

SHORT COMMUNICATION

Influence of unsaturated to saturated ratio of fatty acids reaching the duodenum on postruminal digestion of stearic acid in Holstein steers fed a high-fat finishing diet

Alejandro Plascencia^{1,2} D, Alberto Barreras² D, Yissel Valdés-García² D, Richard A. Zinn³ Departamento de Ciencias Exactas y Naturales, Universidad Autónoma de Occidente, Guasave, México Instituto de Investigaciones en Ciencias Veterinarias, Universidad Autónoma de Baja California, Mexicali, México Department of Animal Science, University of California, Davis, CA, USA

ABSTRACT

Objective: To evaluate the influence of the unsaturated to saturated ratio of fatty acids (FAs) reaching the duodenum on postruminal digestion of FAs, mainly focused on stearic acid (C18:0). **Materials and Methods:** Six Holstein steers [208 \pm 3 kg initial live weight (LW)] with cannulas in the abomasum and proximal duodenum were used in a replicated 3 \times 3 Latin square design. Steers were fed a fixed amount of a basal steam-flaked corn-based diet containing 8% supplemental fat and were daily infused via abomasum with 0, 67, and 165 gm oleic acid (C18:1). The experiment lasted for 42 days.

Results: The daily total FA (TFA) intake (dietary FA intake plus abomasal infusion of oleic acid) represented a 1.78, 2.10, and 2.56 gm TFA/kg LW ratio. The unsaturated to saturated ratio of FAs entering the duodenum increased (p < 0.01) as level C18:1 infusion into the abomasum increased. Infusion of C18:1 tended (quadratic component, p = 0.07) to improve postruminal TFA digestion, being maximal for the 67 gm/day infusions. This increase in TFA digestion was due to increased (quadratic component, p = 0.03) postruminal C18:0 digestion (postruminal digestion of the other FAs was not different, $p \ge 0.13$).

Conclusion: Increasing the unsaturated to saturated ratio of FAs entering the small intestine will enhance intestinal C18:0 digestion. This positive effect is expected to be more likely beneficial when FA intake is high (and thus, the duodenal flow of FA is high), but this benefit looks diminished when the quantity of TFA reaching the intestine exceeds the proportion of 2.13 gm FA/kg LW.

ARTICLE HISTORY

Received July 19, 2021 Revised August 25, 2021 Accepted August 26, 2021 Published September 20, 2021

KEYWORDS

Fatty acid; postruminal digestion; steers; stearic acid; oleic acid; finishing diets



© The authors. This is an Open Access article distributed under the terms of the Creative Commons Attribution 4.0 License (http://creativecommons.org/licenses/by/4.0)

Introduction

The net energy of maintenance (NE_m) value assigned by the current standards [1] for supplemental fats for feedlot cattle is from 5.65 to 6.03 Mcal/kg. In as much as feedgrade fats largely (~90%) comprise fatty acids (FAs), their net energy value is closely associated with FA digestibility [2]. Fat intake level has the greatest impact on fat digestion [3,4]. This effect results from the limitations on intestinal absorption of saturated FA due to upper limits on bile production [5]. Another factor that limits intestinal FA digestion is the saturated-to-unsaturated FA ratio reaching the intestine. As a consequence of ruminal biohydrogenation of unsaturated FA, C18:0 accounts for 65%

or more of the total FA (TFA) entering the intestine. Stearic acid is absorbed almost similarly (90%) to that of unsaturated FA when fat is supplemented in diets at lower levels (i.e., <3%), but this proportions can drop to 80% or less at higher (\sim 6%) levels of fat supplementation [6]. Intestinal digestion of C18:0 alone explains >85% of the variation in net energy value of supplemental fat [7]. As a result, the energy value of supplementary lipids can be increased by lowering ruminal biohydrogenation and/or improving intestinal digestibility of C18:0. On the latter, a synergistic effect of small intestine absorption of saturated FAs in broilers has been determined when the proportion of the unsaturated to saturated ratio that reaches the intestine is

Correspondence Alejandro Plascencia ⊠ aplas_99@yahoo.com 🖫 Departamento de Ciencias Exactas y Naturales, Universidad Autónoma de Occidente, Guasave, México.

