

Chemical, Biological and Pharmacological Activities of *Ficus religiosa*: an Extensive Review

Sifat Ara Sultana¹, Shusmita Talukder¹, Nusrat Jahan Shawon², Fahima Aktar³, Jakir Ahmed Chowdhury⁴, Abu Asad Chowdhury³, Shaila Kabir³ and Md. Shah Amran³

¹Department of Pharmacy, Faculty of Pharmacy, University of Dhaka, Dhaka-1000, Bangladesh

²Department of Clinical Pharmacy and Pharmacology, Faculty of Pharmacy, University of Dhaka Dhaka-1000, Bangladesh

³Molecular Pharmacology and Herbal Drug Research Laboratory, Department of Pharmaceutical Chemistry, Faculty of Pharmacy, University of Dhaka, Dhaka-1000, Bangladesh

⁴Department of Pharmaceutical Technology, Faculty of Pharmacy, University of Dhaka Dhaka-1000, Bangladesh

(Received: August 29, 2023; Accepted: April 10, 2024; Published (web): July 30, 2024)

Abstract

Herbal drugs are one of the world's oldest and most precious gifts. Medicinal herbs and plants have a prominent place in the healthcare system, with *Ficus religiosa*, being one of the most important. It is commonly known as fig, which is widely used in traditional medicinal systems as antiulcer, anticonvulsant, wound healing, antinephropathic, antiviral, analgesic, anti-inflammatory, antimicrobial, immunostimulant, antihyperlipidemic, antioxidant, antiasthmatic, parasympathetic modulatory, anti-acetylcholinesterase, anticancer and cognitive enhancing agents. Secondary metabolites isolated from the plant such as stigmasterol, methyl oleanolate, lanosterol, caffeic acid, bergenin, amides, lupin 3-one, β -sitosterol, n-octacosanol, and flavonoids are considered to be the principal bioactive compounds of *F. religiosa*. The purpose of this review was to outline the phytochemistry, pharmacology and toxicity profile of *F. religiosa*. Aside from plant morphology and phytochemistry, the application of the plant in the most recent discoveries and nanotechnology was also explored. Furthermore, the review addresses recent therapeutic breakthroughs and identifies areas for further research that should be addressed in future studies.

Key words: *Ficus religiosa*, phytochemistry, traditional medicine, pharmacology, toxicity, nanotechnology.

Introduction

In traditional and modern medicinal systems, many of the beneficial therapeutic agents are derived from plant sources (Pochhi and Muddeshwar, 2017). These plants, used as herbal medicines, hold great promise for the discovery of novel drug molecules for diseases for which current treatments are inefficient while exhibiting fewer adverse effects (Kukde *et al.*, 2021). The genus *Ficus* Linn. consisting of 750-1000 species, is the largest tracheophyte medicinal plants genus in the Moraceae

family and is found in the tropical and subtropical regions (Hesami *et al.*, 2018a; Biju *et al.*, 2020). *F. religiosa*, commonly known as Peepal is a perennial, evergreen monoecious tree widely distributed in Bangladesh, India, Pakistan, Sri Lanka, Nepal, Myanmar, China, Iraq, and Iran and an important member of the genus having medicinal and religious values (Hesami and Daneshvar, 2018b; Chaudhary *et al.*, 2017). *F. religiosa* has been planted as an ornamental tree in various regions of South America, North America, Australia, Europe,

Corresponding author: Md. Shah Amran; Email: amranms@du.ac.bd

DOI: <https://doi.org/10.3329/bpj.v27i2.75192>

the Middle East and Africa (Vianna Filho *et al.*, 2017). The term '*Ficus*' is Latin for fig, while the term '*religiosa*' indicates the religious significance of the tree. In the Indian subcontinent, it is known as the mythical 'World Tree' or the 'Tree of Life' which is the earliest depicted tree in literature and art (Gupta *et al.*, 2021). It is also called the 'Sacred Fig' as it is regarded sacred by the believers of Hinduism, Buddhism and Jainism (Hesami *et al.*, 2017; Patil *et al.*, 2020). It's a tall, highly branched tree with heart-shaped and leathery leaves with long pointed edges on long thin axils and purple figs that grow in pairs (Garg *et al.*, 2018). *F. religiosa* can propagate through seeding and vegetative methods and the seeds remain viable for a few months (Daneshvar, 2019; Parab *et al.*, 2021).

F. religiosa is a good source of proteins, carbohydrates, vitamins, minerals, dietary fibers, amino acids and nitrogen (Tahir *et al.*, 2020). The bark contains saponin gluanol acetate, β -sitosterol, tannins, lupeol acetate, lupeol, ceryl behenate, α -amyrin acetate, leucoanthocyanin and leucoanthocyanidin (Selvan and Chourasia, 2017). Among the plants of the *Ficus* genus, the fig of this plant contains the highest level of serotonin (Mamidiseti *et al.*, 2018). The air pollution tolerance index for *F. religiosa* is 10.73-9.39, indicating that the plant can be used as a pollution sink (Bashir *et al.*, 2021; Sastry and Barua, 2017; Agrahari *et al.*, 2018). It also has a rougher surface area, a denser canopy and a larger surface area than most other tropical trees, which contribute to its superior dust-capturing ability (0.82 mg/cm² and 0.88 mg/cm² in industrial and commercial locations) (Roy *et al.*, 2020). The tree is used worldwide as food, medicine, fodder, vegetable & fuel and the wood has been used in the construction of houses, furnishings, packaging, spoons, forks, bowls and other things (Thanomchat *et al.*, 2019; Subash *et al.*, 2019). Its medicinal value has been documented in the management of infectious diseases, microbiological disorders, inflammation, epilepsy, menstrual disorders, gastrointestinal difficulties, respiratory problems, arthritis, seizure, and stress (Aslam and Akhtar, 2019; Mamta *et al.*, 2021; Rajiv and Sivaraj, 2012).

F. religiosa is a general ingredient in many formulations used in traditional medical systems such as Ayurveda, Siddha and Unani (Chavan *et al.*, 2019). The fig tree is widely used in traditional medicinal systems as an antidiabetic, antiulcer, anticonvulsant, wound healing, antineuropathic, antiviral, analgesic, anti-inflammatory, antimicrobial, immunostimulant, antihyperlipidemic, antioxidant, antiasthmatic, parasympathetic modulatory, anti-acetylcholinesterase, anticancer agent and cognitive enhancer (Utami *et al.*, 2020; Prakash *et al.*, 2017; Manorenjitha *et al.*, 2013; Rathod *et al.*, 2018). In various Ayurvedic formulations namely, Nyagrodhadi churna, Nalpamaradi tailam, Saribadyasavam and Chandanasavam the bark of the tree is used (Raisagar *et al.*, 2019). '*F. religiosa* arishta' is an herbal hydro-alcoholic formulation from Ayurvedic pharmacopeia used in the treatment of inflammatory disorders (Jyothisree and Umadevi, 2020). The plant also shows anti-nephrotoxic and nephroprotective properties against a variety of nephrotoxic substances (Fang *et al.*, 2021; Chaure *et al.*, 2021). There have also been reports on the gender-specific adverse effects of oral administration of *F. religiosa* extracts (Akhtar *et al.*, 2020; Goyal, 2014). Even though several literature reviews on this plant have already been published, this one is presented to include all of the most recent scientific updates on its phytoconstituents and therapeutic effects studied through various approaches.

Materials and Methods

An extensive literature search was conducted on the phytochemical and pharmacological effects on *F. religiosa*. Research articles on phytochemical analysis, *in vivo*, *in vitro* and clinical trials were collected. All these papers were collected using bibliographic databases, search engines and journal websites, such as PubMed, Google Scholar, Nature, Researchgate and others.

Results and Discussion

Plant morphology: The matured ficus tree is thirty meters in height with dark barks. The

undeveloped leaves appear pink or red at first, then turn to deep green, and cordate in shape as mature. The heart-shaped and glossy leaves are 12-20 cm in length, 5-8 cm wide and have five to seven veins. They are joined to the stem with a 6 to 8 cm long petiole. The leaves have five to eight sets of transverse veins, as well as a system of extremely fine veins. They are oval in shape with a leathery, shiny surface, short, sheathing stipules, caudate-

acuminate apex and pinnate venation. There are no trichomes in the plant (Sethuramani *et al.*, 2021). The bark is slightly arched or straight, with a thickness ranging from 1.0 to 3.0 cm (5 to 8 mm). It occurs in level or slightly bent sections ranging in thickness from 1.0 to 2.5 cm or more. In February, the little crimson flowers appear and the fruits appear from May to June as figs that are 12-13 mm or 1.5 inches wide (Patil *et al.*, 2020).



