

GUT BACTERIAL DIVERSITY AND THEIR ANTIBIOTIC RESISTANCE IN HONEY BEE COMMUNITIES OF DHAKA CITY

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Abstract: The study was undertaken to isolate and identify the bacteria from the gut of honey bees and to investigate their antibiotic sensitivity. A total 35 honey bees were collected from Sher-e-Bangla Agricultural University and Curzon Hall area of Dhaka city. This study was conducted to identify the gram-positive and gram-negative bacteria based on the morphological features using microscope. In this study, four species of honey bees were identified, of which, *Apis dorsata* was the most dominant (31.43%) species and followed by 17.14% for *Apis cerana* and 2.86% for *Apis mellifera* and *Apis florea*, respectively. In total, five gram-positive bacteria genera including *Lactobacillus* sp. (37.5%), *Streptococcus* sp. (9.38%), *Staphylococcus* sp. (21.88%), *Enterococcus* sp. (6.25%), and *Clostridium* sp. (9.38%); and two gram-negative bacteria i.e., *Pseudomonas* sp. (3.13%), and *Enterobacter* sp. (12.5%) were detected. Gram-positive bacteria were the most dominant at the honey bee gut in two sites, while bacteria did not detect in *A. dorsata* gut which was collected from Sher-e-Bangla Agricultural University. However, five types of bacteria were isolated from the gut of *A. dorsata* collected from Curzon Hall area. In total, the highest varieties (6 out of 7) and number of isolates (15) were found from *A. florea*. Whereas, the highest number of bacteria was isolated from *A. florea* (12) for Curzon Hall area, and *A. cerana* (6) for Sher-e-Bangla Agricultural University. In antibiotic susceptibility test, the isolated *Streptococcus* sp., *Staphylococcus* sp. and *Enterococcus* sp. were sensitive to the majority of the antibiotics. However, the isolated *Lactobacillus* sp., *Clostridium* sp., *Enterobacter* sp. and *Pseudomonas* sp. were resistant to the majority of antibiotics. The results show that the presence or absence of bacterium in honey bees gut was an indicator for their overall health.

Key words: Antibiotic sensitivity, Bacteria isolation, Probiotic bacteria, *Apis dorsata*, *Apis cerana*, *Apis mellifera*, *Apis florea*.

INTRODUCTION

Honey bees are flying insects within the genus *Apis* under the largest

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family Apidae of order Hymenoptera (Danforth *et al.* 2006). In Bangladesh, only one genus of honey bee *Apis* is being found which includes four species: *Apis mellifera*, *Apis dorsata*, *Apis cerana*, and *Apis florea* (Akter *et al.*, 2019). For the past few decades, their number has been declined throughout the world alarmingly (Potts *et al.*, 2010).

Various types of microbes are associated with the animal kingdom and some of them are beneficial and some only cause nuisance and diseases (Hacquard *et al.*, 2015). The honey bees gut bacteria can be a pathogenic and or symbiotic. The presence of pathogenic bacteria has been significantly affected honey bee overall health while the symbiotic association with some microbes are obligatory for their survival (Engel *et al.*, 2016; Anjum *et al.*, 2006). The microbiota associated with the honey bee is mainly composed of yeasts, gram-positive bacteria (such as *Lactobacillus rigidus apis*, *L. constellatus*, *Bacillus* spp., *Streptococcus* and *Clostridium*), and gram-negative or gram variable bacteria (*Achromobacter*, *Citrobacter*, *Enterobacter*, *Erwinia*, *Escherichia coli*, *Flavobacterium*, *Klebsiella*, *Proteus* and *Pseudomonas*) (Gilliam, 1978; Gilliam and Prest, 1987; Rada *et al.*, 1997; Mohr and Tebbe, 2006). Some gram-positive bacteria in honey bee gut i.e., *Lactobacillus* and *Streptococcus* are considered to be beneficial bacterial species which are immensely used as the important probiotics in the dairy products (Isolauri *et al.*, 2002).

