MOLECULAR IDENTIFICATION AND GENETIC DIVERSITY OF TEN PIERID BUTTERFLIES BASED ON MITOCHONDRIAL COI GENE

Muhammad Sohel Abedin, Ananna Ghosh¹, Abdul Jabber Howlader and Md. Monwar Hossain*

DNA Barcoding Laboratory, Department of Zoology, Jahangirnagar University, Dhaka, Bangladesh

ABSTRACT: The IUCN Bangladesh has documented a total of 31 species belonging to the Pieridae family in Bangladesh. There is currently limited knowledge regarding the genetic diversity and molecular relationships among these butterflies. The present investigation involved the generation of mitochondrial COI (mtCOI) gene sequences for ten species of butterflies belonging to the Pierid family. After that, BLAST analysis showed 96% to 100% similarity, and then the sequences were submitted to NCBI's GenBank. MEGA10 and BioEdit were used to identify a 584 bp COI gene fragment with 179 variable sites and 128 parsimony-informative sites. The COI sequences' average base compositions were 38.56% T, 17.12% C, 30.18% A, and 14.12% G. While a significant AT bias (68.74%) existed among the Pierid species. The genetic distance between ten Pierid species was assessed using the Kimura 2-Parameter (K2P) algorithm, and the nucleotide differences ranged from 0.00172% to 0.24261%. Afterwards, the Maximum Likelihood (ML) method was used to construct a phylogenetic tree using ten sequences of Pierid species. These species belongs to two subfamilies, Pierinae and Coliadinae in Bangladesh. On the tree, the Pierinae subfamily of Pieridae formed a paraphyletic arrangement whereas the Coliadinae subfamily was shown to be monophyletic. The data analyses support the following relationships between the two subfamilies as follows: [(Appias + Leptosia) + (Pieris + Appias) + (Delias) + (Eurema + Catopsilia) + (Pareronia)]. This supported the theory that Pierinae and Coliadinae are sister taxa. Nonetheless, Pareronia hippia remains outside the main Pierinae group, requiring further study to resolve this issue. Finally, these studies generated ten mtCOI gene sequences that have the potential to serve as valuable references for the accurate identification of Pierid species. In addition, this result could be used in the future to reveal the subfamily relationships within the Pieridae taxonomic classification.

Key words: COI, Genetic diversity, Pieridae, Bangladesh

INTRODUCTION

Butterflies are essential pollinators, and they also cause some harm to crops and forests. Pieridae is one of the most significant and common

^{*}Author for correspondence: <monwar@juniv.edu>, ¹Insect Biotechnology Division, IFRB, Atomic Energy Research Establishment, Dhaka, Bangladesh

^{©2023} Zoological Society of Bangladesh DOI: https://doi.org/10.3329/bjz.v51i3.72027

Lepidoptera family, which serves as pollinators and does a little damage to cultivated crops (Lee 2023). This family is widespread across the globe and comprises approximately 1,200 species (Ackery et al. 1999, Vane-Wright 2003, Sobti et al. 2007). Pierid adult butterflies are typically medium-sized, white, orange, or yellow in coloration, and their pupae have a highly distinctive morphology (Chapman 1895, Talbot 1939, Mosher 1969). There are numerous studies on the morphological, ecological, and molecular characteristics of Pierid species from the world (Sobti et al. 2007). However, the systematic relationship of the Pieridae is inadequately understood, including their molecular data on taxa to a certain extent (Braby et al. 2006). Though the family is well-known to be monophyletic (Wahlberg et al. 2005, Heikkila et al. 2012), but It has only lately been thoroughly researched how the main lineages of the family's phylogeny relate to one another (Braby et al. 2006). Prior classifications were based solely on morphological characteristics and non-cladistic methods, and only a small number of researchers have investigated Pieridae's upper-level systematics, which includes figuring out how different subgroups are related to each other, creating a systematic framework (Klots 1933, Ehrlich 1958), and finally, doing the in-depth phylogenetic analyses that are still needed (Braby et al. 2006).