Figure 1. *Ficus religiosa* plant with unripe fruits.

Phytochemistry of F. religiosa: The alkaloids, phenolics, flavonoids, phytosterols, furanocoumarins, amino acids, volatile compounds and aliphatic and aromatic alcohols were extracted and identified from different extracts of *F. religiosa* (Hesami and Daneshvar, 2018c; Shaikh *et al.*, 2020). Several

secondary metabolites, such as stigmaterol, methyl oleanolate, lanosterol, caffeic acid, bergenin, amides, lupin 3-one, β -sitosterol, n-octacosanol, and flavonoids, have been characterized, while many are still unidentified (Hesami *et al.*, 2018d; Nawaz *et al.*, 2020). Lupenyl acetate, lanosterol, piperlongumine,

piperine, quercetin, kaempferol, myricetin, and methyl piperate are the active ingredients having anti-inflammatory properties (Yueniwati *et al.*, 2021; Hesami *et al.*, 2019).

The presence of flavonoids, alkaloids, carbohydrates, saponins, glycosides, and tannins in various extracts of *F. religiosa* leaf was discovered in qualitative phytochemical investigations (Ghadigaonkar *et al.*, 2021; Utami *et al.*, 2021). Significant amounts of total phenolic contents (19.97-62.93 g gallic acid equivalents/mg) and total flavonoid contents (29.88-135.95 g QE/mg) were found in leaf extract (Ghadigaonkar *et al.*, 2021). The leaves contain 0.4-0.75 mg/g ascorbic acid that contribute to its high tolerance towards air pollution (Bashir *et al.*, 2021). Among the forty-four compounds identified in the leaf oil, eugenol (27%), 2-Phenylethyl alcohol (8%), 3-Methylcyclopentane-1,2-dione (10.8%) and itanoic anhydride (15.4%) are the four main constituents of the leaf extract. The other components include *n*-hexanol, phenol, adipoin, *n*-nonanal, 2-phenylethyl alcohol, benzeneacetonitrile, (*E*)-linalool oxide, catechol, coumaran, (*E*)-cinnamyl alcohol, *p*-vinylguaiacol, eugenol, dihydroactinidiolide, α -copaene-11-ol, β -eudesmol, α -eudesmol, α -cadinol, pentadecanal, palmitic acid, and phytol (Al-Snafi, 2017). Thirty-one volatile compounds have been identified in the dry leaves including palmitic acid (hexadecanoic acid), propionic acid, stearic acid (octadecanoic acid), citric acid, aspartic acid, and phenylalanine (Suriyakalaa *et al.*, 2021). Religiosin B and C enzyme glycoproteins, also known as serine proteases are found in the decolorized latex of *F. religiosa* (Al-Snafi, 2017; Murugesu *et al.*, 2021). Furanocoumarin derivatives (bergapten and begaptol), flavonoids, phenols, phytosterols, alkaloids, tannins, vitamin K, lanosterol, lupen-3-one, methyl oleanolate, and steroids have been found in the aqueous extract of bark (Devanesan *et al.*, 2018). On the other hand, the petroleum ether extract contained 35 compounds including 13 saturated fatty

acids (31.19%), 8 unsaturated fatty acids (59.24%), 1.76% alkene, 6.02% alkane, 0.97% fatty alcohols, and 0.82% cyclic fatty acid esters (Ali *et al.*, 2017a).

Recently, a new β -steryl naphthyl ester and two novel fatty acids have been characterized in a phytochemical screening of the methanolic extract of the stem bark for the first time (Ali *et al.*, 2017b). Other recent phytochemical studies led to the discovery of a novel lanostane type-triterpenic ester and a novel naphthyl-substituted phytosterol from the bark and the extraction of a new fiber with 55.58% cellulose content from the root of *F. religiosa* (Ali *et al.*, 2020; Moshi *et al.*, 2020a and Moshi *et al.*, 2020b). The fruit of *F. religiosa* contains substantial amounts of flavonoid (1111.1 ± 33.3 mg/kg dry matter) and phenolic contents (Jawanjal *et al.*, 2021; Sultana and Anwar, 2008). The seeds contain amino acids, phytosterols, fatty matter, glycoside, carbohydrates, coloring matter, and β -sitosterol (Devanesan *et al.*, 2018). Endophytes, including actinomycetes, bacteria, and fungi live inside the plant tissue and create secondary metabolites that are similar to those produced by plants (Tiwari *et al.*, 2017). Figure 2 depicts the distinct phytochemicals discovered in *F. religiosa*.

Pharmacological activities: *F. religiosa* is a magical plant with various medicinal activities and uses. It contains different secondary metabolites in different parts, such as leaves, roots, bark and stems. These complex chemical substances play role in different pharmacological activities. Different systems like the endocrine system, central & peripheral nervous systems, reproductive system and digestive system are affected by the chemical compounds of *F. religiosa*. A wide spectra of *in vivo* and *in vitro* studies is conducted to ensure pharmacological activities, like, nephroprotective effect, bronchoconstriction effect, astringent effect, anti-hyperlipidemic effect, antimicrobial effect and so on. These pharmacological effects with mechanism, assay and proper dose are described below.

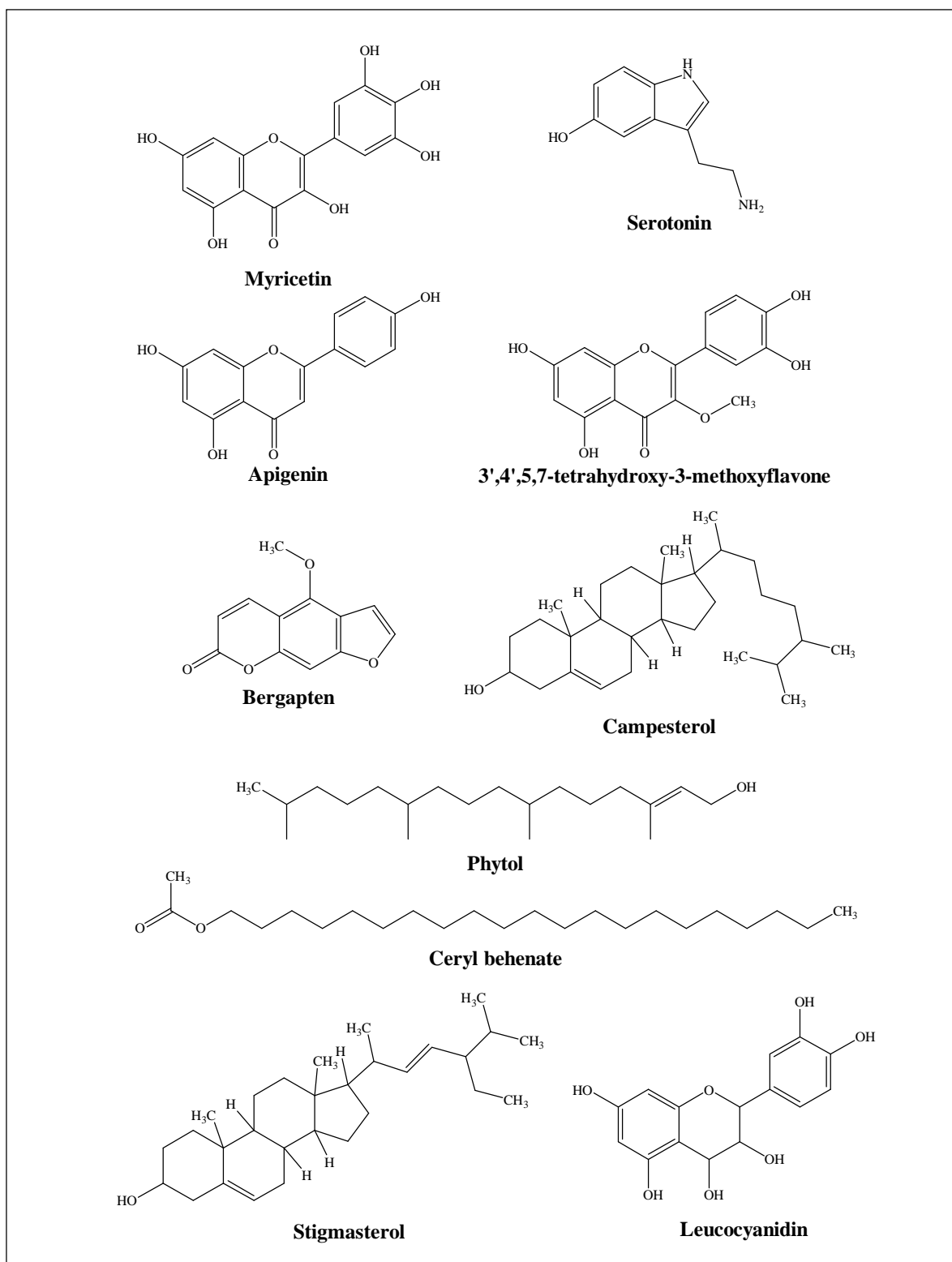
Figure 2. Chemical structures of bioactive compounds of *F. religiosa*.

