

LARVICIDAL EFFICACIES OF FOUR INDIGENOUS PLANT EXTRACTS AGAINST RED FLOUR BEETLE, *TRIBOLIUM CASTANEUM* (HERBST) (COLEOPTERA: TENEBRIONIDAE)

MD. ADNAN AL BACHCHU¹, KISMOT ARA², MD. NIZAM UDDIN AND ROUSHAN ARA³
Department of Entomology, Hajee Mohammad Danesh Science and Technology University (HSTU), Dinajpur, Bangladesh

Abstract

An investigation was carried out on the larvicidal efficacy of four indigenous plant extracts against the 6th instar larvae of red flour beetle, *Tribolium castaneum* (Herbst) during the period of February to July 2016. The result revealed that all the four plant extracts were effective in checking insect infestation and had different toxic effects against the 6th instar larvae of red flour beetle. Average mortality indicated that castor plant extract showed the highest toxic effect (average mortality 57.78%) whereas the lowest toxicity (average mortality 27.46%) was found in the neem leaf extracts. The larval mortality significantly differed among all the concentrations of the plant extracts applied and the highest mortality (60.75%) was recorded in maximum concentration (10.61 mg/cm²) of the plant extract. No larval mortality was observed in untreated control up to 72 HATs. Mortality percentage was also observed directly proportional to the level of concentrations of plant extracts and to the exposure period. Probit analysis of larval mortality also revealed that the castor plant extract showed the highest toxicity at different time interval among all the plant extracts applied.

Key words: Plant extracts, Larvicide, Mortality, Toxicity, *Tribolium castaneum*

Introduction

Stored agricultural products are attacked by more than 600 species of beetles, 70 species of moths and about 355 species of mites which cause quantitative and qualitative losses (Rajendran and Sriranjini 2008). These insect pests are the major cause of grain loss during post harvest storage, particularly in the tropical countries (Rajashekar *et al.* 2012). It was estimated that more than one-third of the food products are lost due to various pests during post-harvest storage (Tripathi *et al.* 2009). During storage food grain losses due to insect infestation are the most serious problem, specifically in the developing countries like Bangladesh (Dubey *et al.* 2008). Among the storage pests, Red flour beetle, *Tribolium castaneum* (Herbst) (Coleoptera: Tenebrionidae) is one of the most comprehensive and destructive pests of stored products, feeding on different stored grain and grain products such as peas, beans, cocoa, nuts, dried fruits and spices but milled grain products such as flour is the most preferred food (Mishra *et al.* 2012a and 2012b). They are attracted to grain with high moisture content where they encourage mold growth and produce a displeasing, musty odor.

Currently, different kinds of preventive and curative control measures are practiced to protect insect pests. Among them, chemical pesticides have been used for a long time, but have serious drawbacks (Sharaby 1988). Indiscriminate and continuous use of chemical insecticides create several problems in agroecosystem such as direct toxicity to beneficial insects, fishes and human (Goodland *et al.* 1985), pesticide resistance, increased environmental and social costs (Pimentel *et al.* 1980). To overcome these problems, it is utmost necessary to search the alternative pest control methods. The botanical pesticides which defend stored grains from pests are very helpful because of diverse distinct advantages. Botanical pesticides are of broad spectrum, safe to apply, unique in action and can be easily processed. Maximum botanical insecticides are non-hazardous and non-toxic to human. The main advantages of botanicals are that they are easily produced by farmers and are less expensive.

Interest in the use of botanical insecticides has increased over the past years (Isman 2000). Scientists in different parts of the world are working for the upliftment and induction of plant based pesticide, generally known as phytopesticide, botanical pesticide, biopesticide or natural pesticides (Siddiqui *et al.* 2009). Plant extracts contain compounds that show ovicidal, repellent, antifeedant and toxic effects in insects (Isman 2006). Previously investigations had been conducted on the toxicity effects of indigenous plant extracts against the adult stages of red flour beetle (Mahdeem 1998, Morgan 2009 and Joel 2015) but few research works had focused on the efficacies of plant extracts directly on the larval mortality of red flour beetle (Khalequzzaman and Sultana, 2006 and Yasir *et al.* 2012). Therefore, the present study was undertaken to evaluate the larvicidal efficacy of neem, thorn apple, castor and custard apple plant extracts against the larvae of *Tribolium castaneum*.