How to cite: Plascencia A, Barreras A, Valdés-García YS, Zinn RA. Influence of unsaturated to saturated ratio of fatty acids reaching the duodenum on postruminal digestion of stearic acid in Holstein steers fed a high-fat finishing diet. J Adv Vet Anim Res 2021; 8(3):465–470.

raised [8], increasing the energy value of supplemental fats. Likewise, in ruminants, protecting supplemental fat from ruminal biohydrogenation improves intestinal digestion of C18:0. Zinn et al. [9] reported that for each percentage unit increase in the proportion of unsaturated FA (mainly oleic acid) entering the small intestine, digestibility of C18:0 increased by 1%. This could be explained by oleic acid's significant amphiphilic properties, which increase the surface area of the bile salt micelle, increasing its hydrophobic capacity and thus alleviating the potential limitation of bile production associated with high-fat intakes, thereby improving the physical–chemical conditions for increased intestinal absorption of saturated FAs [4].

Due to the intense ruminal biohydrogenation of unsaturated FAs, it is complex to precisely manipulate the proportions of saturated to unsaturated FAs that reach the intestine. In ruminants, the reports on this subject are minimal. To our knowledge, the effect of an increasing proportion of unsaturated to saturated FAs flowing to the duodenum on the postruminal digestion of saturated FAs in cattle fed high-fat finishing diets has not been directly assessed. The purpose of this experiment was to determine the effect of increasing the proportion of unsaturated to saturated FAs in Holstein steers fed a high-fat finishing diet on postruminal digestion of stearic acid (C18:0). This was accomplished by directly infusing oleic acid (C18:1) into the abomasum (thereby avoiding ruminal biohydrogenation of oleic acid).

Material and methods

Ethical approval

The protocol of this experiment was approved by the University of California Animal Care and Use Committee (protocol #18811).

Characteristics of the experimental units, basal diet, treatments, and sampling methods

Six Holstein steers [208 ± 3 kg live weight (LW) at start of the experiment] with cannulas in abomasum [10] and proximal duodenum [tygon "T" cannula (1.9 cm i.d.) placed approximately 6 cm from pyloric sphincter] were used in a replicated 3 × 3 Latin square experiment to evaluate the influence of abomasal oleic acid infusion (C18:1) on characteristics of postruminal FA digestion. The steers were maintained in an individual neoprene-floor pen (2.35 × 3.92 m) and drawer feeder with *ad libitum* access to water. Intake of basal diet was limited to 4.60 kg/day (as is fed basis), which was equivalent to 2.2% of the initial LW or 74.6 gm of diet dry matter (DM)/kg LW^{0.75}), offered at equally twice daily (0800 and 2000 h) during all experiments which lasted for 42 days. Ingredient composition and chemical characteristics of the basal diet are shown

in Table 1. The external marker chromic oxide (Cr₂O₃) (4.0 gm/kg of diet) was employed to assess digestion coefficients. Before incorporating into the diet, the external marker was premixed for 5 min with other minor ingredients (trace mineral salt, limestone, and urea) using a 2.5 m capacity mixer. The treatments were as follows: 1) purified water infused via the abomasal cannula at 2.79 ml/h (67 gm/day); 2) oleic acid-infused via the abomasal cannula at 3.14 ml/h (67 gm/day), and 3) oleic acid-infused via the abomasum cannula at 7.72 ml/h (165 gm/day). The oleic acid-infused was oleic acid NLT 98% (relative density = 0.89; Cas# 112-80-1 Mallinckrodt Baker Inc., Phillipsburg, NI). The infusion was accomplished using a variable flow peristaltic pump (Variable Flow Mini-Pump II, VWR Scientific Products, West Chester, PA). Infusion tubing (3.1 mm d.i. × 5.1 m Tygon®; Norton, Akron OH 44309-3660) was moored at the ceiling above each pen using 1

Table 1. Composition of the basal diet fed to cannulated Holstein steers.

Ingredient composition	% in diet			
DM content	88.82			
Ingredient composition	DM basis			
Alfalfa hay	6.00			
Sudangrass hay	6.00			
Steam flaked corn	55.85			
Steam flaked wheat	10.00			
Tallow	8.00			
Cassava	7.00			
Cane molasses	3.00			
Limestone	1.37			
Urea	1.07			
Chromic oxide	0.40			
Dicalcium phosphate	0.16			
NaHCO ₃	0.75			
Trace mineral salt ^a	0.40			
Nutrient composition, % DM basis ^b				
Calculated crude protein	11.92			
Calculated neutral detergent fiber	13.66			
Determined TFA	9.05			
Calculated NE, Mcal/kg				
Maintenance	2.38			
Gain	1.68			

 $^{^{\}mathrm{a}}$ Trace mineral salt contained: CoSO $_{\mathrm{a}}$, 6.8 gm/kg; CuSO $_{\mathrm{a}}$, 10.4 gm/kg; FeSO $_{\mathrm{a}}$, 35.7 gm/kg; ZnO, 12.4 gm/kg; MnSO $_{\mathrm{a}}$, 10.7 gm/kg; KI, 0.52 gm/kg; and NaCl, 923.5 gm/kg.