The family Pieridae is divided into four subfamilies as Pierinae, Coliadinae, Pseudopontiinae, and Dismorphiinae. According to the status of the IUCN in Bangladesh, there are 31 species in the Pieridae family, which is under two subfamilies: Pierinae and Coliadinae. Among them, 23 species belongs to Pierinae and other 8 species to Coliadinae (IUCN 2015). Indeed, one of the least recognized families of butterflies is the Pieridae, and this family is still significant in terms of its biodiversity studies, population survey and significant contributors to evolutionary research (Stavenga et al. 2004, Kemp et al. 2005), and include some species that are significant in agriculture and visual science, respectively (Arikawa 2017). The morphological changes, various color forms, and other elements are a tactic to improve fitness and the survival rate of these insects, but they can pose a significant challenge in accurate identification (Goonesekeraa et al. 2019). In addition, phenotypic variability brought on by aging, sexual dimorphism, seasonal fluctuations, and convergent evolution further confuses the species (Collins and Morris 1985, Kunte et al. 2011, Wilson et al. 2013). To solve these issues, a brief, standardized 650 bp sequence of the mitochondrial DNA (mtDNA), cytochrome c oxidase subunit I (COI) has been suggested as a barcoding technique, or at the very least to confirm species delimitation and evolutionary investigations (Hebert et al. 2003a, Schindel and Miller 2005, Ebach and Holdrege 2005, Costa and Carvalho 2007, Miller 2007). Therefore, the current investigation was conducted to determine the mtCOI gene

sequences of Pieridae butterflies found in Bangladesh. Additionally, the study aimed to reveal genetic divergence and determine the phylogenetic status of this butterfly group.

MATERIAL AND METHODS

Sample collections: Specimens of butterflies were collected from various regions of Bangladesh over the period of 2017 to 2021, as indicated in Table 1. The geographical coordinates of the various places, along with pertinent information such as the collector's name, the date of collection, and additional relevant details, were duly recorded. The specimens were collected from their natural habitat by an insect net and subsequently conserved through desiccation within a small enclosure. Butterflies were morphologically identified using the keys established by Bingham (1905, 1907), Wynter-Blyth (1957), and Talbot (1939, 1947). The voucher samples were generated in accordance with the methodology outlined by Brower (1996).

DNA isolation, amplification and sequencing: The genomic DNA of ten adult Pieridae butterflies was extracted from their legs in accordance with the instructions provided in the Wizard Genomic@ DNA Purification Kit (Promega, USA). The mitochondrial cytochrome c oxidase I (COI) gene region was amplified in PCR using the primers LCO 1490 (5'-GGTCAACAAATCATAAAGATATTGG-3') and HCO 2198 (5'-TAAACTTCAGGGTGACCAAAAAATCA-3'). Using the Q2 Green PCR Master Mix, PCR was performed in a thermal cycler from Applied Biosystems (Veriti, USA). The total volume of the PCR reaction mixture was 20 µl, including 10 µl of 2x master mix, 1 µl of forward primer (10 pM), 1 µl of reverse primer (10 pM), 1 µl of template DNA (depending on the quantity of extracted DNA), and 1 µl of nuclease-free water. The cycle's settings were initial denaturation at 95°C for 5 min, 35 cycles of primer annealing at 49°C for 30 sec, primer extension at 72°C for 45 sec, and a final extension at 72°C for 5 min. 1% agarose gel electrophoresis was used to test the effectiveness of the amplification under ultraviolet light (Bio Doc Analyze, Biometra, Anlytic Jena, Germany). An ABI 3500 sequencer was used to sequence the amplified product.

Phylogenetic analysis: The assembled sequences were aligned using the ClustalW programme for multiple alignment in BioEdit version 7.0 (Hall 1999). Using the MEGA10 software, calculations and summaries of nucleotide compositions and estimates of pairwise distances were done. While the Maximum Likelihood (ML) method was used to construct a phylogenetic tree with 1000 bootstrap replicates in MEGA10 (Saitou and Nei 1987, Kumar *et al.* 2018). In the analysis of each species of COI gene sequences, conspecific sequences from two different geographic regions were used (Table 2). The *Aedes aegypti* originating from Bangladesh was employed as an outgroup in the phylogenetic analysis.

