Immature reticulocyte fraction as a predictor of bone marrow recovery in children with acute lymphoblastic leukaemia on remission induction phase

Yesmin S¹, Sultana T², Roy CK², Rahman MQ², Ahmed ANN²

¹Department of Pathology, Dhaka Community Medical College, Dhaka, ²Department of Clinical Pathology, Bangabandhu Sheikh Mujib Medical University (BSMMU), Dhaka

Abstract

Immature reticulocyte fraction a new routine parameter in the hematology analyzer can give the idea of the earliest morphologic change of bone marrow recovery before other test become positive after chemotherapy. A prospective observational study was carried out in the Department of Clinical Pathology in collaboration with Paediatric Haematology and Oncology, Bangabandhu Sheikh Mujib Medical University during a period of one year starting from October 2009 to September 2010 to evaluate the bone marrow recovery in children with acute lymphoblastic leukaemia by automated reticulocyte analysis. Total fifty patients were enrolled in this study on remission induction phase. All patients were between 8 months to 15 years age range with a mean age of 5.5 ± 3.2 . At the end of the study out of 50 cases, 52% patients showed early immature reticulocyte fraction recovery and concluded that the immature reticulocyte fraction parameter showed earlier haematopoietic recovery than the current practice of absolute neutrophil count recovery.

Introduction

Acute lymphoblastic leukaemia (ALL) is a malignancy of lymphoid tissue¹. ALL occurs most frequently in the children². After diagnosis chemotherapy is the main stay of treatment in ALL. Chemotherapy uses anti-cancer (cytotoxic) drugs to destroy the leukaemic cells. After chemotherapy blood counts generally fall within a week of treatment and may take some time to recover^{3,10,19}. At this period extensive monitoring of bone marrow recovery are needed. Among the hematological parameters immature retculocyte fraction can predict bone marrow recovery over others. So, serial measurement of immature reticulocyte fraction (IRF) is useful to monitor bone marrow regenerative function. Reticulocytes are immature red blood cells. They are released in the peripheral blood after a period of maturation in the bone marrow and undergo further differentiation into mature RBC⁶. Flow cytometric reticulocyte analysis is more precise and more sensitive than manual reticulocyte counting⁷. Besides this, the measured fluorescence intensity allows the quantification of reticulocyte maturity²⁰. In automated flowcytometry method, reticulocyts have been classified morphologically into three maturational stages: low fluroscent reticulocytes (LFR), middle fluroscent reticulocytes (MFR) and high fluroscent reticulocytes (HFR)³. Immature Reticulocyte Fraction (IRF) is defined as the ratio of immature reticulocytes to the total number of

reticulocytes. They are larger, having the greatest light scatter properties due to the highest level of ribonucleic acid (RNA). Immature reticulocytes normally constitute less than five percent (5%) of the total number of reticulocytes⁵. It is released into the peripheral blood during periods of intense erythropoietic stimulation. An increase in the reticulocyte percent >1% used as an indicator of erythroid regeneration^{3,8}. Spanish Multicentric Study Group for Haematopoietic recovery defined IRF >5% as recovery⁸. Absolute neutrophil count (ANC) is defined as the number of mature neutrophils plus bands per unit of volume generally accepted as a primary indicator of successful bone marrow recovery. An increase in ANC $\geq 0.5 \text{ x} 10^9/\text{L}$ defines successful myeloid recovery after chemotherapy^{4,9,10}. The aim of this study is to establish the earliest indicator of marrow recovery among the reticulocyte subpopulations in children with ALL.

Materials and Methods

This study was carried out in the Department of Clinical Pathology and Department of Paediatric Haematology and Oncology, Bangabandhu Sheikh Mujib Medical University (BSMMU), Dhaka during the period of October' 2009 to September' 2010. 50 Children upto 18 years of age irrespective of sex with acute lymphoblastic leukaemia attended in Paediatric Hemato-Oncology outpatients and inpatients department for treatment included in this study. Blood sample (2 ml) was collected in an EDTA tube for complete blood count (CBC), absolute neutrophil count (ANC), reticulocyte profile (both manual and automated method) and peripheral blood film (PBF) examination. Count was done preferrably within 2 hrs of collection.

Supravital staining of unfixed RBCs was done with new methylene blue (NMB) in 50 patients and 40 controls. Mixing of 100 μ l whole blood with 100 μ l of 1% NMB solution was done in a 10x75- mm tube. After incubation at 37⁰ for 15-20 minutes, the dilution was remixed and a wedge smear was performed¹¹. The number of reticulocytes per 1000 RBCs determined microscopically on x100 objective. Then percentage of reticulocyte was done. A reticulocyte was defined as a RBC containing at least 2 granules of reticulum (Figure-1). Absolute reticulocyte count was calculated from RBC count obtained from automated hematology analyzer.