Some gram-positive bacteria i.e., *Clostridium* and *Bacillus* and gram-negative bacteria i.e., *Enterobacter* and *Pseudomonas* are considered as pathogen in honey bee population. These are responsible for diseases and honey bee colony collapsing (Evans and Schwarz, 2011). According to Anjum *et al.* (2021) most of these pathogenic bacteria are also pathogenic to human which has substantially caused honey bee mortality thus honey bee population has been declined gradually. The study on honey bee gut microbiota and their antibiotic resistance in Bangladesh is not available. Therefore, the present study was conducted to identify the gut bacterial diversity and their antibiotic resistance in honey bee community. Additionally, this study was help to understand the overall health status of honey bee population in the Dhaka city.

MATERIAL AND METHODS

This research work was conducted in Entomology Laboratory, Department of Zoology and Microbiology Laboratory, Department of Microbiology, University of Dhaka. Curzon Hall area of University of Dhaka. The Sher-E-Bangla Agricultural University campus was selected as the collection site of honey bees. Honey bees were collected by sterilizing insect net and hand picking. After catching by insect net, they were transferred to air tight jar. Inside the jar there

was some cotton which was mixed with chloroform for anesthetizing. Collected bees were placed into the sterile tube individually inflicting minimal injury to them. Identification was made by examining the external morphology of the bees individually and following standard taxonomic keys (Engel, 1999). After identification, fresh honey bee samples were soaked in 70% ethanol for five minutes to decontaminate their external surface and dried, followed by washing with sterile saline to remove traces of ethanol. The alimentary tract of honey bees was aseptically dissected out using autoclaved sterilized entomological dissecting needles under a dissecting microscope. The instrument was dipped in ethanol and flamed between dissections. The excised gut was homogenized in 9 ml of sterile normal saline water. Then it was diluted. In total, 35 honey bees were collected for the experiment. All collected homogenate were cultured using spread plate technique into Nutrient Agar (NA) media and M-FC agar media followed by incubation at 37°C for 24 hours. Growth on all plates was observed and the numbers of bacteria colonies were counted. Isolated colonies were cultured using NA media to obtain pure culture. Moreover, the pure isolated colonies were then cultured using Mac Conkey media and Mannitol salt agar and M-FC agar for further identification. These were then transferred in the incubation chamber to developed colonies, which were identified based on morphological features of each colony (size, shape, elevation, color, consistency, opacity, pigmentation).

Steps for gram staining were followed by Pelczar *et al.* (1993) to determine the size, shape, arrangement and gram reaction to isolated organisms. The presumptive identification of isolates was done by performing various biochemical tests i.e., motility test, indole test, citrate utilization test, lactose fermentation test and glucose fermentation test, gas formation test. H₂S in Kingler's Iron Agar (KIA) were performed for gram-negative bacteria and catalase test and oxidase test were performed for gram-positive bacteria. All the tests were accomplished by following the standard protocol as described in Bergey's Manual of Systematic Bacteriology (Garrity, 2001). Antibiotic susceptibility testing of bacterial isolates was done by Kirby Bauer disk diffusion method using Mueller Hinton Agar (MHA) plate. Commercial antimicrobial discs were used which include: Ampicillin (AMP 25), Penicillin-G (P), Chloramphenicol (C), Tetracycline (TE), Amoxicillin (AML), Imipenem (IMI), Erythromycin (E).

RESULTS AND DISCUSSION

A total of 35 honey bees of four different species were identified from Sher-e-Bangla Agricultural University and Curzon Hall, which were then used to examine the bacteriological study. In this study, in total, four species of honey bees were identified i.e., *A. dorsata*, *A. florea*, *A. cerana* and *A. mellifera*; these

Table1: Honey bee species distribution in two study areas

Honey bee species	Sher-E-Bangla Agricultural University	Curzon hall	Total
<i>Apis dorsata</i>	2 (5.71%)	9 (25.71%)	11(31.42%)
<i>Apis florea</i>	7(20%)	10 (28.57%)	17 (48.57%)
<i>Apis cerana</i>	6 (17.14%)	0	6 (17.14%)
<i>Apis mellifera</i>	1(2.86%)	0	1(2.86%)
Total	16(45.71%)	19(54.28%)	35