S1 No	Species Name	Country	GenBank Accession No.			
1	Pareronia hippia	India	KX467796			
2	Pareronia hippia	China	NC071367			
3	Appias olferna	Malaysia	KF226283			
4	Appias olferna	Thailand	MW796558			
5	Appias libythea	India	KJ423038			
6	Appias libythea	Thailand	ON437315			
7	Appias lyncida	Malaysia	KF226280			
8	Appias lyncida	China	EF584853			
9	Pieris canidia	India	KT175605			
10	Pieris canidia	Vietnam	OK342235			
11	Delias eucharis	India	ON358410			
12	Delias eucharis	Australia	JX978938			
13	Delias pasithoe	Malaysia	KF226396			
14	Delias pasithoe	China	KU921271			
15	Eurema hecabe	India	KP216719			
16	Eurema hecabe	Korea	GU372559			
17	Catopsilia pomona	India	EU792482			
18	Catopsilia pomona	Pakistan	OR140768			
19	Leptosia nina	India	KJ423032			
20	Leptosia nina	Malaysia	KF226514			
21	Aedes aegypti	Bangladesh	OP942188			

Table 2. List of GenBank-retrieved species used in the analysis

RESULTS AND DISCUSSION

The current investigation involved the generated of mitochondrial COI gene sequences (mtCOI) spanning around 636 bp from ten species belonging to the Pieridae family. The sequenced gene was compared with the available sequences of the GenBank and the result of BLAST analysis showed 96 to 100% similarity to the same species from different geographical areas. Then, ten COI gene sequences were submitted to the NCBI's GenBank in order to get accession numbers (Table 1).

Nucleotide analysis of COI gene: BioEdit and MEGA10 were used to evaluate the COI gene sequences, and the results revealed a 584 bp COI gene fragment with 128 parsimony-informative sites and 179 variable sites (Table 3). The majority of changes were seen at the second and third positions of the codons. In Table 3, it is seen that out of the informative sites, 36 were ranked in

S1	Species Name	GPS	Date of	Voucher	GenBank
No	-	Coordinates	Collection	No	Accession No.
1	Pareronia	23°52'32.9"N	18.07.17	BBV259	MH269418
	hippia	90°16'05.9"E			
2	Appias olferna	24°19'35.3"N	09.07.18	BBV308	MK282887
		91°47'00.8"E			
3	Appias libythea	23°52'33.6"N	17.09.18	BBV271	OQ359494
		90°16'05.8"E			
4	Appias lyncida	24°19'40.1"N	31.10.17	BBV0255	MK317932
		91°47'01.0"E			
5	Pieris canidia	24°19'40.1"N	11.05.18	BBV250	MK282888
		91°47'01.0"E			
6	Delias eucharis	23°52'26.1"N	30.12.21	BBV269	OQ254745
		90°16'03.3"E			
7.	Delias pasithoe	24°19'30.7"N	31.10.17	BBV0270	MK317930
		91°47'14.1"E			
8.	Eurema hecabe	23°52'32.9"N	29.05.17	BBV264	MF784358
		90°16'05.9"E			
9	Catopsilia	23°52'32.9"N	11.05.17	BBV258	MF784359
	pomona	90°16'05.9"E			
10	Leptosia nina	23°52'33.0"N	30.12.21	BBV268	OQ254746
		90°16'05.7"E			

Table 1. List of the species from which COI genes were sequenced and their GenBank accession numbers

the first position, 42 were ranked in the second position, and 46 were ranked in the third position. The average base compositions of the COI sequences were found to be 38.56% thymine (T), 17.12% cytosine (C), 30.18% adenine (A), and 14.12% guanine (G). A significant bias towards the adenine-thymine (AT) base pair was observed, accounting for 68.74% of the total base pairs. The adenine-thymine (A+T) content of the first, second, and third codon positions of the cytochrome c oxidase subunit I (COI) fragment was found to be 59.02%, 57.28%, and 90.04%, respectively (Table 3).

The parsimony informative is employed in molecular phylogenetic and evolutionary studies to elucidate diverse relationships among organisms. Similarly to other protein-coding genes, a significant proportion of changes were detected at the third codon position, as demonstrated by our present findings (Table 3) (Win *et al.* 2015). Additionally, an AT bias was also identified in other Lepidopterans, such as *Pieris* to *Catopsilia*, which aligns with our present results (Table 3) (DeSalle *et al.* 1987, Li *et al.* 2015, Nie *et al.* 2018).