Table 1. Microorganisms found in *F. religiosa* and their therapeutic activities.

Microorganism	Species	Compounds isolated	Therapeutic activity	References
Bacteria	<i>Azoarcus nasutitermitis</i> <i>Azoarcus rhizosphaerae</i>			Lin et al., 2020
Endophytic fungi	<i>Curvularia lunata</i> <i>Aspergillus</i> sp. <i>Aspergillus aculeatus</i> , <i>Penicillium</i> sp., <i>Cephalophora irregularis</i> , <i>A. sydowii</i> , <i>Diaporthe</i> sp., <i>A. quadrilineatus</i> , <i>A. flavus</i> , <i>A. versicolor</i>	1-Eicosane Naphthaquinone antibiotic herbarin, herbaridine A	Antimicrobial, antidiabetic and cytotoxic properties Antidiabetic, antiinflammatory effect Anti-oxidant and anti- diabetic activities	Murugesu et al., 2021 Jayant and Vijayakumar, 2021
Nematode	<i>Parasitodiplogaster religiosae</i>			Zeng et al., 2018
Rhizobacterium	<i>Bacillus safensis</i>			Asif, 2019

(i) *Antioxidant activity*: Tannins, flavonoids and polyphenols are mainly responsible for antioxidant activities. Some assays justify this statement that aqueous extract at a dose of 100 and 200 mg/kg shows antioxidant effect in streptozotocin-induced diabetic rats by decreasing superoxide dismutase [for 100 mg/kg] and by enhancing catalase (CAT) and glutathione peroxidase (GSH-Px) activity (Kirana et al., 2009). In the case of methanolic extract 11.75, 50.65, 400 µg shows activity by bleaching 1,1-diphenyl-2-picrylhydrazyl (DPPH) radical (Anandjiwala et al., 2008). Ethanolic extract at 200 µg/mL to 1000 µg/mL shows antioxidant effect by increasing H₂O₂ in the DPPH method (Zaidi et al., 2009).

(ii) *Wound healing activity*: Flavonoids, terpenoids, sterols, alkaloids and saponin β-sitosterol-D-glucoside are responsible for wound healing activity. In Wistar albino rats, 300 mg/kg dose of leave extract increases wound healing/closure rate, whereas 10% ointment (in case of roots) decreases epithelialization period and increases hydroxyproline content (Zaidi et al., 2009).

(iii) *Nephroprotective effects*: Nephroprotective effects refer to the protection of the nephron or kidney from harm. Alkaloids, glycosides, amino acids, flavonoids and tannins show nephroprotective activities. Methanolic extracts of latex 200 mg/kg dose show this effect in Wistar rat models by decreasing levels of urea and creatinine (Yadav and Srivastava, 2013).

(iv) *Anti-asthmatic activity*: Asthma is a condition that narrows airways and creates difficulty in breathing. Flavonoids, terpenoids, tannins, phenols and sterol help in the maintenance of asthma. Aqueous extract of leaves at a dose of 150 and 300 mg/kg of body weight in guinea pigs delay histamine-induced pre-convulsion dyspnea (Kapoor et al., 2011).

(v) *Anti-acetylcholinesterase activity*: Steroids, alkaloids, flavonoids, phenols, saponins, tannins, glycosides and sterols are responsible for this activity. Methanolic extract of stem bark at a dose of 73.69 µg/mL prolongs the half-life of acetylcholine by hindering acetylcholinesterase (Vinutha et al., 2007).

(vi) *Broncho-constriction effect*: Histamin and acetylcholine has significant effects in broncho-constriction. Glycosides, terpenoids, flavonoids, serotonergic content are key components that shows broncho-constriction effect. Methanolic extract of fruits at various doses such as 0.5, 1 and 2 mg/kg are effective, this assay is conducted in histamine and acetylcholine induced guinea pig and these doses affect the EC(50) doses of both histamine and acetylcholine (Ahuja et al., 2011).

(vii) *Astringent activity*: Components present in *F. religiosa* steroids, flavonoids, alkaloids, phenol content, glycosides, tannins, saponins, polyphenolic compounds, sterols, terpenoids, serotonergic content are responsible for astringent activity. Using tannic acid as standard, entire mature fruit and inner bark shows 0.6% astringency compared to tannic acid (fruit) in bovine serum albumin assay with remazol brilliant blue (Ahuja et al., 2011).

(viii) *Anti-fertility activity*: Flavonoids, serotonergic content, terpenoid, glycosides show anti-fertility activity. Methanolic extract of fruits of *F. religiosa* gives anti-fertility activity, for example- 1% anti-fertility effect was seen on uterus of goats when methanolic extraction is given. It reduces diameter of uterine glands and myometrium thickness of uterus of goats and increased activity (Sharma et al., 2013).

(ix) *Anti-inflammatory activity*: Anti-inflammatory activities are shown by steroids, flavonoids, alkaloids, phenol content, glycosides, tannins, saponins, polyphenolic compound, sterols. Ethanolic extract of bark at a dose of 100 mg/kg when administrated in carrageenan induced golden Syrian hamsters expresses anti-inflammatory action. Mainly production of reactive oxygen species is responsible for anti-inflammatory activity (Murugesan et al., 2012).

(x) *Anti-ulcer activity*: Flavonoids, terpenoids, tannins, phenols, sterols, Steroids, alkaloids, phenol content, glycosides and saponins show anti-ulcer activity. Ethanolic bark extract at 200 and 400 mg/kg doses in reduces the amount of gastric acid and hyper acidity in Wistar rats. On the other hand, ethanolic leaf extract at a dose of 2000 mg/kg in albino mice

decreases gastric secretion (Gregory et al., 2013; Khan et al., 2011).

(xi) *Hepatoprotective activity*: Alkaloids, glycosides, amino acids, flavonoids and tannins show hepatoprotective activity. Methanolic extracts of latex at a dose of 300 mg/kg in cisplatin-induced liver injury in Wistar rats degenerate inflammatory infiltrate (Yadav, 2015).

(xii) *Analgesic effect*: Flavonoids, phenols, terpenoids and glycosides are mainly responsible for analgesic activity. Methanolic extract of fruits in acetic acid writhing test (in mice) causes inhibition of cyclo-oxygenases and shows an analgesic effect at 200 and 400 mg/kg doses (Mamidiseti et al., 2018).

(xiii) *Anti-epileptic activity*: Bark extract of ethanol in Swiss albino mice shows an epileptic effect at 200 and 400 mg/kg doses. This is because high saponin content block voltage-dependent Na⁺ channels and NMDA receptors and thus delay the onset of convulsion and reduce the duration of convulsion (Sapkota et al., 2021).

(xiv) *Antiarthritic activity*: Flavonoids and terpenoids are mainly responsible for the antiarthritic activity. Methanolic bark extract in the case of Freund's adjuvant-induced arthritis in rats causes suppression of inflammatory mediator (Garg et al., 2018).

(xv) *Antihyperlipidemic effect*: Aqueous extract of leaves at a dose of 250 mg/kg in alloxan-induced diabetic rats shows anti-hyperlipidemic activity by improving serum lipids, lipoproteins concentration and lipid metabolizing enzymes activity such as - lipase, HMG CoA reductase, LCAT (Lecithin cholesterol acyl transferase) (Pochhi and Muddeshwar, 2017).

(xvi) *Anti-cancer activity*: Some assays justify the anti-cancer activities of *F. religiosa*. Chloroform extract of leaves at a dose of 110 µg/mL in trypan blue exclusion dye test shows anti-cancer activity. Besides, chloroform extract at various doses such as 62.5, 125, 250, and 500 µg/mL in MDA-MB-231 breast cancer cell line through SRB assay shows oxidative stress and causes retardation of cell growth (Kumaresan et al., 2018). Polyphenol dichloroacetic

acid present in *F. religiosa* shows cytotoxic effect. Combined crude extract of *F. religiosa* and *F. benghalensis* at a dose of 100 µg/mL in mitochondrial reduction assay against HeLa (human cervical cancer cell line) shows a cytotoxic effect. Through depolarizing polyphenol, dichloroacetic acid affects the proliferation of tumor cells and causes apoptosis in glioblastoma cancer (Kumaresan et al., 2018).