Table 2: Number of isolates among different species of honey bees

Study area	Honey bee species	No. of Isolates	Percentage of isolates
Sher-E-Bangla Agricultural University	<i>Apis dorsata</i>	0	0%
	<i>Apis florea</i>	3	9.38%
	<i>Apis cerana</i>	6	18.75%
	<i>Apis mellifera</i>	1	3.12%
Curzon hall	<i>Apis florea</i>	12	37.5%
	<i>Apis dorsata</i>	10	31.25%
Total		32	100%

were recorded in Sher-e-Bangla Agricultural University while two honey bee species *A. florea* and *A. dorsata* were observed in Curzon Hall area. On the basis of percentage composition, honey bees constituted of 54.28% and 45.71% total population in Curzon Hall and Sher-e-Bangla Agricultural University, respectively. Among the four species of Honey bees, *A. florea* (48.57%) was the most dominant species and followed by *A. dorsata* (31.43%) and *A. cerana* (17.14%), while *A. mellifera* (2.86%) was least dominant species (Table 1). The bacteriological characteristics of 35 honey bees were investigated. A total of 32 bacterial isolates were prepared from the bees. Among 12(37.5%) were collected from *A. florea* and 10(31.25%) from *A. dorsata* of Curzon Hall area. Additionally, 3(9.38%) isolates were collected from *A. florea*, 1(3.12%) from *A. mellifera* and 6(18.75%) from *A. cerana* of Sher-E-Bangla Agricultural University campus. Not any type of microorganism was found from the gut of *A. dorsata* collected from Sher-e-Bangla Agricultural University. (Table-2). Based on biochemical tests and growth on selective media, the isolates were presumptively identified as *Lactobacillus* sp. (37.5%), *Streptococcus* sp. (9.38%), *Staphylococcus* sp. (21.88%), *Enterococcus* sp. (6.25%), *Clostridium* sp. (9.38%), *Pseudomonas* sp. (3.13%), and *Enterobacter* sp. (12.5%) (Fig. 1). Seven types of bacteria species were isolated, of which. five (*Lactobacillus* sp., *Streptococcus* sp., *Staphylococcus* sp., *Enterococcus* sp., and *Clostridium* sp.) were gram-positive while the two (*Pseudomonas* sp. and *Enterobacter* sp.) were gram-negative bacteria. Among the isolated gram-positive bacteria, *Lactobacillus* sp. was more frequent and among the gram-negative, *Enterobacter* sp. was the most dominant. *Pseudomonas* sp. and *Enterococcus* sp. did not find from the samples of Curzon Hall. Similarly,

Table 3: Biochemical test result of Gram-negative isolates collected from 2 selected areas

Study area	Honey bee species	Sample no.	Isolate no.	Growth on			Gram stain	Biochemical test						Presumptive Identification		
				MacConkey	MFC			Lact. Iron Agar test	Kingler's test	Gluc.	Gas	H2S	Citrate use		Motility test	Indole production
Sher-E-Bangla Agricultural University	<i>Apis florea</i>	S-8	I	+	+	+	GN-rod	+	+	-	+	+	-	+	+	<i>Enterobacter</i> sp.
		S-9	K	+	+	+	GN-rod	-	+	-	+	+	+	-	-	<i>Pseudomonas</i> spp
Curzon hall	<i>Apis dorsata</i>	C17	H, J	+	+	+	GN-rod	+	-	-	+	-	-	+	+	<i>Enterobacter</i> sp.
		C10	N2	+	+	+	GN-rod	+	-	-	+	-	-	+	+	<i>Enterobacter</i> sp.