^bCalculated from the tabular values for individual feed ingredients [1]; average DM and FA concentration of basal diet were determined in subsamples gathered and composited throughout the experiment.

m lengths of 6.3 mm latex tubing to provide constant tension, thus permitting free movement of steers during infusion. The experiment lasted for 42 days and consisted of 3 experimental periods of 14 days planned as follows: 3 days for infusion treatment adjustment, followed by a sampling period of 4 days; then, following the completion of each infusion period, steers were rested (free of infusion tubing) for 7 days but the intake of basal diet was maintained. Duodenal and fecal samples were taken from each steer following the procedure (sample quantity and schedule sampling) described by Núñez-Benítez et al. [11]. Once the sampling period finished, duodenal and fecal samples from each steer and within each sampling period were composited for subsequent analysis.

Laboratory analysis

Feed, duodenal, and fecal samples were subjected to the DM (oven drying at 105° C until no further weight loss) determination, Cr_2O_3 determination [12], and FA determination. According to Zinn and Plascencia's [3] protocols, direct methyl esterification was used to identify FAs [13], followed by gas chromatographic analysis [14]. The Cr intake estimated the total DM flow to the duodenum and excreted in feces (gm/day) versus concentration of Cr in the duodenal and fecal samples as follows: total DM output, gm/day = gm Cr_2O_3 intake daily/(g Cr_2O_3 /gm of duodenal/feces).

Statistical analysis

The effects of the treatments on the characteristics of postruminal FA digestion were analyzed as a replicated 3 × 3 Latin square design using the MIXED procedure of the Statistical Analytical System software [15]. The fixed effects consisted of treatment and period, and steer as a random effect. The statistical model for the trial was as follows: $Y_{ijkl} = \mu + C_i + A_j + P_k + E_{ijkl}$ where Y_{ijk} is the response variable, C_i is the oleic acid level effect, A_j is the animal effect, P_k is the period effect, and E_{ijk} is the residual error. Orthogonal polynomials were used to investigate the linear and quadratic effects of oleic acid infusion. Contrast coefficients for unequally spaced levels of oleic acid infusion were carried out using the ORPOL function [15]. Treatment effects were considered significant when p-value was ≤ 0.05 , and ≤ 0.10 .

Results and Discussion

In as much as DM intake of the basal diet was constant for all steers, differences in FA intake and saturated to unsaturated FA ratio between treatments was due to the C18:1 infused into the abomasum (Table 2). Accordingly, the daily relative FA intake (expressed as gm FA/kg LW) linearly increased (p < 0.01) from 1.78 (0 gm infused) to

2.63 (165 gm infused/day), while the proportion of unsaturated to saturated FA intake linearly increased (p < 0.01) from 1.10 (approximately 52% unsaturated FA) to 2.02 (approximately 67% unsaturated FA).

Duodenal flow, fecal excretion, and postruminal digestion of FA are shown in Table 3. Considering all treatments, the average TFA flow to the small intestine was 12% greater (average = 498 gm/day) than the average FA intake (446 gm/day). This differential reflects the ruminal indigestibility of dietary FA in combination with ruminal microbial FA synthesis. Analyzing data from other experiments carried out with feedlot cattle fed high-energy diets supplemented with fats which report FA intakes and FA intestinal flows [5,9,16-19], the relationship between FA intake versus the duodenal FA flow is as follows: Y = 1.134X ($R^2 = 0.91$), where Y = duodenal FA flow (gm/day) and X = FA intake. gm/day. When the equation is applied to the current experiment, the average anticipated flow is 505 gm/day, which is quite near the observed value (0.99). When no C18:1 was infused, the saturated to unsaturated ratio of FAs reaching the duodenum was 4.98. Duodenal flow (as % of TFA flow) of saturated FAs increased 1.7-fold (82.0%) when compared with the proportion of 47.5% of saturated FAs in the TFA intake. This effect is due to ruminal biohydrogenation of unsaturated FAs [20], which typically average 70% (ranging between 60% and 90%) [7]. As expected, the saturated to unsaturated ratio of FA entering the duodenum declined (p < 0.01) as level C18:1 infusion into the abomasum increased.

