%) compared with the other quarters. Lee-and Lee (2007) reported that the mean Somatic Cell Score (SCS) of milk samples from the front quarters was lower than those of the milk samples from the rear quarters. Sharma *et al.* (2007) who reported single and hind quarter involvement was maximum in case of subclinical mastitis.

Prevalence of subclinical mastitis on the basis of parity

On the basis of parity the prevalence of subclinical mastitis is presented in Table 5. An increased prevalence of SCM with increased number of parity was observed in this study. The highest prevalence of SCM was found at 3rd-4th parity by all four tests that are 71.43%, 64.16%, 64.29% and 63.39% by CMT, SCC, WST and SFMT respectively (quarter-wise) and 82.14%, 75.00%, 71.43% and 71.43% by CMT, SCC, WST and SFMT respectively (animal-wise). In 3rd - 4th parity, the prevalence of subclinical mastitis was significantly higher (p<0.001) than other no. of parity (quarter wise). The lowest prevalence of SCM was found at 1st parity by all four tests that are 21.88%, 15.62%, 21.88% and 20.31% by CMT, SCC, WST and SFMT respectively (quarter-wise) and 43.75%, 43.75%, 50% and 43.75% by CMT, SCC, WST and SFMT respectively (animal-wise).

Table 4. Quarter-wise prevalence of subclinical mastitis in lactating cows

Test used	Quarter side	No. of samples	Negative No. (%)		Level of significance			
				1+	2+	3+	Total	_
California	LF	111	61(54.95)	30(27.03)	13(11.71)	7(6.31)	50(45.05)	
Mastitis Test	LH	111	36(32.43)	39(35.14)	25(22.5)	11(9.91)	75(67.57)	
(CMT)	RF	111	51(45.95)	39(35.13)	14(12.61)	7(6.31)	60(54.05)	
	RH	111	31(27.93)	38(34.23)	30(27.03)	12(10.1)	80(72.07)	**
	Total	444	179(40.39)	146(32.88)	82(18.47)	37(8.33)	265(59.68)	
Somatic cell	LF	111	67(60.36)	27(24.32)	13(11.7)	4(3.6)	44(39.64)	
count	LH	111	43(38.74)	46(41.44)	16(14.1)	6(5.41)	68(61.26)	
(SCC)	RF	111	65(58.56)	29(26.13)	12(10.81)	5(4.5)	46(41.44)	
	RH	111	39(35.14)	47(42.34)	17(15.32)	8(7.2)	72(64.86)	**
	Total	444	214(48.2)	149(33.55)	58(13.07)	23(5.18)	230(51.8)	
White Side Test	LF	111	71(63.96)	25(22.52)	11(9.91)	4(3.61)	40(36.04)	
(WST)	LH	111	46(41.44)	42(37.84)	16(14.41)	7(6.31)	65(58.56)	
	RF	111	61(54.95)	34(30.64)	12(10.81)	4(3.6)	50(45.05)	
	RH	111	41(36.94)	45(40.54)	17(15.32)	8(7.2)	70(63.06)	**
	Total	444	219(49.32)	146(32.88)	56(12.62)	23(5.18)	225(50.68)	
Surf Field	LF	111	72(64.86)	23(20.72)	12(10.82)	4(3.6)	39(35.14)	
Mastitis Test	LH	111	47(42.34)	44(39.64	14(12.61)	6(5.41)	6(57.66)	
(SFMT)	RF	111	61(54.95)	33(29.73)	12(10.81)	5(4.5)	50(45.66)	
	RH	111	42(37.84)	44(39.64)	17(15.32)	8(7.2)	69(62.16)	**
	Total	444	222(50.00	144(32.43)	55(12.39)	23(5.18)	222(50.00)	

^{**}Significant at 1% level (p<0.01)

LF=Left front quarter, LH=Left hind quarter, RF=Right front quarter, RH=Right hind quarter

This observation supports with the reports of Rasool *et al.* (1985), Devi *et al.* (1997) who reported an increasing prevalence of SCM with advancing parity. Batra and Mcallister (1984) reported that Somatic Cell Count and CMT score increased from first to fourth parity while there was general trend for increasing conductivity from second to fourth parity. Lee and Lee (2007) reported that increased SCC was observed with increase in parity.