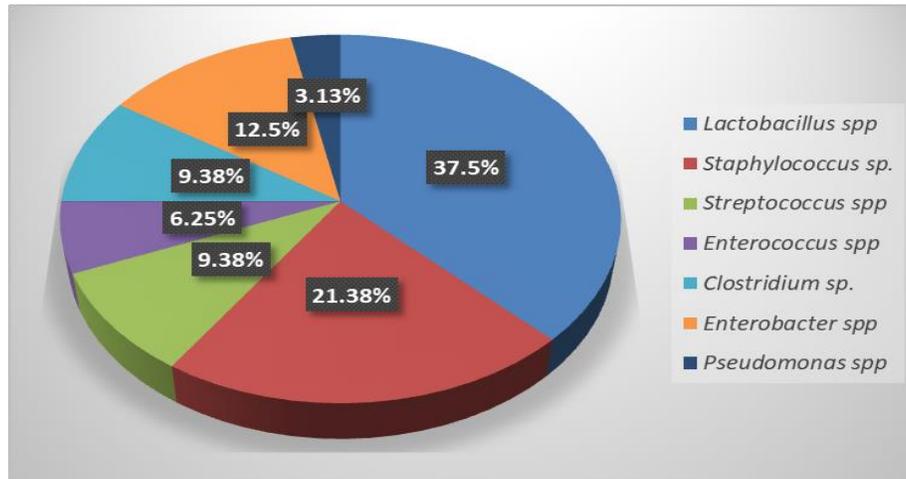


Fig.1. Percentage of different bacteria found in honey bee species

Clostridium sp. and *Staphylococcus sp.* were not detected among the samples of Sher-E-Bangla Agricultural University campus. (Table 3 and Table 4). It was investigated that *A. cerana* carried *Lactobacillus sp.* and *Enterococcus sp.* (Table 4). However, Disayathothonooat *et al.* (2012) isolated bacteria from the gut of *Apis cerana* and found *Klebsiella* and *Enterobacter sp.* It was also found that *A. mellifera* contained only *Enterococcus sp.* (Table 4). This result match with the study investigated by Audisio *et al.* (2010), who found five strain *Enterococcus* from the gut of *A. mellifera*. *A. dorsata* contained *Lactobacillus sp.*, *Streptococcus sp.*, *Staphylococcus sp.*, *Clostridium sp.*, and *Enterobacter sp.* (Table 4). In case of *A. dorsata* the study result was consistent with the study of Tajabad *et al.* (2012) where *Lactobacilli sp.* was isolated from the honey bee stomach. Most of these bacteria are act as the causative agent of various diseases in human and the presence of human pathogenic bacteria is a reason for the reduction of bee community, as these pathogens affect host overall health (Ishii *et al.*, 2014). Malignant external otitis, endophthalmitis, endocarditis, meningitis, pneumonia, and septicemia are the results of *Pseudomonas sp.*, while *Staphylococcus aureus* can cause skin infection and food poisoning (Bodey *et al.*, 1983, Forster *et al.*, 2009). *Enterococcus sp.* is associated with urinary tract and biliary tract infections and *Streptococcus sp.* are responsible for endocarditis, respiratory and urinary tract infections (Patterson, 1996). Various bacteria such as *Lactobacillus sp.*, *Enterococcus sp.* and *Streptococcus sp.* etc. are notably known as probiotic for honey bee (Ljungh and Wadström, 2006; Du, *et al.*, 2021; Hutkins and Goh, 2014). According to Poormuntaseri *et al.* (2014), *Clostridium sp.* do not cause

Table 4: Biochemical test result of Gram-positive isolates collected from 2 selected areas

Study area	Honey bee species	Sample no	Isolate no	Growth on			Biochemical test		Presumptive Identification
				MacCon	Key	MSA	MFC	Catalase	
Sher-E-Bangla Agricultural University	<i>Apis florea</i>	S-3	D	-	+	-	-	-	<i>Streptococcus</i> sp.
		S-10, 12, 13, 14, 15	P, Q1, T, V, X	-	+	-	-	-	<i>Lactobacillus</i> sp.
	<i>Apis cerana</i>	S-12	Q2	-	+	-	-	-	<i>Enterococcus</i> sp.
		S-16	Z	-	+	-	-	-	<i>Enterococcus</i> sp.
	<i>Apis mellifera</i>	C-1	A1	-	+	-	-	-	<i>Streptococcus</i> sp.
		C-4, 8, 9	A2, F, L, M	-	+	-	-	-	<i>Lactobacillus</i> sp.
		C-2	B1	-	+	-	-	+	<i>Lactobacillus</i> sp.
	<i>Apis florea</i>	C-2, 3, 7, 10	B2, E, G1, G2, N1	-	+	-	-	+	<i>Clostridium</i> sp.
		C-12, 13	O, S	-	+	-	-	+	<i>Staphylococcus</i> sp.
	Curzon hall	<i>Apis dorsata</i>	C-12, 16	R, b	-	-	-	-	-
C-14			U, W, Y	-	-	-	-	+	<i>Lactobacillus</i> sp.
C-16			a	-	-	-	-	-	<i>Streptococcus</i> sp.