Analysis of genetic distance: With the help of the MEGA10 program, the pairwise distance was determined. Ten sequences of Pierid butterflies had genetic distances ranging from 0.00172 to 0.24261 (Table 4). The lowest genetic distance (0.00172) was found between *Appias libythea* and *Appias olferna*, and their sequence similarity was 99.83%. While the highest genetic distance (0.24261) was found between *Leptosia nina* and *Delias pasithoe* (Table 4). In the present study, the narrow range of interspecific divergence (0.24261%) may be

Position	No. of	No. of variable	No. informative	Statist				
	sites			Т	С	Α	G	AT %
All								
Positions	584	179	128	38.56	17.12	30.18	14.12	68.74
First								
Position	195	58	36	26.15	15.89	32.87	25.07	59.02
Second								
Position	195	59	42	42.66	27.23	14.62	15.48	57.28
Third								
Position	194	62	46	46.90	8.19	43.14	1.75	90.04

Table 3. Basic statistics for COI gene sequences in 10 Pierid species.

a result of the prevalence of low interspecies hybridization, which is a prevalent occurrence among many butterfly species (Win *et al.*, 2015). While higher levels of interspecific divergence may result from hybridization in areas where it is more common (Comeault and Matute 2018). Nonetheless, sequence divergences between 0% and 1.2% are observed in numerous Papilio species, whereas sequence divergences exceeding 2% are employed for species differentiation in lepidopteran insects (Hebert *et al.* 2003b, Zakharov 2004).

Table. 4. Percentage pairwise distances among ten Pieridae species

Species Name	1	2	3	4	5	6	7	8	9
Pareronia hippia									
Appias olferna	0.15380								
Appias libythea	0.15641	0.00172							
Appias lyncida	0.16452	0.13067	0.13313						
Pieris canidia	0.15561	0.14861	0.15115	0.12044					
Delias eucharis	0.18463	0.19190	0.18911	0.17984	0.18920				
Delias pasithoe	0.20787	0.19245	0.18965	0.15547	0.17391	0.14607			
Eurema hecabe	0.19047	0.17850	0.17575	0.17049	0.16593	0.18511	0.20976		
Catopsilia pomona	0.17411	0.14853	0.14600	0.13990	0.14779	0.21087	0.17152	0.18762	
Leptosia nina	0.22170	0.18630	0.18357	0.17537	0.19268	0.23245	0.24261	0.22148	0.23272

Phylogenetic analysis: Maximum Likelihood (ML) method was used to construct the phylogenetic tree. In the analysis, the phylogenetic tree of the ten species shared a common ancestor. On the tree, the Pierinae subfamily of Pieridae formed a paraphyletic arrangement whereas the Coliadinae was shown to be monophyletic (Fig. 1). In the tree analysis, *Appias libythea* and *Appias olferna* showed very close relationships between them as indicated by their substantial boot strap support (Fig. 1). In nature, this two species can be difficult to differentiate in appearance, and it is probable to their similar genetic affinities as shown in the present results (Fig. 1). Nevertheless, *Appias libythea*, *Appias olferna*, and *Leptosia nina* were shown in a cluster. On the other hand, *Pieris canidia* and *Appias lyncida* demonstrated close connections, as did

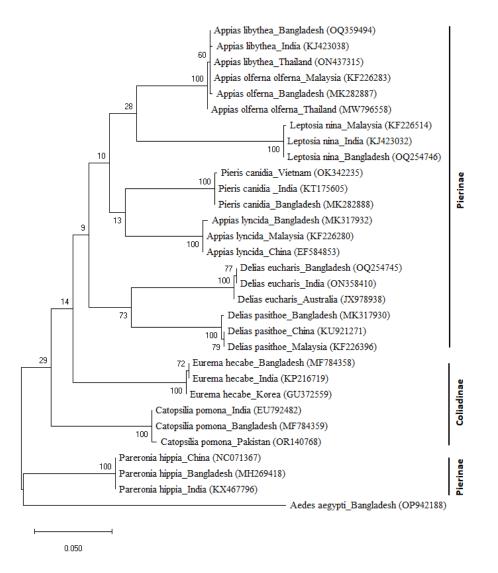


Fig.1: Phylogenetic tree constructed with COI gene sequences using the maximum likelihood (ML) method. Bootstrap values are shown at the branching points. The bar indicates phylogenetic distance.