Prevalence of subclinical mastitis on the basis of lactation stage

The prevalence of subclinical mastitis on the basis of lactation stage is shown in Table 6. It appears from the Table 6 that, all three stages of lactating cows were affected with subclinical mastitis. The highest prevalence of SCM was found at early lactation of stage by all four tests that are 78.43%, 76.47%, 70.59% and 68.63% by CMT, SCC, WST and SFMT respectively (animal-wise) and 70.58%, 64.22%, 63.24% and 61.76% by CMT, SCC, WST and SFMT respectively (quarter-wise). In early stage of lactation, the prevalence of subclinical mastitis was significantly higher (p<0.001) than other stage of lactation (quarter-wise).

The lowest prevalence of SCM was found at mid and late lactation stage by CMT, SCC, WST and SFMT tests that are 65.63%, 56.25%, 59.37% and 53.57% (animal-wise) and 49.22%, 39.06%, 39.06%, and 39.29 % (quarter-wise). This observation supports with the reports of Jyoti *et al.* (1998) who reported 77.77%, 63.76% and 41% prevalence of SCM in early, mid and late lactation respectively in an organized farm. Rahman *et al.* (1997) reported that highest prevalence of SCM during the 3rd months (34.00%) of lactation. Lalintluanga *et al.* (2003) reported that mastitis incidence was higher during the early stage of the 3rd lactation (36.60%). However, SCC is found out to increase in the first few days of lactation and may be high up to the first month of lactation (Atakan, 2008) and increase towards the end of lactation is considered to be physiological. Sederevicius *et al.* (2006) reported a temporary increase in SCC just after calving due to adaptation of the udder from non-lactating to lactating status, while in mid lactations SCC usually remains in normal range.

Table 5. Effect of parity on the prevalence of subclinical mastitis in lactating cows

			Animal-wise		Level of	Total	Quarter-wis	e	Level of	
Test	Parity	Total animal	Positive No. (%)	Negative No. (%)	significance	quarter	Positive No. (%)	Negative No. (%)	- significance	
	1 ST	16	7(43.75)	9(56.25)		64	14(21.88)	50(78.12)		
	2^{nd}	17	10(58.82)	7(41.18)		68	30(44.12)	38(55.88)		
	3^{rd}	30	24(80.00)	6(20.00)		120	85(70.83)	35(29.17)		
CMT	4^{th}	28	23(82.14)	5(17.86)		112	80(71.43)	32(28.57)		
	5 th	20	16(80.00)	4(20.00)	NS	80	56(70.00)	24(30.00)	**	
	Total	111	80(72.07)	31(27.93)		444	265(59.68)	179(40.32)		
	1 ST	16	7(43.75)	9(56.25)		64	10(15.62)	54(84.38)		
	2^{nd}	17	9(52.94)	8(47.06)		68	24(35.29)	44(64.71)		
	3^{rd}	30	22(73.33)	8(26.67)		120	77(64.16)	43(35.84)		
SCC	4^{th}	28	21(75.00)	7(25.00)		112	70(62.50)	42(37.50)		
	5 th	20	15(75.00)	5(25.00)	NS	80	49(61.25)	31(38.75)	**	
	Total	111	74(66.67)	37(33.33)		444	230(51.8)	214(48.20)		
	1 ST	16	8(50.00)	8(50.00)		64	14(21.88)	54(78.12)		
	2^{nd}	17	10(58.82)	7(41.18)		68	28(41.18)	40(58.82)		
WST	3^{rd}	30	20(66.67)	10(33.33)		120	68(56.67)	52(43.33)		
	4^{th}	28	20(71.43)	8(28.57)	NS	112	72(64.29)	40(35.71)		
	5 th	20	14(70.00)	6(30.00)		80	43(53.75)	37(46.25)	**	
	Total	111	72(64.86)	39(35.14)		444	225(50.68)	219(49.32)		
	1 ST	16	7(43.75)	9(56.25)		64	13(20.31)	51(79.69)		
	2^{nd}	17	9(52.94)	8(47.06)		68	29(42.65)	39(57.35)		
	3^{rd}	30	18(60.00)	12(40.00)		120	65(54.17)	55(45.83)		
SFMT	4^{th}	28	20(71.43)	8(28.57)		112	71(63.39)	41(36.61)	**	
	5 th	20	14(70.00)	6(30.00)	NS	80	44(55.00)	36(45.00)		
	Total	111	68(61.26)	43(38.74)		444	222(50.00)	222(50.00)		