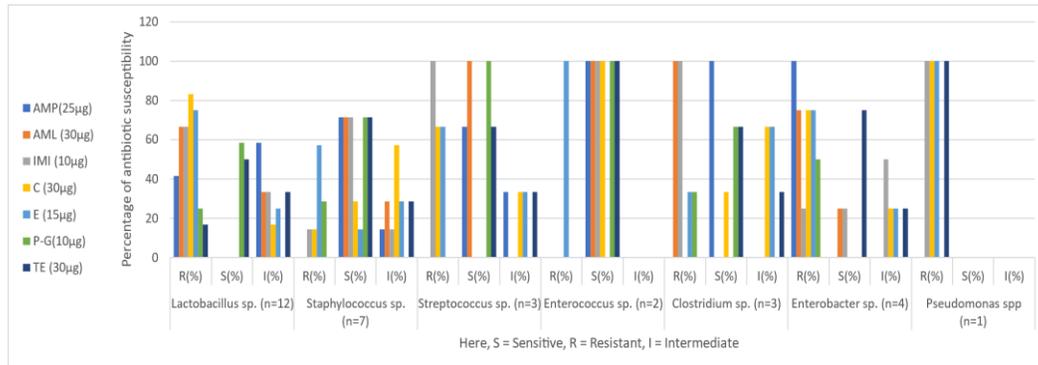


Fig. 2. Percentage of antibiotic susceptibility of different isolated bacteria species.

any disease in bee, but the reason behind botulism in humans, this gram-positive bacterium affects mainly children and those with weakened immune systems. *Enterobacter* sp. are responsible for causing many nosocomial infections, including urinary tract infections (UTI), respiratory infections, soft tissue infections in human and also cause bacterial disease in honey bee (Ramirez and Giron, 2021). Human depends on bee and bee products include honey, which could be contaminated by various sources, but the primary source of microbial contamination is the digestive tracts of honeybees (Peter et al., 2007). Identified isolates from gut of bees were tested for their antibiotic susceptibility against seven commonly used antibiotics. Most of the *Lactobacillus* isolates were resistant to those antibiotics. The highest resistance pattern (83.33%) was found against Chloramphenicol and the lowest (16.67%) was found against Tetracycline antibiotic. It was found that 100% *Pseudomonas spp.* were resistant to Chloramphenicol, Imipenem, Erythromycin and Tetracycline, while most of the individual of other isolates found sensitive to Tetracycline. Among the four isolates of *Enterobacter* spp., highest (75%) susceptibility was observed to Tetracycline and highest (100%) resistance was found against Ampicillin. By contrast, among three isolates of *Clostridium* spp. all of them were resistant (100%) against Amoxicillin and Imipenem and 100% sensitive to Ampicillin (Fig. 2). It was observed that, all of the two isolates of *Enterococcus* sp. were resistant (100%) against these antibiotics, except in Erythromycin, where the isolates found 100% sensitive. Most of the isolates of *Staphylococcus* spp. were sensitive. Among them, they were mostly sensitive (85.71%) to Ampicillin and most resistant (57.14%) against Erythromycin antibiotic. But according to Akter et al., (2021), most of the isolates of *Staphylococcus* spp. found resistant. were equally susceptible and resistant against these antibiotics. All of the three isolates (100%) *Streptococcus* sp. were resistant against Imipenem and sensitive to Amoxicillin and Penicillin-G antibiotic (Fig. 2).