(xvii) *Antidiabetic effect*: Sterols, β-sitosterol-D-glucoside, alkaloids, flavonoids and tannins present in *F. religiosa* are responsible for the antidiabetic effect. Aqueous extract of root, bark, leaves and ethanolic extract of bark in normal fasted male albino rabbits show positive results. Phytosterols present in bark decreases blood glucose level. Besides the antioxidant effect of tannin, flavonoids and polyphenols are suggested as possible mechanisms for anti-diabetic effects. For leaf extract, it reduces plasma glucose levels in a time and dose-dependent manner both in normal and alloxan-treated diabetic mice (Brahmachari and Augusti, 1962; Pandit et al., 2010; Singh et al., 2011).

(xviii) *Antimicrobial activity*: 70% ethanolic extract of bark shows activity against *H. pylori* in pylori plates and strains from antral biopsies patients (Zaidi et al., 2009). Ethanolic extract of bark at a concentration of 10 g of powdered bark gives antimicrobial activity against *B. cereus* and *S. agalactiae* in agar disc diffusion and disc diffusion method (Nair and Chanda, 2006). Aqueous and ethanolic extract of bark at a concentration of 10 g of powdered bark is active against *B. cereus* in agar disc diffusion (for aqueous extract) and agar well-diffusion (for ethanolic extract) (Nair and Chandra, 2007). Aqueous, methanol and chloroform extract of bark show activity against *E. coli* in disc diffusion method where three enterotoxigenic *E. coli* were isolated from diarrhoeal patients (Uma et al., 2009). 80% ethanolic extract of leaves is active against *E. coli*, *P. aeruginosa*, *S. aureus*, and *B. subtilis* in agar dilution method (Valsaraj et al., 1997).

70% ethanolic extract of leaves at a dose of 100µL of 100 mg/mL extract shows antimicrobial

activity against *S. aureus*, *S. paratyphi*, *S. dysenteriae*, *S. typhimurium*, *E. coli*, *S. typhi* and *C. albicans* in agar well diffusion method (Aqil and Ahmad, 2007). Aqueous, ethanol and methanolic extract of leaves for 0.5 mL of extract from 10g powder of leaf with 100 mL of solvent are active against *B. subtilis*, *P. aeruginosa*, *E. coli*, and *S. typhi* in agar plate method (Preethi et al., 2010). Chloroform extract of fruits at a dose of 0.5 mg (in acetone) with a disc of 6.35 mm diameters is active against *A. chroococcum*, *B. cereus*, *B. megaterium*, *S. faecalis*, *S. lactis* and *K. pneumonia* in filter paper disc diffusion method (Mousa et al., 1994). Phenolic compounds, such as phenols, flavanols, alkaloids show anti-fungal effect. N-butanol fraction of stem bark at a dose of 0.125 and 2 mg/mL shows anti-fungal activity while used in antifungal assay (through agar dilution method adopted by Javaid and Sammad) (Shahid et al., 2016).

F. religiosa in recent scientific discoveries: Researchers have recently become interested in nanotechnology because of its cellular-level mechanisms that can benefit the pharmaceutical, nutraceutical and cosmetic industries. The application of plants for nanoparticle manufacturing has become increasingly popular in recent years. When compared to conventional methods, this process is far more ecologically acceptable and generates biodegradable waste. It has been found that the employment of *F. religiosa* extract in solid lipid nanoparticles increased the bioavailability and potency of the principal bioactive compounds (Priyanka et al., 2017). Solid lipid nanoparticle form of *F. religiosa* shows greater potency and bioavailability and increases the plasma half-life of lupeol present in bark extract in male Wister rats (Priyanka et al., 2017). Therefore, the utilization of solid lipid nanoparticles may result in a reduction in dose frequency, a longer duration of action, and improved therapeutic efficacy. Silver nanoparticle production employing plant extract is a fast, low-cost, environment-friendly, one-step biosynthesis technique that is also safe for medicinal uses (Rahman and Prasanna, 2018). Table 2 lists the nanoparticles produced with *F. religiosa* extracts and their therapeutic efficacy or use in different studies.

Natural sources such as *Ficus* provide a safer and more sustainable way to remove hazardous metal ions from industrial wastes than many other approaches (Tariq *et al.*, 2021; Tariq *et al.*, 2019). Senthilkumar *et al.* used organic *Ficus* Chlorophyll-a to synthesize a semi-organic Chl-a doped ADP (CHLAADP) single crystal with electrical and optical conductivity ideal for lenses, electronics, power grids, and sensors (Senthilkumar *et al.*, 2020; Senthilkumar and Manimekalai, 2020). According to theoretical and experimental investigations, *F.*

religiosa can act as a green corrosion inhibitor due to the presence of campesterol, serotonin and myricetin, which reduces the erosion rate of steel at pH below seven (Haldhar *et al.*, 2018). Thus, it can serve as an excellent mild steel corrosion preventative agent in 0.5M H₂SO₄ owing to the abundance of aromatic rings and heteroatoms in the principal components (Haldhar *et al.*, 2018). The biomass carbon obtained from *F. religiosa* leaves could be a viable choice for potassium ion intercalating electrodes in energy-storing technologies (Rajkumar *et al.*, 2021).

Table 2. Application of *F. religiosa* in nanoparticle synthesis and their therapeutic effects.

Nanoparticles	Solution used	Plant part	Evaluated activity/ application	Result	Reference
Copper oxide nanoparticles	Cupric sulphate	Leaf	Anticancer activity	Apoptotic effect in cancer cells	Sankar <i>et al.</i> (2014)
CuO/C porous nanocomposite	Cupric sulphate	Leaf	Antimicrobial, antioxidant	Strong antimicrobial activity	Bhavyasree and Xavier (2021)
Silver nanoparticle	AgNO ₃	Leaf	Antibacterial efficacy	Potent antibacterial efficacy	Kushare <i>et al.</i> (2020)
	AgNO ₃	Leaf	Larvicidal activity	Reduced survival rate of larvae	Kantrao <i>et al.</i> (2017)
	AgNO ₃	Leaf	Antitumor, antioxidant, antiangiogenic	Treatment of Dalton's ascites lymphoma	Antony <i>et al.</i> (2013)
SnO ₂ nanoparticles	Tin chloride dihydrate	Leaf	Glucose detector	Successful detection of aqueous glucose	Narayana <i>et al.</i> (2020)
Solid lipid nanoparticles	Glyceryl monostearate, poloxamer	Stem bark	Enhance bioavailability	Enhanced C _{max} and prolonged t _{1/2} of lupeol	Priyanka <i>et al.</i> (2017)
		Lipid phase	Andiabetic effects	Decreased blood glucose level	Priyanka <i>et al.</i> (2018)
	Glyceryl monostearate	Stem bark	Andiabetic effects	Reduced blood glucose levels	Karunanidhi <i>et al.</i> (2021)
TiO ₂ nanoparticles	TiO(OH) ₂	Leaf	Larvicidal activity	Increased larvicidal activity	Soni and Dhiman (2020)
Zinc oxide nanoparticles	ZnNO ₃ and NaOH	Leaf	Larvicidal activity	Increased larvicidal activity	
	ZnNO ₃	Leaf	Antibacterial properties	Good antibacterial activity	Raghavendra <i>et al.</i> (2017)

Plant toxicity and biosafety: *F. religiosa* is considered to be safe as there is no evidence of its toxic effect from ancient traditional use. Even now the majority of studies done to determine toxicity show no sign of toxicity. Although the aqueous and alcoholic extract of leaves is considered to be safe,

some extract shows toxicity varying on the dose. A table (Table 3) is given below regarding those extracts and the toxic effect with dose. Chloroform extract of fruits of *F. religiosa* shows toxicity with LC₅₀ of 400 µg/mL, but having LC₅₀ > 100 µg/mL is considered non-toxic. From this point of view, fruit

extract can be considered safe (Mousa *et al.*, 1994; Moshi *et al.*, 2010). Overall studies show that toxicity depends on therapeutic dose and type of extract and

maximum extracts of *F. religiosa* are considered to be safe.

Table 3. Toxicity profile of *F. religiosa* and adverse effects in different animal models.