^{**}Significant at 1% level (p<0.01)

Table 6. Effect of stage of lactation on the prevalence (animal-wise and quarter-wise) of subclinical mastitis in lactating cow

	Lactation	Total	Animal-w	rise	Level of	Total	Quarter-v	vise	Level of
Test	stage	animal	Positive	Negative	significance	quarter	Positive	Negative	significance
			No. (%)	No. (%)			No. (%)	No. (%)	
	Early	51	40	11		204	144	60	
	(6-90d)		(78.43)	(21.57)			(70.58)	(29.42)	**
CMT	Mid	28	19	9		112	58	54	
	(91-180d)		(67.86)	(32.14)	NS		(51.79)	(48.21)	
	Late	32	21	11		128	63	65	
	(>180d)		(65.63)	(34.37)			(49.22)	(50.78)	
	Total	111	80	31		444	265	179	
			(72.07)	(27.93)			(59.68)	(40.32)	
	Early	51	39	12		204	131	73	
	(6-90d)		(76.47)	(23.53)			(64.22)	(35.78)	**
SCC	Mid	28	17	11		112	49	63	
	(91-180d)		(60.71)	(39.29)	NS		(43.75)	(56.25)	
	Late	32	18	14		128	50	78	
	(>180d)		(56.25)	(43.75)			(39.06)	(60.93)	
	Total	111	74	37		444	230	214	
			(66.67)	(33.33)			(51.80)	(48.20)	
	Early	51	36	15		204	129	75	
	(6-90d)		(70.59)	(29.41)			(63.24)	(36.76)	**
WST	Mid	28	17	11	NS	112	46	66	
	(91-180d)		(60.71)	(39.29)			(41.07)	(58.93)	
	Late	32	19	13		128	50	78	
	(>180d)		(59.37)	(40.63)			(39.06)	(60.94)	
	Total	111	72	39		444	225	219	
			(64.86)	(35.14)			(50.68)	(49.32)	
	Early	51	35	16		204	126	78	
	(6-90d)		(68.63)	(31.37)			(61.76)	(38.24)	**
SFMT	Mid	28	15	13		112	44	68	
	(91-180d)		(53.57)	(46.43)	NS		(39.29)	(60.71)	
	Late	32	18	14		128	52	76	
	(>180d)		(56.25)	(43.75)			(40.63)	(59.37)	
	Total	111	68	43		444	222	222	
			(61.26)	(38.74)			(50.00)	(50.00)	

^{**}Significant at 1% level (p<0.01)

In conclusion, SCC was the most accurate test after cultural isolation, followed by CMT, WST and SFMT. Taking into consideration that laboratory tests such as cultural isolation and SCC require adequate laboratory facilities, personnel and time, CMT is a reliable diagnostic method for use as a regular mastitis screening test in field conditions even by less trained dairymen.

ACKNOWLEDGEMENTS

Authors are grateful to Md. Mizanur Rahman, Manager, Rajshahi Dairy and Breed Development Farm, Rajabari, Rajshahi and the farmers of the adjacent villages of Bangladesh Agricultural University, Mymensingh for their assistance and co-operation to conduct the research.

REFERENCES

1. Atakan KOC (2008). A study of somatic cell counts in the milk of Holstein-Friesian cows managed in Mediterranean climatic condition. *Turkish Journal of Veterinary and Animal Sciences* 32: 13-18.