As some bacteria in bee affects host nutrition, weight gain, endocrine signaling, immune function, and pathogen resistance (Zheng *et al.*, 2019), it is important to investigate whether these pathogens create any impact on other beneficial and obligatory microbes present in bee gut. Honey bee health could be improved by using probiotic (Kaznowski, 2005) and in this study we found three natural probiotics namely, *Lactobacillus* sp., *Enterococcus* sp. and *Streptococcus* sp. They assist in healthy replacement of good bacteria in the intestinal tract following antibiotic therapy (Sanders, *et al.*, 2000). Therefore, it can be said that the *Lactobacillus* sp. which originates from the honey stomach can be selected as a food probiotic and food preservatives supplement for human consumption.

LITERATURE CITED

- AKTER T., AKHTER S., SULTANA, J. A. JHORNA and BEGUM, S. 2019. The bees (Apocrita: Hymenoptera) of Dhaka city, Bangladesh. *J. biodivers. Conserve. Bioresour. Manag.* **5**(1):113-119.
- AKTER T., JAHAN, S., AHMED, S., SULTANA, S. and BEGUM, S. 2021. Isolation of multi-drug resistant potential pathogenic bacteria from blow fly collected from different areas of dhaka city. *Bangladesh J. Zool.* **49** (2): 205-214.
- ANJUM, S. I., ALDAKHEEL, F., SHAH, A. H., KHAN, S., ULLAH, A., HUSSAIN, R., KHAN, H., ANSARI, M. J., MAHMOUD, H. A. and MOHAMMED, O. B. 2021. Honey bee gut an unexpected niche of human pathogen. *J King Saud U Sci.ens.* **33** (1).
- AUDISIO, M. C., TORRES, M. J., SABATE, D. C., IBARGUREN, C. and APELLA, M. C. 2011. Properties of different lactic acid bacteria isolated from *Apis mellifera* L. bee-gut. *Microbiol. Res.* **166**: 1-13.
- BODEY, G. P., BOLIVAR, R., FAINSTEIN, V. and JADEJA, L. 198. 3Infections caused by *Pseudomonas aeruginosa*. *Rev. Infect. Dis.* **5**(2):279-313.
- DANFORTH, B. N., SIPES, S., FANG, J. and BRADY, S. G. 2006. The history of early bee diversification based on five genes plus morphology. *PNAS.* **103** (41): 15118-15123.
- DISAYATHANOOWAT, T., YOSHIYAMA, M., KIMURA, K. and CHANTAWANNAKUL, P. 2012. Isolation and characterization of bacteria from the midgut of the Asian honey bee (*Apis cerana indica*). *J. Apic. Res.* **51**(4): 312-319.
- DU, Y., LUO, S. and ZHOU, X. 2021. *Enterococcus faecium* Regulates Honey Bee Developmental Genes. *Int. J. Mol. Sci.* **22**(22).
- EL-LEITHY, M. and EL-SIBAEL, K. 1992. Role of microorganisms isolated from bees, its ripening and fermentation of honey. *Egypt. J. Microbiol.* **75**:679-681.
- ENGEL, M. S. 1999. The Taxonomy of Recent and Fossil Honey Bees (Hymenoptera: Apidae; *Apis*). *J. Hymenopt. Res.* **8**(2):165-196.
- ENGEL, P. *et al.* 2016. The bee microbiome: impact on bee health and model for evolution and ecology of host-microbe interactions. *mBio.* **7**(2).