Infusion of C18:1 tended (quadratic component, p = 0.07) to increase postruminal FA digestion, being maximal for the 67 gm/day infusions. This increase in TFA digestion was due to increased (quadratic component, p = 0.03) postruminal C18:0 digestion (postruminal digestion of the others FA was not different, p > 0.13). Due to its greater facility for the formation of micelles, small

Table 2. Ingestion of FA in steers (208 kg LW) infused with 0, 67, or 165 gm of oleic acid/day.

Item	C18:1 infusion, gm/day			
	0	67	165	
Intake, gm/day				
DMª	4,086	4,154	4,222	
FAs (from feed)	370	376	382	
FAs infused	0	67	165	
TFA intake	370	443	547	
FA intake, gm FA/kg LW	1.78	2.13	2.63	
Saturated FA intake	176	176	176	
Unsaturated FA intake	194	260	356	
Unsaturated: saturated FA ratio	1.10	1.48	2.02	

^aInclude DM of feed plus DM of FA infusion.

Table 3. Treatments' effects on postruminal digestion of FA of Holstein steers (208 kg LW).

Item	C18:1	C18:1 infusion, gm/day			Contrast p-value	
	0	67	165	SEM	L	Q
FA intake	370	443	547			
Flow to duodenum, gm/day						
TFA	400	478	595	28.9	0.04	0.99
C14:0	16.5	16.6	16.2	0.12	0.31	0.35
C16:0	124	143	145	12.3	0.39	0.55
C18:0	192	192	225	9.96	0.13	0.39
C18:1	49	103	180	5.08	<0.01	0.80
C18:2	18.5	23.3	29.3	2.31	0.08	0.90
Saturated FAs	333	352	387	22.3	0.22	0.94
Unsaturated FAs	68	126	209	6.30	<0.01	0.80
Unsaturated: saturated FA ratio	0.20	0.36	0.54	0.03	<0.01	0.03
Fecal excretion, gm/day						
TFA	108	88	173	19.7	0.12	0.19
C14:0	1.42	3.46	2.52	0.89	0.55	0.28
C16:0	30.0	26.9	41.8	15.2	0.61	0.72
C18:0	70.3	43.9	112.6	4.0	0.01	<0.01
C18:1	4.87	10.9	14.4	4.45	0.28	0.72
C18:2	1.35	1.89	1.95	0.43	0.45	0.63
Saturated FAs	102	74	157	15.64	0.10	0.12
Unsaturated FAs	6.23	12.8	16.3	4.66	0.27	0.70
Unsaturated: saturated FA ratio	0.06	0.17	0.10	0.07	0.14	0.27
Posruminal digestion, %						
TFA	73.02	81.58	70.92	2.24	0.52	0.07
C14:0	91.40	78.76	84.52	5.91	0.56	0.30
C16:0	75.63	82.25	73.25	9.36	0.82	0.58
C18:0	63.22	77.05	49.51	2.71	0.06	0.03
C18:1	89.95	89.97	92.21	3.50	0.68	0.86
C18:2	92.64	91.21	93.11	1.61	0.80	0.50
Saturated FAs	69.31	78.96	59.81	2.33	0.08	0.04
Unsaturated FAs	90.83	89.84	92.30	3.15	0.72	0.77

intestine absorption of unsaturated FAs is greater than the saturated FAs [7]. Thereby, in marked difference with saturated, the intestinal digestion of unsaturated FA is hardly affected when their supply to the small intestine increases [21]. Accordingly, intestinal FA digestion and hence, the feeding value of fat is a function of FA intake level and the expected profile of FAs entering the small intestine.

With regard to finishing diets commonly offered for feedlot cattle in the USA, Plascencia et al. [6] informed that accurately known TFAs intake and intestinal digestion of FAs can be predicted as follows: FA digestion (gm/kg LW): Y = 87.560 - 8.591X ($R^2 = 0.89$, n = 25), where Y = intestinal digestion and X = FA intake per day (gm/kg LW). Due to

observed limitations of FA intake on subsequent FA digestion, the authors concluded that finishing diets should be formulated not to exceed 0.88 g FA/kg LW (approximately 0.98 gm lipid/kg LW). According to their equation, the estimated postruminal FA digestion rates are 72.3%, 69.3%, and 65.8% for infusions of 0, 67, and 165 gm/day, respectively. The observed postruminal FA digestion for 0 infused FA treatment was in close agreement (101%) with the expectation. However, for the 67 and 165 gm/day FA infusion treatments, the observed postruminal FA digestion was 15.1% and 7.2% greater than expected, respectively.