Materials and Methods

The present study on the larvicidal efficacies of four indigenous plant extracts against red flour beetle, *Tribolium castaneum* (Herbst) was conducted in the laboratory of the Department of Entomology, Hajee Mohammad Danesh Science and Technology University (HSTU), Dinajpur during the period of February to July 2016.

Collection and preparation of plant extract: The fresh mature plant leaves of custard apple (*Annona reticulata*), neem (*Azadirachta indica*), castor (*Ricinus communis*) and thorn apple (*Datura stramonium*) were collected from the HSTU campus, Dinajpur and the nearest area. Collected plant materials were air dried in shade followed by one day sun dried for 4 hours. The dried leaves were made powder separately by an electric grinder in the laboratory. The dust was passed through a 60 μ m mesh sieve to get fine powder. The powdered leaves were extracted in the methanol solvent. Hundred gram of every plant powders were taken separately in a 500 ml conical flask and mixed with 300 ml of methanol. The mixture was stirred for 30 minutes and then allowed to shake in the shaker machine for 24 hours. Next they were filtered through a filter paper (Whatman no. 1) and allowed to evaporate the solvents in the vacuum rotary evaporator and finally

hard different color extracts were collected. The collected crude extracts were preserved in tightly corked vials and stored in a refrigerator for further experimental use.

Preparation of food medium: Standard mixture of wheat flour with powdered dry yeast in a ratio of 19:1 was used as food medium (Park 1962). Food medium was sterilized at 60 °C temperature for 6 hours in an oven. After sterilization food was not used until at least 15 days to recover its moisture contents (Mondal 1984). In the micrometer sieve, both flour and yeast were passed through and then were mixed thoroughly for homogeneous mixing.

Collection and rearing of Tribolium castaneum: Adult beetles were collected from naturally infected wheat flour from the local market of Dinajpur town. Beetles were reared in glass beaker (500 ml) with the food medium (wheat flour). The beakers were kept in an incubator at 30 ± 0.5 °C temperature without light and humidity control. About 500 adults in each beaker were introduced with 500 g of wheat flour. The cultures were checked regularly and eggs along with larvae were separated to increase the population properly. A crumpled filter paper was placed inside the beaker for the easy movement of the beetles as well as to avoid the cannibalism of eggs. The beakers were covered with pieces of muslin cloth tightly fixed with the help of rubber bands to avoid possible escape of the beetles.

Determination of larval instars: About 500 beetles from the culture were placed in a beaker containing standard food medium. The beaker was covered with a cloth and kept in an incubator at 30 °C for egg collection. In regular interval, the eggs were collected by sieving the food medium by two sieves of 500 and 250 µm mesh separating the adults and eggs, respectively following the methods of Khan and Selman (1981). For egg collection, mainly 250 µm sieves were used. The collected eggs were kept in 60 mm in diameter petridishes and incubated at the same temperature (30 °C). Eggs hatched after 3 – 5 days and the newly hatched larvae were collected with a fine camel hair brush and then transferred to the fresh food medium for culture. The larval instars were determined by counting the number of exuviate (larval skin) deposited in the food medium (Mondal 1984). The 2nd day larvae were found as first instar larvae while the second, third, fourth, fifth and sixth instar larvae were found from the larval culture on 3rd, 6th, 9th, 12th and 16th day after hatching, respectively. Larval culture was also maintained in an incubator in the temperature at $30^{\circ} \text{C} \pm 0.5^{\circ} \text{C}$ without light and humidity control. The food medium was replaced by three days interval to avoid conditioning by the larvae.