Automated reticulocyte count was done in 50 patients in Sysmex xt-2000i haematology analyzer. The measuring principle of the system is based on flow cytometry combined with hydrodynamic focusing. EDTA blood (100 µl) is aspirated and intracellular RNA is stained by auramine O, which is fluroscent under argon laser light¹². The reticulocyte population is further subdivided into LFR, MFR and HFR (Figure-2). The percentage of reticulocyte is given as the sum of LFR, MFR and HFR. The IRF was manually calculated by MFR plus HFR. IRF>5% were taken as bone marrow recovery³. ANC was calculated from total leucocyte count and differential count of WBC. Serial hemogram with reticulocyte count, IRF and ANC measurements was done in every 4 days interval upto 32 days of the therapy.

All necessary and relevant data were processed. Data were evaluated by standard statistical methods. Analysis was done by Statistical package for social science (SPSS) 16 by applying appropriate formula. Bone marrow recovery were evaluated by Mean, Median, Paired t test, Coefficient of variance test, ANOVA test.

Results

During study randomly selected 40 healthy subjects were taken to establish the reference values for reticulocyte profile in the laboratory. Fifty children with ALL were 8 months to 15 years of age range and mean age of the patients was 5.5 ± 3.2 years. Maximum patients were male. Male and female ratio was 1.5:1. In this study, during induction remission phase hemoglobin (Hb) level were

gradually decreased upto 12th day (p<0.05). However from 16th day onwards the Hb level remain the similar with baseline status which was not significant (p>0.05) (Paired t-test). At diagnosis mean total count of WBC was 14.74±25.47 (x10⁹/L). During induction remission total count of WBC was sharply declined at day 8 and continued up to last follow up. Statistically significant differences were observed between different follow-ups day (p<0.05) (Paired t-test).

At diagnosis mean total count of RBC was 4.0 ± 0.9 (x 10^{12} /L). During induction remission total count of RBC was significantly declined upto 20^{th} day (p<0.05) (Paired t-test). In this study ret% significantly declined upto day 4 and then gradually increased or remain the same with the diagnosis day in both automated and manual method (Paired t-test). The absolute reticulocyte concentration significantly declined upto day 4 then it remain the same upto last follow up.

This study showed IRF declined gradually in 24 cases and reaches to low-level mean of 12.1±7.1 % with a median of 6 days (days 4-8). But IRF showed high range at diagnosis before the chemotherapy (Paired t-test). Recovery day was ranged from 16.6±4.6 (median day 16) (Table-I) (ANOVA test). In this study the children were profoundly neutropenic (<500/ml) in day 12 and day 16. The ANC were remain the same with the day of diagnosis from day 20 to 32 day of the therapy. The ANC were significantly (p<0.05) declined upto day 16 of therapy (Paired t-test). The range of ANC recovery was 10-32 days (median 23 day) (Table-I) (ANOVA test). This study found that 52 % cases recovery of IRF occurred earlier than ANC and the median difference of IRF preceding ANC was days 7. Late recovery occurred in 22% cases. It was on the same day in 26% cases. Thus the similar or earlier recovery was in 78% cases.

Table I: Comparison of IRF and ANC recovery in days (n=50)

Parameter	Recovery in days		P Value
IRF			
Mean+SD	16.6+4.6		0.016*
Median	16		
Range (Min-max)	(10-30)		
ANC			
Mean+SD	23.3+5.7		0.001*
Median	23		
Range (Min-max)	(10-32)		
* Significant			
5			
Table II: IRF recovery versus ANC recovery			
Parameter	Earlier IRF	Same	Later IRF
	recovery	IRF	recovery
	•	recovery	2
No. of cases (50 cases)	26 (52%)	13 (26%)	11 (22%)
Median difference	7	-	8
(Day post CT)	(10-30 days)		(14-32 days



Fig. 1: Photomicrograph of reticulocytes in children with ALL on day 8 induction of remission phase. Stained supravitally with new methylene blue (x1000)



Fig-2: IRF scattergram in Sysmex XT-2000i hematology analyzer

Discussion

Several studies showed that the younger reticulocytes detected by the flow cytometric method are better indicators of recovery than the detection of ANC in post chemotherapy patients^{5,13,17}. Grotto et al. in 1999 used immature reticulocyte fraction (IRF) as a indicator of early recovery. This study also evaluated IRF as a predictor of bone marrow recovery in post chemotherapy children with ALL and it was compared with other hematological indicators of bone marrow recovery.

This study showed that IRF was the first sign of haematological recovery in 52% of the patients studied preceding the rise in ANC. The IRF-signaled recovery appeared on a median of 7 days when compared to the ANC. Das et al. (2006) showed earlier IRF recovery at a median 21 days. This study found ANC recovery at a median of 23 days but Das et al. (2006) found ANC recovery between 10-35 days (median day 19). In this study IRF showed 78% similar or early recovery. A study showed, 91.2% IRF recovery occurred before ANC in chemotherapy patients on remission induction

phase⁸. Lesesve et al. (1995) found earlier recovery of IRF in 88.4% cases than ANC by a period of 33.3 days. IRF recovery was 72.1% similar or earlier than ANC¹³. In other studies, IRF were also used as a marker of recovery and showed similar results^{3,16}.