- EVANS, J. D. and SCHWARZ, R. S. 2011. Bees brought to their knees: microbes affecting honey bee health. *Trends Microbiol.* **19**:614-620.
- GARRITY, G. M., BOONE, D. R. and CASTENHOLZ, R.W. 2001. *Bergey's Manual of Systematic Bacteriology*. Springer-Verlag, New York, NY. **2**(1)
- GILLIAM, M. 1978. Bacteria Belonging to the Genus *Bacillus* Isolated from Honey Bees, *Apis mellifera*, Fed 2, 4-D and Antibiotics. *Apidologie.* **9**(3), 213-222.
- GILLIAM, M., PREST, D. B. and LORENZ, B. J. 1989. Microbiology of pollen and bee bread: taxonomy and enzymology of molds. *Apidologie.* **20**: 53-68.
- GILLIAM, M. and PREST, D. B. 1987. Microbiology of feces of the larval honey bee, *Apis mellifera* . *J. Invert. Pathol.* **49**(1):70-75.
- HACQUARD, S. et al. 2015. Microbiota and host nutrition across plant and animal kingdoms. *Cell Host Microbe.* **17**, 603-616.
- HUTKINS, R. and GOH, Y. J. 2014. *Streptococcus thermophilus*. Encyclopedia of Food Microbiology (2nd Ed.). 554-559.
- ISHII, K. and HAMAMOTO, K. H. 2014. Sekimizu Establishment of a bacterial infection model using the European honeybee, *Apis mellifera* L. *PloS One.* **9**(2).
- ISOLAURI, E., RAUTAVA, S., KALLIOMÄKI, M., et al. 2006. Role of probiotics in food hypersensitivity. *Curr. Opin. Immunol. Clin. Allergol.* **2**:263-271.
- KAZNOWSKI, A., SZYMAS, B., JAZDZINSKA, E., KAZIMIERCZAK, M., PAETZ, H. and MOKRACKA, J. 2005. The effects of probiotic supplementation on the content of intestinal microflora and chemical composition of worker honey bees (*Apis mellifera*). *J. Apic. Res.* **44**:10-14.
- LJUNGH A. and WADSTRÖM, T. 2006. Lactic acid bacteria as probiotics. *Intest. Microbiol.* **7**(2):73-89.
- MOHR, K. I. and TEBBE, C. C. 2006. Diversity and phylotype consistency of bacteria in the guts of three bee species (Apoidea) at an oilseed rape field. *Environ. Microbiol.* **8**(2):258-272.
- PATTERSON, M. J. and BARON S. 1996. In: Medical Microbiology. 4th edition. Galveston (TX): University of Texas Medical Branch at Galveston. Chapter 13.
- PETER, B. O., OLUFEMI, E. A. and IYABO, O. O. 2007. Honey: a reservoir for microorganisms and an inhibitory agent for microbes. *Afr. Health Sci.* **7**(3): 159-165.
- PELCZAR, M. J., CHAN, E. C. S. and KRIEG, N. R. 1993. Microbiology: Concept and application International edition. McGraw-Hill, USA.
- POORMONTASERI, M., HOSSEINZADEH, S. and SHEKARFOROUSH, S. S. 2014. Characterization of *Clostridium botulinum* spores and its toxin in honey. *Iran. J. Vet. Res.* **15**:36-39.
- POTTS, S. G., BIESMEIJER, J. C., KREMEN, C., NEUMANN, P., SCHWEIGER, O., and KUNIN, W. E. 2010. Global pollinator declines: trends, impacts and drivers. *Trends Ecol. Evol.* **25**, 345-353.
- RADA, V., MACHOVA, M., HUK, J., MAROUNE, K M. and DUSKOVA, D. 1997. Microflora in the honeybee digestive tract: counts, characteristics and sensitivity to veterinary drugs. *Apidologie.* **28**: 357-365.

- RAMIREZ, D. and GIRON, M. 2022. Enterobacter Infections. In: StatPearls .Treasure Island (FL): StatPearls Publishing.
- SANDERS, M. E. 2000. Considerations for Use of Probiotic Bacteria to Modulate Human Health. *J. Nutr.* **130**(2): 384–390.
- TAJABADI, N., MARDAN, M., SAARI, N., MUSTAFA, S., BAHREINI, R., MANAP, M.Y. A., SHUHAIMI, M., MEIMANDIPOUR, A. and NATEGHI, L. 2011. Detection and identification of *Lactobacillus* bacteria found in the honey stomach of the giant honeybee *Apis dorsata*. *Apidologie*, Springer Verlag, **42** (5): 642-649.
- ZHENG, H., STEELE, M. I., LEONARD, S. P., MOTTA, E. V. S. and MORAN, N. A. 2018. Honey bees as models for gut microbiota research. *Lab. Anim. (NY)*. **47**: 317-325.

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