Bioassay test (mortality test): Residual film method (Busvine 1971) was used to larvicidal test of different plant extracts against the 6th instar larvae of *Tribolium castaneum* (Herbst). One ml plant extract of each dose was dropped separately on petridishes (60 mm) with the help of pipette, covering uniformly the whole area of the petridish internally. The petridishes were then kept open for sometimes to evaporate the

solvents fully. Then 10 larvae of 6th instar were released in each petridish. Only methanol solvent was used for the control treatment. Three replications were made for each dose of all the treatments. The petridishes were then kept without food in the laboratory and larval mortality was recorded at 12, 24, 36, 48, 60 and 72 hours after treatments (HAT). The percentage of mortality was corrected using Abbott's formula (Abbott 1987).

$$P = \frac{P' - C}{100 - C} \times 100$$

Where,

P = Percentage of corrected mortality

P' = Observed mortality (%)

C = Mortality (%) at control.

Statistical analysis: The experiment was conducted using CRD. The data obtained from the experiments were statistically analyzed by MSTATC computer program. The significance of the mean difference was tested by DMRT. The observed mortality was also subjected to probit analysis.

Results and Discussion

The results of the experiment are presented and discussed under the following subheadings:

Toxicity effect of different plant extracts against 6th instar larvae of red flour beetle: Average mortality percentages of 6th instar larvae of red flour beetle at 12, 24, 36, 48, 60 and 72 hours after treatment (HAT) indicated that castor plant extract showed the highest (average mortality, 57.78%) toxic effect, whereas neem plant extract showed the lowest (average mortality, 27.46%) toxic effect (Table 1). Mortality percentages of four plant extracts were directly proportional to the time after treatment. There was significant difference ($p < 0.01$) among the toxicity effects of plant extracts applied on the 6th instar larva of *T. castaneum*. The order (highest to lowest) of toxicity effect of four plant extracts against the 6th instar larvae of red flour beetle were: castor > custard apple > thorn apple > neem. Mortality percentage also differed significantly among all the concentration level at different time interval (Table 2). The highest mortality (60.75%) was at the maximum concentration (10.61 mg/cm²) of plant extract. Mortality percentage is directly proportional to the level of concentration. The interaction of different plant extracts of different doses at different time interval indicated that castor plant extracts showed the highest average per cent larval mortality at maximum dose (10.61 mg/cm²) which was statistically different from all other plant extract at different concentration level (Table 3). Mortality percentages of red flour beetle of different plant extracts of different dose level at different hours were found statistically significant.

Table 1. Toxicity effect of different plant extracts against 6th instar larvae of *Tribolium castaneum* at different HAT (interaction of plant extracts and time).

Plant extracts used	Percentage of larval mortality at different time intervals						Average mortality
	12HAT	24HAT	36HAT	48HAT	60HAT	72HAT	
Neem	6.667 c	17.22 c	22.78 c	31.48 d	35.80 c	50.79 c	27.46 d
Thorn apple	9.444 bc	16.11 c	28.33 c	36.79 c	43.83 b	60.26 b	32.46 c
Custard apple	12.78 b	24.44 b	42.22 b	51.67 b	73.27 a	79.88 a	47.38 b
Castor	22.22 a	43.89 a	55.56 a	70.00 a	75.80 a	79.20 a	57.78 a
LSD	4.104	4.606	6.368	5.182	4.340	3.996	2.652
CV (%)	47.92	27.04	25.53	16.28	11.32	8.83	9.59
s _x	1.443	1.620	2.240	1.822	1.526	1.405	0.9327

HAT = Hour after treatment. Within column values followed by different letter(s) are significantly different at 5% level of probability by DMRT.

From the above result, it is apparent that among the tested four plant extracts, castor showed the most toxic effect against the 6th instar larvae of the red flour beetle. The reduction of larval population by using the leaf extracts of castor is similar to the previous findings of Basheer (2014). He reported that the castor leaf extract obtained as the best with the mortality of the larvae of *Anopheles arabiensis* was 96% after 24 hours with an LC₅₀ at 0.390 mg/l, 100% mortality was observed after 48 hours with LC₅₀ at 0.284 mg/l. Collavino *et al.* (2006) reported that castor bean leaf powder is effective

Table 2. Toxicity effect of different doses of plant extracts against 6th instar larvae of *Tribolium castaneum* at different HAT (interaction of dose and time).