Delias eucharis and Delias pasithoe. While Eurema hecabe and Catopsilia pomona under the subfamily Coliadinae, were constituted an additional cluster in the analysis. In the tree, only Pareronia hippia remarkably showed distance among other Pierinae species. According to the data analysis, the two subfamilies are related as follows: [(Appias + Leptosia) + (Pieris + Appias) + (Delias) + (Eurema + Catopsilia) + (Pareronia)] (Fig. 1). In the present study, it is shown that Pierinae and Coliadinae are sister taxa, which is also consistent with

the result of Ding and Zhang (2016). Nonetheless, *Pareronia* remain outside the main Pierinae group, and this subfamily played an important role in many evolutionary studies that require further study to resolve this issue in the future (Kemp *et al.* 2005, Braby *et al.* 2006).

CONCLUSION

The current investigation involved the generation and submission of mtCOI gene sequences for ten species of Pierid butterflies to the GenBank database. These generated ten mtCOI genes with the potential to serve as valuable references for the precise identification of Pierid butterflies. In the present study, COI gene sequences of 584bp contained 179 variable sites and 128 parsimony-informative sites. The genetic distance between ten species of Pierid butterfly ranged from 0.00172% to 0.24261%. In Bangladesh, there are two subfamilies of Pieridae, Pierinae and Coliadinae. According to the phylogenetic tree, the Coliadinae subfamily of Pieridae is monophyletic, whereas the Pierinae subfamily is paraphyletic. In the present investigation, the theory supported that Pierinae and Coliadinae are sister taxa. However, *Pareronia hippia* remains outside the primary group of Pierinae, necessitating additional research to resolve this issue.

Acknowledgements: The authors are thankful to the University Grants Commission of Bangladesh (CP No. 3424) for funding the project "Enhancement of Entomological Research Capability Using DNA Barcoding." Thanks are also extended to Wazed Miah Science Research Center, Jahangirnagar University, for providing the laboratory space.

LITERATURE CITED

- ACKERY, P.R., DE-JONG, R. and VANE-WRIGHT, R.I. 1999. The butterflies: Hedyloidea, Hesperoidea and Papilionoidea. In: Kristensen NP (ed.) Lepidoptera: Moths and Butterflies, 1. Evolution Systematics and Biogeography. Handbook of Zoology. Vol. IV. Part 35. Walter de Gruyter, Berlin. 263–300 pp.
- ARIKAWA, K. 2017. The eyes and vision of butterflies. Symposium Review. J. Physiol. **595** (16):5457–5464.
- BINGHAM, C.T. 1905. Fauna of British India. Butterflies, Vol. I. Taylor and Francis, London. 1-511.
- BINGHAM, C.T.1907. Fauna of British India. Butterflies, Vol. II. Taylor and Francis, London. 1-480.
- BRABY, M.F. and TRUEMAN, J.W.H. 2006. Evolution of larval host plant associations and adaptive radiation in Pierid butterflies. *J. Evol. Biol.* **19**: 1677–1690.