A synergistic effect of small intestine absorption of saturated FAs in broilers when the proportion of unsaturated

to saturated FAs ratio that reaches the intestine is raised has been reported [8]. In a similar manner, reducing the ruminal biohydrogenation index of the unsaturated FAs by protecting fat of the ruminal biohydrogenation increases TFAs absorption from 80.3% to 87.8% [9]. It has been established that there is a direct relationship (1:1) between the increase in the proportion of unsaturated to saturated FAs that reach the intestine and the increase in C18:0 intestinal digestibility. Nevertheless, limitations on intestinal FA digestion are also constrained by limits in bile production. Plascencia et al. [5] observed that bile production in cattle is relatively constant, and unlike non-ruminants, refractory to the level of fat intake, explaining 69% of the variation in postruminal FA digestion. In such a way that even when high proportions of unsaturated FAs reach the intestine, the potential for bile production will be the limiting factor for its absorption.

Based on the physiological fuel value of lipids (9 kcal metabolizable energy per gram) and the efficiency with which this energy can be used for weight gain [7], the net energy value for gain (NE $_{\rm g}$) of dietary fat is 6.03 kcal/gm of intestinally absorbable fat. Therefore, NE $_{\rm m}$ and NE $_{\rm g}$ values for supplemental tallow were 5.52 and 4.40, 6.16 and 4.95, and 5.40 and 4.33 for 0, 67, and 165 gm/day infusions, respectively. These findings demonstrate that increasing the fraction of unsaturated FA reaching the small intestine improves FA digestion in ruminants and non-ruminants alike. The extent of this increase in FA usage on growth performance responses, on the other hand, is uncertain. Additional research is needed to determine the effects of this method on feedlot cattle performance.

Conclusion

Decreasing saturated to unsaturated ratio of FAs entering the small intestine will enhance intestinal C18:0 digestion. Accordingly, protecting supplemental fat from ruminal biohydrogenation is an alternative to improve intestinal FA digestion and hence, the energy value of dietary fat in finishing diets for feedlot cattle. However, TFA supply to the small intestine is a major constraint on intestinal FA digestion (by limited bile production). Modifications to the unsaturated to saturated ratio of FAs entering the small intestine is expected to be more likely beneficial when expected FA flow to the intestine does not exceed 2.13 gm FA/kg LW.

List of Abbreviations

 $\rm Cr_2O_3$: chromic oxide, DM: dry matter, gm: gram, kg: kilogram, LW: live weight, NE_g: net energy of gain, NE_m: net energy of maintenance, NLT: not less than, ORPOL: orthogonal polynomials, SAS: Statistical Analytical System, TFA: total fatty acids.

Conflict of interest

No potential conflict of interest was reported by the authors.

Authors' contributions

AP: data curation, data interpretation, wrote the final version of the manuscript; AB: collaborated with statistical analyses; YSVG: contributed to writing and original draft preparation; RAZ: designed and supervised the experiment and laboratory analyses, revised and edited the final version of the manuscript.