Doses	Percentage of larval mortality at different time intervals						Average mortality
	12HAT	24HAT	36HAT	48HAT	60HAT	72HAT	
10.61	21.67 a	40.83 a	57.50 a	68.52 a	80.74 a	95.23 a	60.75 a
5.30	18.33 ab	35.83 a	52.50 a	63.24 ab	76.95 a	89.06 b	55.99 b
2.65	15.00 bc	29.17 b	43.33 b	58.98 b	67.87 b	80.78 c	49.19 c
1.33	12.50 cd	25.00 bc	37.50 bc	50.09 c	60.65 c	72.71 d	43.08 d
0.66	9.167 d	21.67 c	32.50 c	44.07 c	56.85 c	67.40 e	38.61 e
Control	0.00 e	0.00 d	0.00 d	0.00 d	0.00 d	0.00 f	0.00 f
LSD	5.027	5.641	7.799	6.346	5.315	4.895	3.248
CV (%)	47.92	27.04	25.53	16.28	11.32	8.83	9.59
s _x	1.768	1.984	2.743	2.232	1.869	1.721	1.142

HAT = Hour after treatment. Within column values followed by different letter(s) are significantly different at 5% level of probability by DMRT.

Table 3. Toxicity effect of different plant extracts of different doses against 6th instar larvae of *Tribolium castaneum* at different HAT (interaction of plant, dose and time).

Plant extracts used	Doses (mg/cm ²)	Percentage of larval mortality at different time intervals						
		12HAT	24HAT	36HAT	48HAT	60HAT	72HAT	Average mortality
Neem	10.61	13.3 cdef	33.33 de	46.67 de	53.71 ef	62.97 c	95.83 ab	50.97 ghi
	5.30	10.0 defg	30.00 de	36.67 efg	49.63 fg	55.56 cd	78.57 cd	43.41 j
	2.65	6.66 efg	16.67 fgh	26.67 fgh	50.00 fg	48.15 de	64.88 ef	35.51 k
	1.33	6.66 efg	13.33 gh	13.33 hi	17.78 i	29.63 f	39.28 g	20.00 m
	0.66	3.33 fg	10.00 hi	13.33 hi	17.78 i	18.52 g	26.19 h	14.86 m
	0.00	0.00 g	0.00 i	0.00 i	0.00 j	0.00 h	0.00 i	0.00 n
	10.61	20.00 bcd	36.67 cd	50.00 cde	60.37 def	66.67 c	88.4 abc	53.69 fghi
Thorn apple	5.30	13.33 cdef	26.67 def	43.33 def	50.00 fg	66.67 c	84.72 bc	47.45 ij
	2.65	10.00 defg	16.67 fgh	26.67 fgh	39.26 gh	48.15 de	68.98 de	34.95 k
	1.33	6.667 efg	10.00 hi	26.67 fgh	39.26 gh	40.74 ef	61.58 ef	30.82 kl
	0.66	6.667 efg	6.667 hi	23.33 gh	31.85 h	40.74 ef	57.87 f	27.86 l
	0.00	0.00 g	0.00 i	0.00 i	0.00 j	0.00 h	0.00 i	0.00 n
	10.61	20.00 bcd	33.33 de	56.67 bcd	70.00 bcd	96.67 a	100.0 a	62.78 cde
	5.30	16.6 bcde	30.00 de	56.67 bcd	66.67 cde	92.59 ab	100.0 a	60.43 def
Custard apple	2.65	16.6 bcde	30.00 de	53.33 cde	63.33 cdef	85.93 ab	92.96 ab	57.04 efg
	1.33	13.33 cdef	30.00 de	50.00 cde	60.00 def	82.59 b	93.33 ab	54.88 fgh
	0.66	10.00 defg	23.33 efg	36.67 efg	50.00 fg	81.85 b	92.96 ab	49.13 hij
	0.00	0.00 g	0.00 i	0.00 i	0.00 j	0.00 h	0.00 i	0.00 n
	10.61	33.33 a	60.00 a	76.67 a	90.00 a	96.67 a	96.67 a	75.56 a
	5.30	33.33 a	56.67 ab	73.33 ab	86.67 a	92.96 ab	92.96 ab	72.65 ab
	2.65	26.67 ab	53.33 ab	66.67 abc	83.33 ab	89.26 ab	96.30 a	69.26 abc
Castor	1.33	23.33 abc	46.67 bc	60.00 abcd	83.33 ab	89.63 ab	96.67 a	66.61 bcd
	0.66	16.6 bcde	46.67 bc	56.67 bcd	76.67 abc	86.30 ab	92.59 ab	62.59 cde
	0.00	0.00 g	0.00 i	0.00 i	0.00 j	0.00 h	0.00 i	0.00 n
	LSD	10.05	11.28	15.60	12.69	10.63	9.789	6.496
	CV (%)	47.92	27.04	25.53	16.28	11.32	8.83	9.59
	s _x	3.536	3.967	5.486	4.464	3.738	3.443	2.285

