

MULTIDRUG RESISTANT ACINETOBACTER SPP. BLOOD STREAM INFECTION IN A NEONATAL INTENSIVE CARE UNIT OF AN URBAN SPECIALIZED HOSPITAL IN DHAKA

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Abstract

Background: Acinetobacter spp. are ubiquitous in the environment, that is, soil and water, and occasionally isolated from mucous membrane, secretion, and skin of hospitalized patients, also on surfaces of hospital environment. Antibiotic resistant Acinetobacter spp. blood stream infection is a leading problem. Multidrug resistant Acinetobacter spp. blood infection in the neonatal intensive care unit (NICU) patients create a great problem in hospital settings.

Methods: This study was carried out from January to December 2011 based on the clinical specimens obtained from suspected cases of septicemia patients admitted in the NICU of Ad-din Women's Medical College Hospital, Dhaka, Bangladesh. Blood collected from suspected cases by maintaining strict aseptic precaution and sent immediately to microbiology laboratory of the same institution. Samples were also taken from different sites in NICU and Department of Obstetrics & Gynaecology operation theatre for detection of the sources of infection in outbreak situation. Isolated organisms were identified by standard bacteriological method and drug susceptibility test had been done by modified Kirby Bauer disk diffusion technique.

Results: A total 87 blood culture positive cases were included in this study. Out of 87 culture positive cases Acinetobacter spp. was the predominant pathogen 32 (35.58%). Acinetobacter spp. isolated from the blood of newborn and the mortality rate was 3.33%. Acinetobacter blood stream infection is more common in the first 7 days of life (early onset), birth weight less than 2 kg (56.6%), also affected the baby with birth weight more than 2.5 Kg (23.3%) and most of the neonates having gestational age less than 37 weeks (70%). Twenty four babies (80%) were delivered in the same hospital. Acinetobacter spp. showed increased resistance against majority of antibiotics such as colistin 3.12%, levofloxacin 40.62%, imipenem 53.12%, cotrimoxazole 54.54%, piperacillin tazobactam 59.37%, amikacin 75%, ciprofloxacin 78.12%, gentamycin 84.37%, amoxyclavulonic acid 93.75%, ceftriaxone, ceftazidime and cefixime 96.87% cefuroxime, cephadrine and ampicillin 100% respectively. We categorized the Acinetobacter spp. in to 3 categories, such as multidrug resistant (MDR) 37.5%, extensive drug resistant (XDR) 28.12% and pandrug resistant (PDR) 3.12% Miscellaneous pattern of sensitivity 31.15%. A cluster of cases occurred in month June, which was quite higher than other months, Surveillance had been done. Acinetobacter spp. isolated from the environment of operation theatre of Obstetrics and Gynaecology department. In NICU, Acinetobacter spp. were isolated from suction water, suction tube, incubator door handle and body surface of the neonates. Acinetobacter spp. which were isolated from suction tube and suction water were only sensitive to colistin and resistant to other drugs. Two Acinetobacter spp. were also isolated from OT environment and body surface of neonates which were found sensitive to only imipenem and colistin. Acinetobacter spp. isolated from incubator door handle was sensitive to colistin, imipenem, amikacin, gentamycin, levofloxacin, ciprofloxacin, piperacillin tazobactam and cotrimoxazole.

Key words: Multidrug resistance, Acinetobacter, septicemia, antibiotic resistance, nosocomial infection.

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Introduction

Acinetobacter baumannii is an opportunistic pathogen that is frequently involved in out breaks of infection occurring mostly in intensive care units from contaminated common sources, e.g. ventilator equipments, catheters, etc¹. *Acinetobacter spp. areaerobic* Gram negative, non-motile, non-spore forming encapsulated or biofilm producing coccobacilli belonging to family Neisseriaceae². Commonly present in environment (water, soil) and hospital (catheters, lotions, ventilator equipments). *Acinetobacter baumannii* is the major species of *Acinetobacter*. It is an important pathogen found to be associated with wide spectrum of nosocomial infection. Risk factors for *Acinetobacter* infection are hospitalization, ICU, surgery, antibiotic exposure and catheters³. It is clearly pathogenic when recovered from blood and normally sterile body sites³. Septicemia is a major cause of morbidity and mortality in newborns, especially in developing countries. There are many studies documented worldwide in the literatures emphasizing the *Acinetobacter spp.* as an important nosocomial agent of septicemia in neonatal intensive care units⁴⁻⁶. Management of multidrug resistant *Acinetobacter spp.* infections is a great challenge for physicians and clinical microbiologist. *Acinetobacter spp.* has the ability to survive in a hospital milieu and its ability to persist for extended periods of time on surfaces makes it a frequent cause for health care associated infection and it leads to multiple outbreaks². Early diagnosis and appropriate antimicrobial therapy of septicemia are of utmost importance to prevent morbidity and mortality.

Definitions of multidrug-resistant *Acinetobacter* species vary when referring to a wide array of genotypes and phenotypes⁷. Different terms like 'multidrug resistant (MDR)', 'extensive drug resistant (XDR)', and 'pandrug resistant (PDR)' have been used with varied definitions to describe the extent of antimicrobial resistance among *Acinetobacter spp.* However, to date, unlike *Mycobacterium tuberculosis*, internationally, there are no accepted definitions for the extent of resistance in the bacteria. Arbitrarily used terms have thus

caused great confusion making it difficult for the available literature to be analyzed⁸.