Plant extract	Toxic dose	Safe dose	Animal model	Adverse effects	Reference
Methanolic fruit extract	above 1000 mg/kg	25, 50 and 100 mg/kg	Rotarod	Neurotoxic effect	Singh and Goel, (2009)
Aqueous bark extract	above 2000 mg/kg	Up to 2000 mg/kg	Swiss female albino mice	Catharsis/allergies	Pandit <i>et al.</i> (2010)
Phytosterolin isolated from the bark	40-400 mg/kg	Not specified	Male albino rats	Increased motor activity and temperature, hyperirritability, piloerection, at last death after clonic convulsions	Balachandran and Govindarajan (2005)
Leaves with tannic acid	2 doses/day for 40 days	Leaves without tannic acid	In goats	Necrosis of the epithelial cells of both cortical and medullary regions of the renal tubule, vacuolar degeneration, myocardium congestion, focal hemorrhage, patchy degenerations with complete absence of epidermis	Tripathy <i>et al.</i> (1984)

Conclusions

In the Indian subcontinent, herbs have long been the primary means of treatment. The presence of diverse complex organic compounds of varying compositions identified as secondary plant metabolites confers therapeutic effects on medicinal plants (Chandrasekar *et al.*, 2010). Due to its therapeutic, aesthetic and religious value, *F. religiosa* L. is one of the most prominent plants in the Moraceae family and is recognized as a multifaceted forest tree (Hesami and Rahmati-Joneidabad, 2018e). It is an extensively used bark drug in traditional medicines and it's one of the five bark drugs in 'Panchavalkala' (Mallya *et al.*, 2018). It has been used in Bangladesh to treat a variety of ailments, including infectious infections, inflammation, and cancer (Bhalerao and Sharma, 2014). When administered in the recommended amount, it has no harmful effects on humans (Lakshmi HimaBindu *et al.*, 2013). The differing proportions of numerous phytochemicals, in selecting of suitable extracting solutions, could be one of the factors for the functions of these plant extracts (Abusufyan *et al.*, 2018). Its wide spectrum of secondary metabolites such as lanosterol, stigmasterol, caffeic acid, β -

sitosterol, bergenin, methyl oleanolate, lupin-3-one, amides and flavonoids could be another factor behind its success as a traditional remedy (Hesami *et al.*, 2020). *F. religiosa* are important elements of tropical ecosystems because their wider canopy areas promote plants dispersed by frugivores and other species (Chakraborty *et al.*, 2021). Therefore, *F. religiosa* stem bark has future prospective to be used in a variety of ways. The antioxidant, anti-inflammatory, anticancer, antidiabetic, antibacterial, activities of the species were validated in the investigations, providing a scientific foundation for its use in herbal medicines. Antifertility, antiproliferative, anticancer, anti-stress, immunomodulatory and antimutagenic properties are a few potential therapeutic effects of *F. religiosa* that require further investigations. *In vivo* studies can be carried out in the future to assess the pharmacological actions demonstrated. Therefore, this review of phytochemicals and their therapeutic actions will provide forthcoming information on current studies, and therapeutic factors that may call for additional emphasis, as well as practical outcomes to be included in this plant species.

Acknowledgement

We would like to thank the authors of the articles we have cited. We also express our gratitude to the authority of the Department of Pharmaceutical Chemistry for using their computer of the Molecular and Herbal Drug research Laboratory established under the HEQEP Project (CP-3258).

Authors contribution

MSA has conceived the original idea. SAS and ST extensively consulted the literature. SAS, ST, and MSA prepared the initial manuscript and arranged the reference section. FA, JAC, AAC and SK critically reviewed the overall activities. MSA supervised the whole activity. All the authors read the review article meticulously and agreed to submit the article.

Funding statement

Part of this work has been funded by a grant from the Ministry of Science and Technology, Government of the Peoples Republic of Bangladesh. Serial: 523; Ref.No.:39.00.0000.009.14.019.21, Date: 15-12-2021.

Conflict of interest

There is no conflict of interest according to the authors.

References

- Abusufyan, S., Ibrahim, M. and Mohib, K. 2018. Comparative *in vitro* antidiabetic and antioxidant activity of various extracts of *Ficus* species. *Pharmacogn. J.* **10**, 349-354.
- Agrahari, P., Richa, R., Swati, K., Rai, S., Singh, V.K. and Singh, D.K. 2018. *Ficus religiosa* Tree leaves as bioindicators of heavy metals in Gorakhpur city, Uttar Pradesh, India. *Pharmacogn. J.* **10**, 416-420.
- Ahuja, D., Bijjem, K.R. V and Kalia, A.N. 2011. Bronchospasm potentiating effect of methanolic extract of *Ficus religiosa* fruits in guinea pigs. *J. Ethnopharmacol.* **133**, 324-328.
- Akhtar, N., Iqbal, S., Shahzad, M.F., Latif, M. and Iqbal, F. 2020. Oral supplementation of *Ficus religiosa* leaf extract adversely affects the selected behavioral aspects of male albino mice. *Biologia* **75**, 2295-2300.
- Al-Snafi, A.E. 2017. Pharmacology of *Ficus religiosa*-a review. *IOSR J. Pharm.* **7**, 49-60.
- Ali, A., Jameel, M. and Ali, M. 2017a. Fatty acids analysis of *Ficus religiosa* stem bark by gas chromatography-mass spectrometry. *Int. J. Adv. Pharm. Med. Bioallied Sci.* **2017**, 112.
- Ali, A., Jameel, M. and Ali, M. 2017b. A new naphthyl substituted β -sitosterol and fatty acids from the bark of *Ficus religiosa* L. *Indian Drugs* **54**, 18-22.
- Ali, A., Jameel, M. and Ali, M. 2020. New Naphthyl substituted phytosterol and Lanostane type-triterpenic esters from the stem bark of *Ficus religiosa* L. *Indian J. Pharm. Educ. Res.* **54**, 750-754.
- Anandjiwala, S., Bagul, M.S., Parabia, M. and Rajani, M. 2008. Evaluation of free radical scavenging activity of an ayurvedic formulation, Panchvalkala. *Indian J. Pharm. Sci.* **70**, 31.
- Antony, J.J., Sithika, M.A.A., Joseph, T.A., Suriyakalaa, U., Sankarganesh, A., Siva, D., Kalaiselvi, S. and Achiraman, S. 2013. *In vivo* antitumor activity of biosynthesized silver nanoparticles using *Ficus religiosa* as a nanofactory in DAL induced mice model. *Colloids Surf. B.* **108**, 185-190.
- Aqil, F. and Ahmad, I. 2007. Antibacterial properties of traditionally used Indian medicinal plants. *Methods Find. Exp. Clin. Pharmacol.* **29**, 79-92.
- Asif, G.M.S.U. 2019. Draft genome sequence of *Bacillus safensis* strain sami, isolated from leaf veins of *Ficus religiosa*. *Microbiol. Resour. Announc.* **8**.
- Aslam, I., and Akhtar, N. 2019. Development of *Ficus religiosa* extract-loaded emulsion system for topical application: characterization and stability evaluation. *Trop. J. Pharm. Res.* **18**, 2003-2010.
- Balachandran, P. and Govindarajan, R. 2005. Cancer - an ayurvedic perspective. *Pharmacol. Res.* **51**, 19-30.
- Bashir, I., Awan, M.U.F., Muhammad, S., Sardar, A.A., Luqman, M. and Arooj, F. 2021. Physiochemical alterations and air pollution tolerance index in *Ficus religiosa* L. planted along polluted roadsides of Lahore city. *Rev. de Chim.* **72**, 112-121.
- Bhalerao, S.A., and Sharma, A.S. 2014. Ethenomedicinal, phytochemical and pharmacological profile of *Ficus religiosa* Roxb. *Int. J. Curr. Microbiol. Appl. Sci.* **3**, 528-538.
- Bhavyasree, P.G., and Xavier, T.S. 2021. A critical green biosynthesis of novel CuO/C porous nanocomposite via the aqueous leaf extract of *Ficus religiosa* and their antimicrobial, antioxidant and adsorption properties. *Chem. Eng. J. Adv.* **8**, 100152.
- Biju, C.R., Jyotisree, G., Amita, S. and Sruthi, R. 2020. A comparative evaluation of *Ficus religiosa* with *Ficus* species for its anti-inflammatory activity: a review. *J. Appl. Pharm. Sci.* **8**, 13-16.