HAT = Hour after treatment. Within column values followed by different letter(s) are significantly different at 5% level of probability by DMRT.

against male moth larvae, *Plodia interpunctella* HBN (Lepidoptera: Phycitinae). Castor bean oil and pure compounds of *R. communis* had been reported to exhibit high toxic effects in target animals (Kumar *et al.* 2007). Castor bean also contains the alkaloid ricinin, the polyphenolic molecule epicatechin and fatty acids in their leaves (Zahir *et al.* 2012) all which have insecticidal properties. The biological activity of castor plant extracts might be attributed to its alkaloid contents such as saponins, lectins, trypsin inhibitor etc. which caused mortality to the larvae.

Table 4. Relative toxicity (probit analysis) of different plant extracts treated against 6th instar larvae of *Tribolium castaneum* after 12, 24, 36, 48, 60 and 72 HAT.

Plant extracts used	No of larvae used	LD ₅₀ values (mg/cm ²)	95% fiducially limits		χ^2 values (df)
			Lower	Upper	
12HAT					
Neem	30	1321.64	0.29	5856718	0.27 (3)
Thorn apple	30	2464.44	0.02	2.916262E ⁺⁰⁸	0.13 (3)
Custard apple	30	443.13	1.16	168276.4	0.24 (3)
Castor	30	77.75	1.27	4730.86	0.26 (3)
24HAT					
Neem	30	37.25	5.02	276.12	0.41 (3)
Thorn apple	30	4105.10	1.98E ⁻⁰⁶	8.4938E ⁺¹²	0.28 (3)
Custard apple	30	23.47	6.43	85.67	0.08 (3)
Castor	30	1.50	0.26	8.59	0.14 (3)
36HAT					
Neem	30	12.78	4.95	33.03	0.66(3)
Thorn apple	30	2.37	0.77	7.32	0.53(3)
Custard apple	30	12.91	3.07	54.17	0.87(3)
Castor	30	0.32	0.03	3.31	0.01(3)
48HAT					
Neem	30	5.82	3.15	10.77	3.61 (3)
Thorn apple	30	0.47	0.04	5.04	0.16 (3)
Custard apple	30	4.55	1.90	10.91	0.51 (3)
Castor	30	0.0096	4.07E ⁻⁰⁶	22.99	0.08 (3)
60HAT					
Neem	30	4.07	2.42	6.84	0.72 (3)
Thorn apple	30	0.049	0.001	1.61	0.83 (3)
Custard apple	30	2.02	0.97	4.19	1.30 (3)
Castor	30	0.0093	1.08E ⁻⁰⁵	8.15	0.28 (3)
72HAT					
Neem	30	1.67	1.24	2.26	0.84 (3)
Thorn apple	30	0.002	5.79E ⁻⁰⁸	74.72	0.21 (3)
Custard apple	30	0.96	0.55	1.67	1.07 (3)
Castor	30	1.7E ⁻⁰⁷	5.88E ⁻³²	5.07E ⁺¹⁷	0.17 (3)

HAT = Hour after treatment.

Values were based on five concentrations, three replications of 10 insects each.

χ^2 = Goodness of fit.

The tabulated value of χ^2 is 5.99 (d. f = 2 at 5% level).