References

- NASEM. Nutrient requirements of beef cattle. 8th Rev. edition, National Academies of Sciences, Engineering, and Medicine, National Academy Press, Washington, DC, 2016.
- [2] Varona E, Tres A, Vichi S, Barroeta AC, Guardiola F. Composition and nutritional value of acid oils and fatty acid distillates used in animal feeding. Animals (Basel) 2021; 11:196; https://doi. org/10.3390/ani11010196
- [3] Zinn RA, Plascencia A. Influence of level and method of supplementation on the utilization of supplemental fat by feedlot steers. J Anim Vet Adv 2004; 3(7):470-4.
- [4] Bionaz M, Vargas-Bello-Pérez E, Busato S. Advances in fatty acids nutrition in dairy cows: from gut to cells and effects on performance. J Anim Sci Biotechnol 2020; 11:110; https://doi. org/10.1186/s40104-020-00512-8
- [5] Plascencia A, Mendoza GD, Vazquez C, Zinn RA. Influence of levels of fat supplementation on bile flow and fatty acid digestion in cattle. J Anim Vet Adv 2004; 3(11):763–8.
- [6] Plascencia A, Mendoza GD, Vazquez C, Zinn RA. Relationship between body weight and level of fat supplementation on fatty acid digestion in feedlot cattle. J Anim Sci 2003; 81(11):2653–9; https://doi.org/10.2527/2003.81112653x
- [7] Zinn RA, Jorquera AP. Feed value of supplemental fats used in feedlot cattle. In: Hollis LC, Olson KC (eds.). Contribution to a book of veterinary food animal practice, Elsevier, Mosby Saunders, Philadelphia, PA, pp 247–68, 2007.
- [8] Rodríguez-Sánchez R, Tres A, Sala R, Garcés-Narro C, Guardiola F, Gasa J, et al. Effects of dietary free fatty-acid content and saturation degree on lipid-class composition and fatty-acid digestibility along the gastrointestinal tract in broiler starter chickens. Poult Sci 2019; 98:4929-41; https://doi.org/10.3382/ps/pez253
- [9] Zinn RA, Gulati SK, Plascencia A, Salinas J. Influence of ruminal biohydrogenation on the feeding value of fat in finishing diets for feedlot cattle. J Anim Sci 2000; 78(7):1738–46; https://doi. org/10.2527/2000.7871738x
- [10] Nichols K, Dijkstra J, van Laar H, Pacheco S, van Valenberg HJ, Bannink A. Energy and nitrogen partitioning in dairy cows at low or high metabolizable protein levels is affected differently by postrumen glucogenic and lipogenic substrates. J Dairy Sci 2019; 102(1):395–412; https://doi.org/10.3168/jds.2018-15249
- [11] Núñez-Benítez VH, Barreras A, Estrada-Angulo A, Castro-Pérez BI, Urías-Estrada JD, Zinn RA, et al. Evaluation of a standardized mixture of synbiotic-glyconutrients as a feed additive in steers fed a finishing diet: site and extent of digestion, ruminal fermentation, and microbial protein synthesis. Livest Sci 2021; 243:104373; https://doi.org/10.1016/j.livsci.2020.104373
- [12] Hill FN, Anderson DL. Comparison of metabolizable energy and productive determinations with growing chicks. J Nutr 1958; 64(4):587-603; https://doi.org/10.1093/jn/64.4.587

- [13] Sukhija P, Palmquist DL. Rapid method for determination of total fatty acid content and composition of feedstuff and feces. J Agric Food Chem 2008; 36(6):1202-6; https://doi.org/10.1021/ jf00084a019
- [14] Zinn RA, Plascencia A. interaction of whole cottonseed and supplemental fat on digestive function in cattle. J Anim Sci 1993; 71(1):11–7; https://doi.org/10.2527/1993.71111x
- [15] Statistical Analysis Software (SAS). Version 90 SAS Institute Inc. Cary, NC, 2004.
- [16] Zinn RA, Plascencia A. Effect of forage level on the comparative feeding value of supplemental fat in growing-finishing diets. J Anim Sci 1996; 74(6):1194–201; https://doi.org/10.2527/1996.7461194x
- [17] Plascencia A, Estrada M, Zinn RA. Influence of free fatty acid content on the feeding value of yellow grease in finishing diets for feedlot cattle. J Anim Sci 1999; 77(10):2603-9; https://doi. org/10.2527/1999.77102603x
- [18] Plascencia A, Zinn RA. Comparative effects of "solid" fat sources as a substitute for yellow grease on characteristics of digestion of growing diets for feedlot cattle. Anim Prod Sci 2019; 59(8):1520–7; https://doi.org/10.1071/AN16820
- [19] Plascencia A, Zinn RA. Influence of method supplementation of yellow grease on growth performance, dietary energetics, carcass characteristics and nutrient digestion of feedlot steers. Iran J Appl Anim Res 2021; 11(1)39–46.
- [20] Lourenco M, Ramos-Morales E, Wallace RJ. The role of microbes in rumen lipolysis and biohydrogenation and their manipulation. Animal 2010; 4(7):1008–23; https://doi.org/10.1017/ S175173111000042X
- [21] Boerman JP, Firkins JL, Dt-Pierre NR, Lock AL. Intestinal digestibility of long-chain fatty acids in lactating cows: a meta-analysis and meta-regression. J Anim Sci 2015; 98(12):8889–903; http:// dx.doi.org/10.3168/jds.2015-9592