- Brahmachari, H.D. and Augusti, K.T. 1962. Effects of orally effective hypoglycaemic agents from plants on alloxan diabetes. *J. Pharm. Pharmacol.* **14**, 617.
- Chakraborty, A., Mahajan, S., Bisht, M.S. and Sharma, V.K. 2022. Genome sequencing and comparative analysis of *Ficus benghalensis* and *Ficus religiosa* species reveal evolutionary mechanisms of longevity. *iScience.* **25**, 105100.
- Chandrasekar, S.B., Bhanumathy, M., Pawar, A.T. and Somasundaram, T. 2010. Phytopharmacology of *Ficus religiosa*. *Pharmacogn. Rev.* **4**, 195-199.
- Chaudhary, A., Yadav, B.S., Singh, S., Maurya, P.K., Mishra, A., Srivastva, S., Varadwaj, P.K., Singh, N.K. and Mani, A. 2017. Docking-based screening of *Ficus religiosa* phytochemicals as inhibitors of human histamine h2 receptor. *Pharmacogn. Mag.* **13**, 706-714.
- Chaure, A.R., Gangane, G.R., Moregaonkar, S.D., Shete, H.J. and Choudhari, A.N. 2021. Pathological studies on subacute toxicity of *Amaranthus retroflexus* and its amelioration of *Ficus religiosa* in wistar rats. *Pharma innov.* **10**, 177-179.
- Chavan, A., Bedekar, G., Miniyar, P. and Gawande, V. 2019. Phytochemical screening and antimicrobial investigation of *Ficus religiosa* leaves. *Curr. Trends Biotechnol. Pharm.* **1**, 31-42.
- Daneshvar, M.H. 2019. *In vitro* and *in vivo* seed germination of *Ficus religiosa* L. 9th Congress of Iranian Horticulture Science. **2019**.
- Devanesan, E.B., Anand, A.V., Kumar, P.S., Vinayagamoorthy, P. and Basavaraju, P. 2018. Phytochemistry and pharmacology of *Ficus religiosa*. *Syst. Rev. Pharm.* **9**, 45-48.
- Fang, C.Y., Lou, D.Y., Zhou, L.Q., Wang, J.C., Yang, B., He, Q.J., Wang, J.J. and Weng, Q.J. 2021. Natural products: potential treatments for cisplatin-induced nephrotoxicity. *Acta Pharmacol. Sin.* **42**, 1951-1969.
- Garg, K., Sharma, J., Bhargava, A. and Bajwa, P.S. 2018. Antiarthritic activity of different plant extracts of *Ficus religiosa* stem bark in complete Freund's adjuvant induced arthritis in rats. *Asian Pac. J. Health Sci.* **5**, 183-188.
- Ghadigaonkar, S., Reddy, A.G., Kalakumar, B., Lakshman, M. and Rajkumar, U. 2021. Quantification of total phenolic content, total flavonoid content and evaluation of *in vitro* free radical scavenging activities in *Ficus religiosa* Linn. *Pharma. innovation* **10**, 84-88.
- Goyal, A.K. 2014. Phytochemistry and *in vitro* studies on anti-fertility effect of *Ficus religiosa* fruits extract on uterine morphology of goat (*Capra hircus*). *Int. J. Drug Dev. Res.* **6**, 141-158.
- Gregory, M., Divya, B., Mary, R.A., Viji, M.H., Kalaichelvan, V.K. and Palanivel, V. 2013. Anti-ulcer activity of *Ficus religiosa* leaf ethanolic extract. *Asian Pac. J. Trop. Biomed.* **3**, 554-556.
- Gupta, A.K., Gupta, S. and Charu, B. 2021. A critical review on Ashvattha leaves (*Ficus religiosa* Linn.): an ayurvedic perspective and current practice. *Int. J. Ayurveda. pharm. Res.* **9**, 62-68.
- Haldhar, R., Prasad, D., Saxena, A. and Kumar, R. 2018. Experimental and theoretical studies of *Ficus religiosa* as green corrosion inhibitor for mild steel in 0.5 M H₂SO₄ solution. *Sustain. Chem. Pharm.* **9**, 95-105.
- Hesami, M., Daneshvar, M.H., Yoosefzadeh-Najafabadi, M. and Alizadeh, M. 2018. Effect of plant growth regulators on indirect shoot organogenesis of *Ficus religiosa* through seedling derived petiole segments. *J. Genet. Eng. Biotechnol.* **16**, 175-180.
- Hesami, M., Tohidfar, M., Alizadeh, M. and Daneshvar, M.H. 2020. Effects of sodium nitroprusside on callus browning of *Ficus religiosa*: An important medicinal plant. *J. For. Res.* **31**, 789-796.
- Hesami, M. and Daneshvar, M.H. 2018a. *In vitro* adventitious shoot regeneration through direct and indirect organogenesis from seedling-derived hypocotyl segments of *Ficus religiosa* L.: an important medicinal plant. *Hortscience.* **53**, 55-61.
- Hesami, M. and Daneshvar, M.H. 2018b. Indirect organogenesis through seedling-derived leaf segments of *Ficus religiosa* - a multipurpose woody medicinal plant. *J. Crop Sci. Biotechnol.* **21**, 129-136.
- Hesami, M., Daneshvar, M.H. and Lotfi-Jalalabadi, A. 2017. Effect of sodium hypochlorite on control of *in vitro* contamination and seed germination of *Ficus religiosa*. *Iran. J. Plant Physiol.* **7**, 2157-2162.
- Hesami, M., Daneshvar, M. H. and Yoosefzadeh-Najafabadi, M. 2018. Establishment of a protocol for *in vitro* seed germination and callus formation of *Ficus religiosa* L., an important medicinal plant. *Jundishapur J. Nat. Pharm. Prod.* **13**.
- Hesami, M., Daneshvar, M.H. and Yoosefzadeh-Najafabadi, M. 2019. An efficient *in vitro* shoot regeneration through direct organogenesis from seedling-derived petiole and leaf segments and acclimatization of *Ficus religiosa*. *J. For. Res.* **30**, 807-815.
- Hesami, M. and Rahmati-Joneidabad, M. 2018. Morphological characterization of *Ficus religiosa* genotypes in Iran by multivariate analysis. *Hortscience.* **53**, 932-936.

- Javaid, A. and Samad, S. 2012. Screening of allelopathic trees for their antifungal potential against *Alternaria alternata* strains isolated from dying back *Eucalyptus* spp. *Nat. Prod. Res.* **26**, 1697-1702.
- Jawanjal, P., Bedarkar, P., Patgiri, B. and Shukla, V.J. 2021. Phytochemical and high-performance thin-layer chromatography analysis of Ashawattha (*Ficus religiosa* Linn.) Kaanda Twaka (outer portion of stem bark) Churna (powder). *BLDE Univ. J. Health Sci.* **6**, 43.
- Jayant, K.K. and Vijayakumar, B.S. 2021. *In vitro* anti-oxidant and anti-diabetic potential of endophytic fungi associated with *Ficus religiosa*. *Ital. J. Mycol.* **50**, 10-20.
- Jyothisree, G. and Umadevi, Dr.S. 2020. Development and validation of a new HPTLC Finger printing analytical method for newly formulated Arishta from *Ficus religiosa* root bark. *Int. J. Pharm. Sci. Invent.* **9**, 01-07
- Kantrao, S., Ravindra, M.A., Akbar, S.M.D., Jayanthi, P.K. and Venkataraman, A. 2017. Effect of biosynthesized Silver nanoparticles on growth and development of *Helicoverpa armigera* (Lepidoptera: Noctuidae): Interaction with midgut protease. *J. Asia-Pac. Entomol.* **20**, 583-589.
- Kapoor, M., Jasani, N., Acharya, N., Acharya, S. and Kumar, V. 2011. Phytopharmacological evaluation and anti-asthmatic activity of *Ficus religiosa* leaves. *Asian Pac. J. Trop. Med.* **4**, 642-644.
- Karunanidhi, P., Verma, N., Kumar, D.N., Agrawal, A.K. and Singh, S. 2021. Triphenylphosphonium functionalized *Ficus religiosa* L. extract loaded nanoparticles improve the mitochondrial function in oxidative stress induced diabetes. *AAPS Pharm. Sci. Tech.* **22**, 158.
- Khan, M.S.A., Hussain, S.A., Jais, A.M.M., Zakaria, Z.A. and Khan, M. 2011. Anti-ulcer activity of *Ficus religiosa* stem bark ethanolic extract in rats. *J. Med. Plants Res.* **5**, 354-359.
- Kirana, H., Agrawal, S.S. and Srinivasan, B.P. 2009. Aqueous extract of *Ficus religiosa* Linn. reduces oxidative stress in experimentally induced type 2 diabetic rats. *Indian J. Exp. Biol.* **47**, 822-826.
- Kukde, V.S., Bhongade, M.A. and Kamde, S.S. 2021. Standardization and phytochemical screening of *Ficus religiosa*. *Res. J. Pharm. Technol.* **14**, 971-976.
- Kumaresan, S., Ramasamy, R. and Jayachandran, P.R. 2018. Antioxidant and cytotoxic activity of combined extracts prepared using *Ficus religiosa* and *Ficus benghalensis* leaves against cervical cancer cell line (HeLa). *Asian J. Pharm. Clin. Res.* **11**, 407-410.
- Kushare, V.N., Pawar, R.K., Ghotekar, S.V. and Kabra, P.S. 2019. Comparison of efficacy of biogenic silver nanoparticles using *Ficus* species. *Int. J. Pharmacomet. Integr. Biosci.* **4**, 10-13.
- Lin, S.Y., Hameed, A., Tsai, C.F., Huang, G.H., Hsu, Y.H. and Young, C.C. 2020. Description of *Azoarcus nasutitermitis* sp. nov. and *Azoarcus rhizosphaerae* sp. nov., two nitrogen-fixing species isolated from termite nest and rhizosphere of *Ficus religiosa*. *Antonie Leeuwenhoek.* **113**, 933-946.
- Mallya, S.V., Udipi, V., Prabhu, S.N. and Narayana, S.K.K. 2018. Identification of *Ficus religiosa* by DNA SCAR marker. *Med. Sci.* **3**, 340-343.
- Mamidiseti, Y.D., Yammada, N., Siddamsetty, H.K., Bakshi, V. and Boggula, N. 2018. Phytochemical and analgesic, anti-inflammatory screening of methanolic extract of *Ficus religiosa* fruits: an *in vivo* design. *Pharma. innovation*, **7**, 69-74.
- Mamta, P.S., Poonam, S., Jyotika, S., Payal, B. and Dharendra, P.S. 2021. Antimicrobial activity of *Ficus religiosa* against some pathogenic microorganisms. *IJPRSE.* **2**, 759-763.
- Manorenjitha, M.S., Norita, A.K., Norhisham, S. and Asmawi, M.Z. 2013. GC-MS analysis of bioactive components of *Ficus religiosa* (Linn.) stem. *Int. J. Pharm. Bio. Sci.* **4**, 99-103.
- Moshi, A.A.M., Ravindran, D., Bharathi, S.R.S., Indran, S., Saravanakumar, S.S. and Liu, Y. 2020a. Characterization of a new cellulosic natural fiber extracted from the root of *Ficus religiosa* tree. *Int. J. Biol. Macromol.* **142**, 212-221.
- Moshi, A.A.M., Ravindran, D., Bharathi, S.R.S., Indran, S. and Priyadarshini, G.S. 2020b. Characterization of surface-modified natural cellulosic fiber extracted from the root of *Ficus religiosa* tree. *Int. J. Biol. Macromol.* **156**, 997-1006.
- Moshi, M.J., Innocent, E., Magadula, J.J., Otieno, D.F., Weisheit, A., Mbabazi, P.K. and Nondo, R.S.O. 2010. Brine shrimp toxicity of some plants used as traditional medicines in Kagera Region, north western Tanzania. *Tanzan. J. Health Res.* **12**, 63-67.
- Mousa, O., Vuorela, P., Kiviranta, J., Wahab, S.A., Hiltunen, R. and Vuorela, H. 1994. Bioactivity of certain Egyptian *Ficus* species. *J. Ethnopharmacol.* **41**, 71-76.
- Lakshmi HimaBindu, M.R., Angala Parameswari, S. and Gopinath, C. 2013. Determination of flavanoid content by *Ficus religiosa* Linn leaf extract by TLC and HPTLC. *Int. J. Pharm. Phytochem. Res.* **5**, 120-127.
- Murugesan, P., Rajaram, K., Victor, A.S., Pandian, M.R. and Manoharan, S. 2012. Evaluation on anti-inflammatory effects of *Ficus religiosa* (Linn.) in carrageenan induced acute inflammation in golden Syrian hamsters. *Elixir Int. J.* **51**, 11124-11128.