Probit analysis for direct toxic effect of different plant extracts against 6th instar larvae of red flour beetle: Probit analysis for direct toxic effect of red flour beetle at 12, 24, 36, 48, 60 and 72 HAT of different plant extracts against 6th instar larvae of red flour beetle are presented in Table 4. Among the treatments, LD₅₀ values at 12 HAT indicated that castor (77.75 mg/cm²) plant extract was the most toxic followed by custard apple (443.13 mg/cm²) plant extract while thorn apple plant extract (2464.44 mg/cm²) was the least toxic. Castor plant extract also maintained its highest toxicity when the LD₅₀ values were compared at 24, 36, 48, 60 and 72 HAT (1.50, 0.32, 0.0096, 0.0093 and 1.72E⁻⁰⁷ mg/cm²). The chi-square values were insignificant at 5% level of probability of different plant extracts at different HAT and mortality data did not show any heterogeneity.

From the results of the probit analysis on the 6th instar larvae of the red flour beetle, it is apparent that all the tested plants would be more or less effective for controlling red flour beetle but castor will be the most effective extracts against the 6th instar larval of the red flour beetle. This result is in agreement with results reported by Ramos-Lopez *et al.* (2012) who evaluated the effect of ingested ricin oil, ricinin and hexanic, acetatoethylic and methanolic extracts from 16 to 24,000 ppm on first instar *Spodoptera frugiperda* larva. All treatments with ricinin (560 ppm) and acetatoethylic extracts (1600 ppm) from *R. communis* seeds had reduced weight of the pupae by 21.6% to 4.9% respectively.

It may be concluded that the botanicals used in the present study had direct toxic effect on the 6th instar larval of *T. castaneum*. Among the tested plants, castor extracts showed the highest toxic effect. The larvicidal potential of indigenous plant extracts against *T. castaneum* has good prospects. Moreover, additional studies are needed to develop appropriate formulation and application method of *R. communis* based pesticides against stored product pest.

Acknowledgements

The authors express gratitude to the University Grant Commission, Bangladesh, for financial support through the Institute of Research and Training, Hajee Mohammad Danesh Science and Technology University, Dinajpur.

References

- Abbott, W.S. 1987. A method of computing the effectiveness of an insecticide. *J. American Mosquito Cont. Asso.* **3**(2): 302-303.
- Basheer, A.G.M. 2014. *Ricinus communis* (castor) as larvicide on *Anopheles arabiensis* Patton. *Inter. J. Advan. pharma. Biol. Chem.* **3**(2): 319–328.
- Busvine, J.R. 1971. A critical review of the techniques for test in insecticides. *Communication of Agricultural Bureau*, (CAB) London, U. K. pp. 263–288.
- Collavino, M., A. Pelicano and R.A. Giménez. 2006. Actividad insecticida de *Ricinus communis* L. sobre *Plodia interpunctella* HBN. (Lepidoptera: Phycitinae). *Revista Facultad de Ciencias Agropecuarias Universidad Nacional de Cuyo.* **38**: 13-18.