- 2. Barbosa CB, Benadetti E, Ribeiro SC, Guimaraes EC and Ribeiro SC (2002). The relationship between somatic cell count (SCC) and result of the California Mastitis Test (CMT) to diagnose bovine mastitis. Bioscience Journal 18: 93-102.
- 3. Barua M, Prodhan MAM, Islam K, Chowdhury S, Hasanuzzaman M, Imtiaz MA and Das GB (2014). Subclinical mastitis prevalent in dairy cows in Chittagong district of Bangladesh: detection by different screening test. *Veterinary World* 7: 483-488.
- Batra TR and McAllister AJ (1984). A comparison of mastitis detection methods in dairy cattle. Canadian Journal of Animal Sciences 64: 305-310.
- 5. Buswell J (1995). Simple mastitis bacteriology for the practice. *In Practice* 17: 426-432.
- Devi BK, Shukla PC and Bagherwal RK (1997). Incidence of subclinical mastitis in cows. *Indian Journal Dairy Science* 50: 477-478.
- 7. Goswami SN, Roy A and Kalyani IH (2003). A comparative study on various indirect tests to direct cultural isolation for detection of subclinical mastitis (SCM). In: *Proceedings of XXI Indian Society for Veterinary Medicine (ISVM) Conference*, Anand, India. pp. 101-102.
- 8. Greiner M, Pfeiffer D and Smith RD (2000). Principles and practical application of the receiver-operating characteristics analysis for diagnostic tests. *Preventive Veterinary Medicine* 45: 23-41.
- 9. Ikram M (1997). Diagnostic microbiology. In: W Paul and VMD Pratt (editors), Laboratory Procedures for Veterinary Technicians. RR Donnelley and Sons Company. St. Louis, Missouri159-160.
- 10. Islam MA, Islam MZ, Islam MA, Rahman MS and Islam MT (2011). Prevalence of subclinical mastitis in dairy cows in selected areas of Bangladesh. *Bangladesh Journal of Veterinary Medicine* 9: 73-78.
- 11. Joyoti B, Dutta GN and Buragohain J (1998). Control of bovine mastitis through treatment of subclinical mastitis during lactation. *Indian journal Dairy Science* 51: 309-314.
- 12. Kader MA, Samad MA and Saha S (2003). Influence of host level factors on prevalence and economics of subclinical mastitis in dairy cows in Bangladesh. *Indian Journal of Dairy Science* 56: 235-240.
- 13. Kahir MA, Islam MA, Rahman AKMA, Nahar A, Rahman MS and Song HJ (2008). Prevalence and risk factors of subclinical bovine mastitis in some dairy farms of Sylhet district of Bangladesh. *Korean Journal of Veterinary Services* 31: 497-504.
- 14. Lalrintluanga C, Ralte EC and Hmarkunga (2003). Incidence of mastitis, bacteriology and antibiogram in dairy cattle in Aizawl, Mizoram. Disease Investigation Laboratory, AH and Veterinary Department, Mizoram, India. *Indian Veterinary Journal* 80: 931-932.
- 15. Lee JC and Lee CY (2007). Study on the relationship between udder and teat characteristics and somatic cell count. *Journal of Veterinary Clinics* 24: 172-176.
- 16. Miller GY, Bartlett PC, Lance SE, Anderson J and Heider LE (1993). Costs of clinical mastitis and mastitis prevention in dairy herds. *Journal of the American Veterinary Medical Association* 202: 1230-1236.
- 17. Muhammad G, Athar M, Shakoor A, Khan MZ, Fazal-ur-Rehman and Ahmad MT (1995). Surf Field Mastitis Test: An inexpensive new tool for evaluations of wholesomeness of fresh milk. *Pakistan Journal of Food Science* 5: 91-93.
- 18. Patel PR, Raval SK, Rao N, Mandali GC and Jani RG (2000). Status of mastitis in Gujarat State. In: *Proceedings of Round Table Conference of the Indian Association for the Advancement of Veterinary Research (IAAVR) on Mastitis*, IVRI, Izatnagar, India, pp. 45-52.
- 19. Quinn PJ, Carter ME, Markey BK and Carter GR (1994). Clinical Veterinary Microbiology. 1st edn., Mosby-Year book Europe, Ltd., Wolfe, Spain.
- 20. Rabbani AFMG and Samad MA (2011). Host determinants based comparative prevalence of subclinical mastitis in lactating Holstein-Friesian cross cows and Red Chittagong cows in Bangladesh. *Bangladesh Journal of Veterinary Medicine* 8: 17-21.
- 21. Rahman MA, Bhuiyan MMU, Kamal MM and Shamsuddin M (2009). Prevalence and risk factors of mastitis in dairy cows. *Bangladesh Veterinary Journal* 26: 54-60.
- 22. Rahman MM, Islam MR, Uddin MB and Aktaruzzaman M (2010). Prevalence of subclinical mastitis in dairy cows reared in Sylhet District of Bangladesh. *International Journal of Biological Research* 1: 23-28.
- 23. Rahman MM, Munsi N, Kabir H, Ekram F, Rahman MT and Saha S (2014). Prevalence of subclinical mastitis in cows at Anwara, a coastal Upazila of Chittagong district in Bangladesh. *Journal of Veterinary Advances* 4: 594-508
- 24. Rahman MS, Nooruddin M and Rahman MM (1997). Prevalence and distribution of mastitis in cross-bred and exotic dairy cows. *Bangladesh Veterinary Journal* 14: 1-4.