- Murugesu, S., Selamat, J. and Perumal, V. 2021. Phytochemistry, pharmacological properties, and recent applications of *Ficus benghalensis* and *Ficus religiosa*. *Plants* **10**, 2749.
- Nair, R. and Chanda, S. 2006. Activity of some medicinal plants against certain pathogenic bacterial strains. *Indian J. Pharmacol.* **38**, 142-144.
- Nair, R. and Chanda, S.V. 2007. Antibacterial activities of some medicinal plants of the western region of India. *Turk. J. Biol.* **31**, 231-236.
- Narayana, A., Tarannum, N., Shaik, M.S., Shobha, B.N., Sundar, R.M. and Lokesh, S.V. 2020. Synthesis of SnO₂ nanoparticles using *Ficus religiosa* leaf extract and their application in fabrication of OFETs for glucose monitoring. *Adv. Mater. Res.* **1159**, 67-77.
- Nawaz, H., Waheed, R. and Nawaz, M. 2020. Phytochemical composition, antioxidant potential and medicinal significance of *Ficus*. *Intech Open.* **1**, 20.
- Pandit, R., Phadke, A. and Jagtap, A. 2010. Antidiabetic effect of *Ficus religiosa* extract in streptozotocin-induced diabetic rats. *J. Ethnopharmacol.* **128**, 462-466.
- Parab, A.R., Han, K.Y., Chew, B.L. and Subramaniam, S. 2021. Morphogenetic and physiological effects of LED spectra on the apical buds of *Ficus carica* var. Black Jack. *Sci. Rep.* **11**, 23628.
- Patil, P., Barsagade, P. and Umekar, Dr.M.J. 2020. A review on *ficus religiosa* an alternative treatment for heart blockage. *Sch. Acad. J. Pharm.* **9**, 108-119.
- Pochhi, M. and Muddeshwar, M. 2017. Hypoglycemic and antihyperlipidemic effect of aqueous leaves extract of *Ficus religiosa* in alloxan induced diabetic rats. *Asian J. Med. Sci.* **8**, 50-55.
- Prakash, V., Gandotra, S., Kumar, P. and Singh, N. 2017. Phytochemical screening and antimicrobial activity of *Ficus religiosa*. *J. Pharm. Sci. Res.* **9**, 100-101.
- Preethi, R., Devanathan, V.V. and Loganathan, M. 2010. Antimicrobial and antioxidant efficacy of some medicinal plants against food borne pathogens. *Adv. Biol. Res.* **4**, 122-125.
- Priyanka, K., Kosuru, R., Sharma, R.P., Sahu, P.L. and Singh, S. 2017. Assessment of pharmacokinetic parameters of lupeol in *Ficus religiosa* L. extract after oral administration of suspension and solid lipid nanoparticles to Wistar rats. *J. Drug Deliv. Sci. Technol.* **41**, 58-67.
- Priyanka, K., Sahu, P. L. and Singh, S. 2018. Optimization of processing parameters for the development of *Ficus religiosa* L. extract loaded solid lipid nanoparticles using central composite design and evaluation of antidiabetic efficacy. *J. Drug Deliv. Sci. Technol.* **43**, 94-102.
- Raghavendra, G., Mahija, D., Sreevani, T., Yamala, A. and Bindu, P.H. 2017. Cell viability studies of green synthesised ZnO nanoparticles for antibacterial properties. *Am. J. Mater. Sci.* **2**, 56-60.
- Rahman, A. and Prasanna, A. 2018. Characterization of silver nanoparticles biosynthesized using *Ficus religiosa* plant leaf extract. *Int. Res. J. Eng. Tech.* **5**, 1449-1452.
- Raisagar, A., Kaur, C.D., Sawarkar, H.A., Kumar, L., Raisagar, A., Karmakar, A. and Sahu, M. 2019. Comparative study of wound healing effect of bark extracts of *Ficus religiosa* & *Ficus benghalensis* by mice model. *J. Pharmacogn. Phytochem.* **8**, 1815-1821.
- Rajiv, P. and Sivaraj, R. 2012. Screening for phytochemicals and antimicrobial activity of aqueous extract of *Ficus religiosa* Linn. *Int. J. Pharm. Pharm. Sci.* **4**, 207-209.
- Rajkumar, K., Ragupathi, M., Lee, Y.S. and Selvan, R.K. 2021. Preparation of sponge-like porous carbon from *Ficus religiosa* leaf and its K-ion intercalation properties. *Mater. Lett.* **301**, 130298.
- Rathod, V.D., Digambar, N.W., Pillai, S., Bhangale, J.O. and Bhangale, P.J. 2018. Antiarthritic activity of ethanolic extract of *Ficus religiosa* leaves in FCA induced arthritis in rats. *World J. Pharm. Res.* **7**, 778-789.
- Roy, A., Bhattacharya, T. and Kumari, M. 2020. Air pollution tolerance, metal accumulation and dust capturing capacity of common tropical trees in commercial and industrial sites. *Sci. Total Environ.* **722**, 137622.
- Sankar, R., Maheswari, R., Karthik, S., Shivashangari, K.S. and Ravikumar, V. 2014. Anticancer activity of *Ficus religiosa* engineered copper oxide nanoparticles. *Mater. Sci. Eng. C* **44**, 234-239.
- Sapkota, S.B., Khanal, A., Neupane, N., Karki, P., Sharan, V.K., Marasini, D.R., Budha, C.B., Barakoti, H., Rokaya, R.K., Pandey, J., Adhikari, A. and Bhandari, R. 2021. Evaluation of antiepileptic activities of *Ficus religiosa* bark and *Aegle marmelos* leaves using Swiss albino mice. *Int. J. Green Pharm.* **15**, 39-44.
- Sastry, A. and Barua, D. 2017. Leaf thermotolerance in tropical trees from a seasonally dry climate varies along the slow-fast resource acquisition spectrum. *Sci. Rep.* **7**, 11246.
- Selvan, A. and Chourasia, V. 2017. Hepatoprotective activity of *ficus religiosa* leaf extract in rats. *Curr. Res. Pharm. Sci.* **7**, 64-68.