In this study IRF parameter showed earlier haematopoietic recovery than the current practice of ANC recovery for monitoring in children with acute lymphoblastic leukemia after chemotherapy. This early laboratory indicator will guide the clinicians to make important therapeutic decisions, which will be economic and live saving for the patients. This study concluded that reticulocyte counting with IRF can be routinely and widely used in the laboratory to evaluate the bone marrow erythropoietic activity after chemotherapy.

References

- 1. Satake N, Yoon JM. Acute Lymphoblastic Leukaemia. E medicine Pediatric 2009; 12:1-26.
- Baruchel A, Leblanc T, Schaison G. Pathology of acute lymphoblastic leukaemias, In: Lilleyman, Hann I, and Blanchette V (eds), Pediatric Haematology 2nd ed, London, Harcourt Brace and Company 1999; 519-536.
- Kuse R. The appearance of reticulocytes with medium or high RNA content is a sensitive indicator of beginning granulocyte recovery after aplasiogenic cytostatic drug therapy in patients with AML. Ann Hematol 1993; 66: 213-214.
- Greinix HT, Linkesch W, Keil F et al. Early detection of hematopoietic engraftment after bone marrow and peripheral blood stem cell transplantation by highly fluorescent reticulocyte counts. Bone Marrow Transplant 1994; 14: 307-313.
- Davis BH, Bigelow NC. Flow cytometric reticulocyte quantification using thiazole orange provides clinically useful reticulocyte maturity index. Arch Pathol Lab Med 1989; 113: 684-689.
- Riley S, Jonathon M, Ezra B, Goel R, Tidwell A. Reticulocytes and Reticulocyte Enumeration. Journal of Clinical Laboratory Analysis 2001;15:267-294.
- 7. Hoewen B. Reticulocyte maturation. Blood cells 1992; 18:167-186.
- Remacha. Flow cytometric reticulocyte quantification in the evaluation of hematologic recovery. Spanish Multicentric Study Group for Hematopoietic Recovery. Eur J Haematol 1994;53: 293-297.
- Hellgriel K. The clinical significance of reticulocyte determination, In: Basting T, 5th eds. The Emerging Importance of Accurate Reticulocyte Counting. Miles, Inc. Diagnostic Division, Tarrytown 1993: 7-10.
- Davis BH. Immature reticulocyte fraction (IFR): by any name, a useful clinical parameter of erythropoietic activity. Lab Hematol 1996; 2:2-8.

- Bain BJ, Lewis SM, Bates I. Basic haematological techniques, In: Lewis SM, Bain BJ, Bates I, Dacie and Lewis Practical Haematology, 10th eds. Philadelphia: Churchill living stone 2008; 25-57.
- Imazu M. General Description of the Automated Hematology Analyzer, XT-2000i', Sysmex. J Int 2002;12: 13-17.
- Das R, Dip NB, Garewal G, Marwaha RK, Vohra H. Automated Reticulocyte response is a good predictor of bone-marrow recovery in pediatric malignancies. Pediatric Hematology and Oncology 2006; 3:299-305.
- Bhatnagor S, Chandra J, Naayanb S. Hematological Changes and Predictors of Bone Marrow Recovery in Patients with Neutropenic Episodes in Acute Lymphoblastic Leukemia. Journal of Tropical Pediatrics 2002; 48(4): 200-203.
- Dekoninck A, Brusselmans C, Goossens W. Indicators for hematopoietic recovery in patients after bone marrow transplantation or intensive chemotherapy, Department of laboratory Medicine, University Hospital Leuven, Leuven, Belgium 2002; pp.39.

- Kuse R, Foures C, Jou JM, d'Onofrio G, Paterakis G. Automated reticulocyte counting for monitoring patients on chemotherapy for acute leukaemias and malignant lymphomas. Clin Lab Haematol 1996; 18(1): 39-43.
- Norhana J.F, De Souza CA, Vigorito AC et al. Immature reticulocytes as an early predicator of engraftment in autologous and allogenic bone marrow transplantation. Clin Lab Haemtol 2003;25:47-54.
- Lesesve JF, Lacombe F, Marit G et al. High fluorescence reticulocytes are an indicator of bone marrow recovery after chemotherapy. Eur J Haematol 1995;54: 61-63.
- Thomas ED, Marmont AM, Sale GE. Aplastic anemia and hematopoiesis after bone marrow transplantation, In: Zucker-Franklin D, Greaves MF, Grossi CE, Marmont AM, editors. Atlas of blood cells. 2nd edition. Philadelphia: Lea & Febiger 1988; 733–760.
- Rowan R.M, Cavill I, Corberand J.X. The reticulocyte count: progress towards the resurrection of a useful clinical test. Clinical and Laboratory Haematology 1996;18(1): 3-8.