- BROWER, A.V.Z. 1996. A new mimetic species of *Heliconius* (Lepidoptera:Nymphalidae), from southeastern Colombia, as revealed by cladistic analysis of mitochondrial DNA sequences. *Zool. J. Linn. Soc.* **116**: 317-332.
- CHAPMAN, T.A. 1895. Notes on butterfly pupae, with some remarks on the phylogenesis of the Rhopalocera. *Entomol.'s Rec. J. Var.* **6**: 101–107, 125–131, 147–152.
- COLLINS, N.M. and MORRIS, M. G. 1985. *Threatened Swallowtail Butterflies of the World:* The IUCN Red Data Book. Gland, Switzerland. 1-401 pp.
- COMEAULT, A. A and MATUTE, D. R. 2018. Genetic divergence and the number of hybridizing species affect the path to homoploid hybrid speciation. *Proc. Natl. Acad. Sci. U S A.* **115**(39): 9761–9766.
- COSTA, F.O. and CARVALHO, G.R. 2007. The Barcode of Life Initiative: synopsis and prospective societal impacts of DNA barcoding of Fish. *Genomics Soc. Pol.* **3**:29-40.
- DESALLE, R. T. FREEDMAN, PRAGER, E.M. and WILSON, A.C. 1987. Tempo and mode of sequence evolution in mitochondrial DNA of Hawaiian Drosophila. *J. Mol. Evol.* **26**:157-164.
- DING, C. and ZHANG, Y. 2016. Phylogenetic relationships of Pieridae (Lepidoptera: Papilionoidea) in China based on seven gene fragments. *Entomol. Sci.* **20**(1): 15–23.
- EBACH, M.C. and HOLDREGE, C. 2005. DNA barcoding is no substitute for taxonomy. *Nature*. **434**: 697.
- EHRLICH, P.R. 1958. The comparative morphology, phylogeny and higher classification of the butterflies (Lepidoptera: Papilionoidea). *Univ. Kans. Sci. Bull.* **39**: 305–364.
- GOONESEKERAA, K., LEEB, P.L.M., POORTENC, G.V. and RANAWAKA, G.R. 2019. The phylogenetic history of the old world butterfly subtribe Mycalesina extended: the *Mycalesis* (Lepidoptera: Nymphalidae) of SriLanka. *J. Asia Pac. Entomol.* **22**(1): 121-133.
- HALL, T.A. 1999. Bioedit, a user-friendly biological sequences alignment editor and analysis program for windows95/98/NT. *Nucleic Acids Symp.* Ser. **41**: 95-98.
- HEBERT, P.D.N., CYWINSKA, A., BALL, S.L. and deWAARD, J.R. 2003a. Biological identifications through DNA barcodes . *Proc. Royal Soc. B.* **270** (1512): 313–321.
- HEBERT, P.D.N., RATNASINGHAM, S. and deWAARD, J.R. 2003b. Barcoding animal life: cytochrome c oxidase subunit 1 divergences among closely related species. *Proc. R. Soc. B.* 270 (Suppl): 96-99.
- HEIKKILA, M., KAILA, L., MUTANEN, M., PENA, C. and WAHLBERG, N. 2012. Cretaceous origin and repeated tertiary diversification of the redefined butterflies. *Proc. Royal Soc. B: Biol. Sci.* 279: 1093–1099.
- IUCN Bangladesh. 2015. Red List of Bangladesh Volume 7: Butterflies. IUCN, International Union for Conservation of Nature, Bangladesh Country Office, Dhaka, Bangladesh. 1-400 pp. https://www.iucn.org/resources/research-publication/redlist-bangladesh-volume-7butterflies
- KEMP, D.J., RUTOWSKI, R.L. and MENDOZA, M. 2005. Colour pattern evolution in butterflies: a phylogenetic analysis of structural ultraviolet and melanic markings in North American sulphurs. *Evol. Ecol.Res.* 7: 133–141.