- Senthilkumar, S., Chidambaram, S. and Manimekalai, R. 2020. Effect of optical, mechanical and thermal properties of bio-organic chlorophyll-a of *Ficus religiosa* added ADP optical single crystal: a novel NLO material. *Optik* **211**, 164530.
- Senthilkumar, S. and Manimekalai, R. 2020. Impact of bio-organic additive chlorophyll-b of *Ficus religiosa*, on optical, thermal, mechanical behaviors of KDP optical single crystals: A novel NLO material. *Optik* **204**, 164045.
- Sethuramani, A., Soundariya, R. and Dharani, V. 2021. Morpho-Playnological study on *Ficus religiosa* L. Leaves. *J. pharmacogn. Phytochem.* **10**, 1358-1365.
- Shahid, A., Saddiqe, Z. and Jabeen, K. 2021. 70. Antifungal and antioxidant activity of stem bark extracts of *Ficus religiosa* L. *Pure Appl. Biol.* **5**, 1304-1315.
- Shaikh, A., Tekale, S., Wagh, S. and Padul, M. 2020. Metabolite profiling of arginase inhibitor activity guided fraction of *Ficus religiosa* leaves by LC-HRMS. *Biomed. Chromatogr.* **34**, 4966.
- Sharma, R.K., Goyal A.K., Yadav, S.K. and Bhat R.A. 2013. Anti-fertility activity of *Ficus religiosa* fruits extract on goat uterus *in vitro*. *Int. J. Drug Dev. Res.* **5**, 330-335.
- Singh, D. and Goel, R.K. 2009. Anticonvulsant effect of *Ficus religiosa*: role of serotonergic pathways. *J. Ethnopharmacol.* **123**, 330-334.
- Singh, D., Singh, B. and Goel, R. K. 2011. Traditional uses, phytochemistry and pharmacology of *Ficus religiosa*: a review. *J. Ethnopharmacol.* **134**, 565-583.
- Soni, N. and Dhiman, R.C. 2020. Larvicidal and antibacterial activity of aqueous leaf extract of Peepal (*Ficus religiosa*) synthesized nanoparticles. *Parasite Epidemiol. Control.* **11**, 00166.
- Subash, B., Vijayan, P. and Baranitharan, M. 2019. Laboratory efficacy of *Ficus religiosa* extracts against mosquito-borne disease. *Indian J. Appl. Res.* **9**, 1-2.
- Sultana, B. and Anwar, F. 2008. Flavonols (kaempferol, quercetin, myricetin) contents of selected fruits, vegetables and medicinal plants. *Food Chem.* **108**, 879-884.
- Suriyakalaa, U., Ramachandran, R., Doulatunnisa, J.A., Aseervatham, S.B., Sankarganesh, D., Kamalakkannan, S., Kadalmani, B., Angayarkanni, J., Akbarsha, M.A. and Achiraman, S. 2021. Upregulation of Cyp19a1 and PPAR- γ in ovarian steroidogenic pathway by *Ficus religiosa*: a potential cure for polycystic ovary syndrome. *J. Ethnopharmacol.* **267**, 113540.
- Tahir, A., Khan, N.A., Mansha, M.Z., Ikram, K., Aslam, H.M.U., Aatif, H.M., Hanif, C.M.S. and Ashfaq, M. 2020. Yield analysis of oyster mushroom (*Pleurotus ostreatus*) on *Ficus religiosa* leaves in combination with agricultural waste materials. *Pure Appl. Biol.* **10**, 12-18.
- Tariq, M., Farooq, U., Athar, M., Salman, M., Tariq, M., Shahida, S. and Farooqi, Z.H. 2019. Fluoride removal using simple protonated and xanthate modified protonated *Ficus religiosa* branch powder in a fixed-bed column. *Desalin. Water Treat.* **150**, 204-212.
- Tariq, M., Farooq, U., Athar, M., Salman, M., Tariq, M., Shahida, S. and Farooqi, Z.H. 2021. Lab-scale continuous flow studies for comparative biosorption of cadmium (II) on untreated and xanthated *Ficus religiosa* biomass. *Water Environ. Res.* **93**, 2681-2695.
- Thanomchat, P., Paopun, Y. and Kermanee, P. 2019. Character and property evaluation of sacred fig tree (*Ficus religiosa* L.: Moraceae) Wood. *Microsc. Microanal. Res.* **32**, 30-33.
- Tiwari, P., Nathiya, R. and Mahalingam, G. 2017. Antidiabetic activity of endophytic fungi isolated from *Ficus religiosa*. *Asian J. Pharm. Clin. Res.* **10**, 59-61.
- Tripathy, K.C., Sahu, B.K., Panda, N.C. and Nayak, B.C. 1984. Toxicity of tannic acid in goats. *Indian J. Anim. Sci.* **54**, 1091-1093.
- Uma, B., Prabhakar, K. and Rajendran, S. 2009. *In vitro* antimicrobial activity and phytochemical analysis of *Ficus religiosa* L. and *Ficus bengalensis* L. against diarrhoeal enterotoxigenic *E. coli*. *Ethnobot. Leaflet.* **13**, 472-474.
- Utami, W., Aziz, H.A., Fitriani, I.N., Zikri, A.T., Mayasri, A. and Nasrudin, D. 2020. *In silico* anti-inflammatory activity evaluation of some bioactive compound from *Ficus religiosa* through molecular docking approach. *J. Phys. Conf. Ser.* **1563**, 012024
- Utami, W., Aziz, H.A., Nasrudin, D., Kusmawan, A., Anwar, Z., Maulana, M. and Daryanto, M., 2021. Investigating the potency of bioactive compounds from *Ficus religiosa* as anti-inflammatory agent. *J. Phys. Conf. Ser.* **1869**, 012022.
- Valsaraj, R., Pushpangadan, P., Smitt, U.W., Adersen, A. and Nyman, U. 1997. Antimicrobial screening of selected medicinal plants from India. *J. Ethnopharmacol.* **58**, 75-83.
- Vianna Filho, M. D. M., Alves, R.J.V., Peng, YQ. and Pereira, R.A.S. 2017. Naturalization of the Bodhi fig tree (*Ficus religiosa* L.-Moraceae) in Brazil. *Biosci. J.* **33**, 177-182.

- Vinutha, B., Prashanth, D., Salma, K., Sreeja, S.L., Pratiti, D., Padmaja, R., Radhika, S., Amit, A., Venkateshwarlu, K. and Deepak, M. 2007. Screening of selected Indian medicinal plants for acetylcholinesterase inhibitory activity. *J. Ethnopharmacol.* **109**, 359-363.
- Yadav, Y.C. 2015. Hepatoprotective effect of *Ficus religiosa* latex on cisplatin induced liver injury in Wistar rats. *Rev. Bras. Farmacogn.* **25**, 278-283.
- Yadav, Y.C. and Srivastava, D.N. 2013. Nephroprotective and curative effects of *Ficus religiosa* latex extract against cisplatin-induced acute renal failure. *Pharm. Biol.* **51**, 1480-1485.
- Yueniwati, Y., Syaban, M.F.R., Erwan, N.E., Putra, G.F.A. and Krisnayana, A.D. 2021. Molecular docking analysis of *Ficus religiosa* active compound with anti-inflammatory activity by targeting tumour necrosis factor alpha and vascular endothelial growth factor receptor in diabetic wound healing. *Open Access Maced. J. Med. Sci.* **9**, 1031-1036.
- Zaidi, S.F.H., Yamada, K., Kadowaki, M., Usmanghani, K. and Sugiyama, T. 2009. Bactericidal activity of medicinal plants, employed for the treatment of gastrointestinal ailments, against *Helicobacter pylori*. *J. Ethnopharmacol.* **121**, 286-291.
- Zeng, Y., Zeng, W., Zhang, Y., Ye, W., Cheng, D., Kanzaki, N. and Giblin-Davis, R.M. 2018. Morphological and molecular characteristics of *Parasitodiplogaster religiosae* n. sp. (Nematoda: Diplogastrina) associated with *Ficus religiosa* in China. *Plos one.* **13**, 0199417.