- Dubey, N.K., B. Srivastava and A. Kumar. 2008. Current status of plant product as botanical pesticides in storage pest management. *J. Biopest.* **1**(2): 182-186.
- Goodland, R., C. Watson and G. Ledec. 1985. Biocides bring positioning and pollution to third world. *The Bangladesh Observer*, 16th and 17th January 1995. p.3.
- Isman, M.B. 2000. Plant essential oils for pest and disease management. *Crop Prot.* **19**: 603-608.
- Isman, M.B. 2006. Botanical insecticides, de-terrents, and repellents in modern agriculture and an increasingly regulated world. *Ann. Review Entomol.* **51**: 45-66.
- Joel, O.O. 2015. Efficacy of selected plant extracts against *Tribolium castaneum* Herbst in stored groundnut (*Arachis hypogaea* L.). *African J. Plant Sci.* **9**(2): 90-96.
- Khalequzzaman, M. and S. Sultana. 2006. Insecticidal activity of *Annona squamosa* L seed extracts against the red flour beetle, *Tribolium castaneum* (Herbst). *J. Biol. Sci.* **14**: 107-112.
- Khan, A.R. and B.J. Selman. 1981. Some techniques for minimizing the difficulties in egg counting in *Tribolium castaneum* (Herbst). *The Entomol. Record J. Vari.* **93**: 36-37.
- Kumar, O., R.P.V. Lakshmana, S. Pradhan, R. Jayaraj, A.S. Bhaskar, A.B. Nashikkar and R. Vijayaraghavan. 2007. Dose dependent effect of ricin on DNA damage and antioxidant enzymes in mice. *Cell and Mol. Biol.* **53**: 92-102.
- Mahdeem, H. 1998. "*Annona reticulata*". Neglected Crops. Department of Horticulture & Landscape Architecture, Purdue University. Archived from the original on 11 May 2008. Retrieved 16 April 2008.
- Mishra, B.B., S.P. Tripathi and C.P.M. Tripathi. 2012a. Response of *Tribolium Castaneum* (Coleoptera: Tenebrionidae) and *Sitophilus oryzae* (Coleoptera: Curculionidae) to potential insecticide derived from essential oil of *Mentha arvensis* leaves. *Biol. Agri. Hort.* **28**: 34-40.
- Mishra, B.B., S.P. Tripathi and C.P.M. Tripathi. 2012b. Repellent effect of leaves essential oil from *Eucalyptus globules* (Mirtaceae) and *Ocimum basilicum* (Lamiaceae) against two major stored grain insect pests of coleopterans. *Nat. Sci.* **10**(2): 50-54.
- Mondal, K.A.M.S.H. 1984. A method of determining the larval instars of *Tribolium castaneum* Herbst (Coleoptera: Tenebrionidae). *Lab. Prac.* **33**(10): 120-121.
- Morgan, E.D. 2009. Azadirachtin, a scientific gold mine. *Bioorg. Med. Chem.* **17**(12): 4096-4105.
- Park, T. 1962. Beetles, competition and population. *Science* **138**: 1369-1375.
- Pimentel, D., D. Andow, R. Dyson-Hudson, D. Gallahan, S. Jacobson, M. Irish, S. Croop, A. Moss, I. Schreiner, M. Shepard, T. Thompson and B. Vinzant. 1980. Environmental and social cost of pesticides. A preliminary assessment. *Oikos.* **34**: 125-140.
- Rajashekar, Y., N. Bakthavatsalam and T. Shivanandappa. 2012. Botanicals as grain protectants. *Psyche.* **2012**: 1-13.
- Rajendran, S. and V. Sriranjini. 2008. Plant products as fumigants for stored product insect control. *J. Stored Prod. Res.* **44**: 126-135.
- Ramos-Lopez, M.A., M.M. Gonzalez-Chavez, N.C. Cardenas-Ortega, M.A. Zavala-Sanchez and G.S. Perez. 2012. Activity of the main fatty acid components of the hexane leaf extract of *R. communis* against *Spodoptera frugiperda*. *African J. Biotech.* **11**: 4274-4278.
- Sharaby, A. 1988. Evaluation of some myrtaceae plant leaves as protectants against the infestation by *Sitophilus oryzae* L. and *Sitophilus granarius* L. *Insect Sci. Appl.* **9**: 465-468.
- Siddiqui, B.S., S.T. Ali, R.M. Tariq, T. Gulzar, M. Rasheed and R. Mehmood. 2009. GC-based analysis of insecticidal constituents of the flower of *Azadirachta indica* A. Juss. *Nat. Prod. Res.* **23**(3): 271-283.

- Tripathi, A.K., S. Upadhyay, M. Bhuiyan and P.R. Bhattacharya. 2009. A Review on Prospects of Essential Oils as Biopesticides in Insect-Pest Management. *J. Pharmacol. Phyto.* **1**: 52-63.
- Yasir, M., M. Sagheer, M. Hasan, S.K. Abbas, S. Ahmad and Z. Ali. 2012. Growth, development and reproductive inhibition in the red flour beetle, *Tribolium castaneum* (Herbst) (Coleoptera: Tenebrionidae) due to larval exposure to flufenoxur on-treated diet. *Asian J. Pharma. Biol. Res.* **2**(1): 51-5.
- Zahir, A. A., A.A. Rahuman, A. Bagavan, K. Geetha, C. Kamaraj and G. Elango. 2012. Evaluation of medicinal plant extracts and isolated compound epicatechin from *R. communis* against *Poromphistomum cervi*. *Parasitol. Res.* **111**(4):1629-35.

(Revised copy received on 5-12-2017)