- 25. Rasool G, Jabbar MA, Kazi SE and Ahmed A (1985). Incidence of subclinical mastitis in Nili-Ravi buffaloes and Sahiwal cows. *Pakistan Veterinary Journal* 5: 76-78.
- 26. Reddy LV, Choudhuri PC and Hamza PA (1998). Sensitivity, specificity and predictive values of various indirect tests in the diagnosis of subclinical mastitis. *Indian Veterinary Journal* 75: 1004-1005.
- 27. Sargeant JM, Leslie KE, Shirley JE, Pulkrabek BJ and Lim GH (2001). Sensitivity and specificity of somatic cell count and California mastitis test for identifying intramammary infection in early lactation. *Journal of Dairy Science* 84: 2018-2024.
- 28. Sarker SC, Parvin MS, Rahman AK and Islam MT (2013). Prevalence and risk factors of subclinical mastitis in lactating dairy cows in north and south regions of Bangladesh. *Tropical Animal Health and Production* 45: 1171-1176
- 29. Schalm OW, Carrol EJ and Jain NC (1971). Bovine Mastitis. 1st edn., Lea and Febiger, Philadelphia, U.S.A.
- 30. Sederevicius A, Balsyte J, Lukauskas K, Kazlauskaite J and Biziulevicius GA (2006). An enzymatic cow immunity-targeted approach to reducing milk somatic cell count: A comparative field trial. *Food and Agricultural Immunology* 17: 1-7.
- 31. Sharma N, Maiti SK and Pandey V (2008). Sensitivity of indirect tests in the detection of subclinical mastitis in buffaloes. *Veterinary Practitioner* 9: 29-31.
- 32. Sharma N, Maiti SK and Sharma KK (2007). Prevalence, etiology and antibiogram of microorganisms associated with subclinical mastitis in buffaloes in Durg, Chhattisgarh State (India). *International Journal of Dairy Science* 2: 145-151.
- 33. Sudhan NA, Singh R, Singh M and Soodan JS (2005). Studies on prevalence, etiology and diagnosis of subclinical mastitis among cross bred cows. *Indian Journal of Animal Research* 39: 127-130.
- 34. Tanwar RK, Vyas SK, Fakhruddin and Singh AP (2001). Comparative efficacy of various diagnostic tests in diagnosis of SCM in Rathi cows. In: *Proceedings of Round Table Conference of the Indian Association for the Advancement of Veterinary Research (IAAVR) on Mastitis* 161-163.