- KLOTS, A.B. 1933. A generic classification of the Pieridae (Lepidoptera) together with a study of the male genitalia. *Entomol. Am.* 12: 139–242.
- KUMAR, S., STECHER, G., LI, M., KNYAZ, C. and TAMURA, K. 2018. MEGA X: Molecular Evolutionary Genetics Analysis across computing platforms. *Mol. Biol. Evol.* **35**:1547-1549.
- KUNTE, K., SHEA, C., AARDEMA, M.L., SCRIBER, J.M., JUENGER, T.E., GILBERT, L.E. and KRONFORST, M.R. 2011. Sex Chromosome Mosaicism and Hybrid Speciation among Tiger Swallowtail Butterflies. *PLoS Genet.* 7(9): e1002274.
- LEE, M. 2023. Environmental factors affecting honey bees (*Apis cerana*) and cabbage white butterflies (*Pieris rapae*) at urban farmlands. *Peer J.* 11:e15725.
- LI, H., SHAO, R.F., SONG, N., SONG, F., JIANG, P., LI, Z.H. and CAI, W.Z. 2015. Higher-level phylogeny of paraneopteran insects inferred from mitochondrial genome sequences. *Sci. Rep.* **5** (8527).
- MILLER, S.E. 2007. DNA barcoding and the renaissance of taxonomy. PNAS. 104 (12):4775-4776.
- MOSHER, E. 1969. Lepidoptera pupae. Five collected works on the pupae of North American Lepidoptera. East Lansing: Entomological Reprint Specialists. 323 pp.
- NIE, L., WANG, Y., HUANG, D., TAO, R., SU, C., HAO, J. and ZHU, C. 2018. Mitochondrial genomes of four Pierid butterfly species (Lepidoptera:Pieridae) with assessments about Pieridae phylogeny upon multiple mitogenomic datasets. *Syst. Zool.* **43**(4): 387–409.
- SAITOU, N. and NEI, M. 1987. The neighbor-joining method: A new method for reconstructing phylogenetic trees. *Mol. Biol. Evol.* **4**: 406-425.
- SCHINDEL, D.E. and MILLER, S.E. 2005. DNA barcoding a useful tool for taxonomists. Nature. **5**: 435(7038):17.
- SOBTI, R.C., SHARMA, V.L., KUMARI, M., GILL, T.K., SINGH, J., SODHI, M., MUKESH, M., BANSAL, S., ARYA, S. and BISNOI, S. 2007. Genetic relatedness of six North-Indian butterfly species (Lepidoptera:Pieridae) based on 16S rRNA sequence analysis. *Mol. Cell. Biochem.* 295: 145– 151.
- STAVENGA, D.G., STOWE, S., SIEBKE, K., ZEIL, J. and ARIKAWA, K. 2004. Butterfly wing colours: scale beads make white pierid wings brighter. *Proc. Royal Soc. B: Biol. Sci.* 271: 1577–1584.
- TALBOT, G. 1939. The Fauna of British India including Ceylon and Burma. Butterflies. Vol. I. Taylor
 & Francis, London (Reprinted by Today and Tomorrow's Printers and Publishers, New Delhi).
 xxix+600.
- TALBOT, G. 1947. The Fauna of British India including Ceylon and Burma. Butterflies. Vol. II. Taylor & Francis, London (Reprinted by Today and Tomorrow's Printers and Publishers, New Delhi). xv+506.
- VANE-WRIGHT, R.I. 2003. Evidence and identity in butterfly systematics. In: Boggs CL, Watt WB, Ehrlich PR (eds) *Butterflies: Ecology and Evolution Taking Flight*, University of Chicago Press, Chicago, IL.477–513 pp.
- WAHLBERG, N., BRABY, M.F., BROWER, A.V.Z., DE JONG, R., LEE, M.M., NYLIN, S., PIERCE, N., SPERLING, F.A., VILA, R., WARREN, A.D. and ZAKHAROV, E. 2005. Synergistic effects of

combining morphological and molecular data in resolving the phylogeny of butterflies and skippers. *Proc. Royal Soc. B: Biol. Sci.* **272**: 1577–1586.

- WILSON, J.J., SING, K. W. and SOFIAN-AZIRUN, M. 2013. Building a DNA barcode reference library for the true butterflies (Lepidoptera) of Peninsula Malaysia: what about the subspecies? *PLoS ONE*. 8(11): e79969.
- WIN, N.Z., CHOI, E.Y., JANG, D-J., PARK, J. and PARK, J.K. 2015. Molecular comparison of the genus Junonia (Lepidoptera:Nymphalidae) in Myanmar. J. Asia-Pac. Biodivers. 8 (4): 287-294.
- WYNTER-BLYTH, M.A. 1957. *Butterflies of India region*. Bombay Natural History Society, Bombay. 523 pp.
- ZAKHAROV, E.V., CATERINO, M.S. and SPERLING, F.A.H. 2004. Molecular phylogeny, historical biogeography, and divergence time estimates for swallowtail butterflies of the genus Papilio (Lepidoptera:Papilionidae). Syst. Biol. 53: 193-215.

(Manuscript received on 01 September; 2023 revised on 11 November 2023)