In the present study 'MDR *Acinetobacter spp.*' would be defined as the isolate resistant to at least three classes of antimicrobial agents such as all penicillins and cephalosporins (including inhibitor combinations), fluoroquinolones, and aminoglycosides. 'XDR *Acinetobacter spp.*' would be the *Acinetobacter spp.* isolate that is resistant to the three classes of antimicrobials described above (MDR) and shall also be resistant to carbapenems. Finally, 'PDR *Acinetobacter spp.*' would be the XDR *Acinetobacter spp.* that is resistant to polymyxins and tigecycline. These definitions further help to clearly define the extent of resistance and rational antimicrobial therapy⁹.

The current study highlighted *Acinetobacter spp.* as important pathogens in neonatal blood stream infection, relationship of *Acinetobacter* infection with onset of disease, gestational age, birth weight, inborn or out born delivery, identification of the sources of infection in outbreak situation and evaluation of its antimicrobial sensitivity.

Methods

The study was done in Ad-din Women's Medical College and Hospital, Dhaka, Bangladesh, from January to December of 2011. Blood collected from suspected cases by maintaining strict aseptic precaution in to blood culture bottle and sent immediately to microbiology laboratory of the same hospital. Only symptomatic babies who have had blood culture positive for *Acinetobacter spp.* were included in the study. Blood culture test was done by FAN method. Subcultures of the positive samples were done on blood agar and McConkey's agar media. Organisms were identified by standard bacteriological method and drug susceptibility test was done by modified Kirby Bauer disk diffusion technique¹⁰⁻¹². The surveillance study to detect the sources of infection was carried out. Samples of NICU from various articles were taken for cultures. Swabs were taken from suction tubes, water for suction, incubator door handle, baby cots, body surface of the newborn and hand swabs of nurses working in NICU. Samples from labor room and Obstetrics

& Gynaecology operation theatre were also taken. wabs were taken from baby trays, resuscitation equipments, suction apparatus. Air sampling was done by keeping two open plates of blood agar and McConkey's agar in to the operation rooms and NICU for 30 minutes^{13,14}. All the samples and plates were immediately sent to microbiology laboratory and samples were immediately inoculated in to blood agar and MacConkey's agar media and plates were incubated in to incubator at 37°C. Isolated organisms were identified by standard biochemical test. Identification of *Acinetobacter spp.* was made as per standard criteria¹¹. Susceptibility pattern were done by modified Kirby Bauer disc diffusion technique¹⁰.

Results

Table-I shows that organisms isolated from the blood of the newborn are *Acinetobacter spp.* 32(34.48%), followed by *Pseudomonas spp.* 19(21.83%), CoNS 16(18.39%), *Klebsiella* and *Moraxella spp.*6(6.89%) respectively, *Staph.*

aureus 4(5.97%) *Enterobacter spp.*3(3.44%), *Esch. coli* (2.29%) and *Enterococci spp.* 1(1.49%).The other results are shown in the tables II-V.

Table-I

Different organisms isolated from blood of neonates in NICU

Name of organisms	Number	Percentage (%)
<i>Acinetobacter spp.</i>	32	34.48
<i>Pseudomonas spp.</i>	19	21.83
CoNS	16	18.39
<i>Klebsiella spp.</i>	06	6.89
<i>Moraxella spp.</i>	06	6.89
<i>Staph. aureus</i>	04	5.97
<i>Enterobacter spp.</i>	03	3.44
<i>Esch.coli</i>	02	2.29
<i>Enterococci spp.</i>	01	1.49
Total	87	

Table-II

Relationship of Acinetobacter spp. blood infection with onset of disease mortality rate, birth weight, place of delivery and gestational age (n=30)

Mortality	Age of onset		Birth weight (kg)			Gestational age (week)		Place of delivery		Total
	Early	Late	1- 1.9 kg	2- 2.5 kg	>2.5 kg	< 37 wks	>37 wks	In-born	Out- born	
1	21	09	17	06	07	21	9	24	6	30
(3.33%)	(70%)	(30%)	(56.6%)	(20%)	(23.3%)	(70%)	(30%)	(80%)	(20%)	

Table-III

Susceptibility pattern of Acinetobacter spp. to different antimicrobial agents (n=32)

Antibiotics	Sensitive		Intermediate sensitive		Moderate	
	No.	%	No.	%	No.	%
Colistin	31	96.87			01	3.12
Imepenem	15	46.87			17	53.12
Piperacillin-tazobactam	13	40.62			19	59.37
Amikacin	08	25			24	75
Levofloxacin	08	25	11	34.37	13	40.62
Ciprofloxacin	06	18.75	01	3.12	25	78.12
Gentamycin	05	15.62			27	84.37
Ceftriaxone	01	3.12			31	96.87
Ceftazidime	01	3.12			31	96.87
Cefuroxime	01	3.12			31	96.87
Cephradin	00	00			32	100
Ampicillin	00	00			32	100
Amoxyclavulonic acid	02	6.25			30	93.75
Cotrimoxazole	15	46.87			17	53.12

Table-IV
Rate of MDR, XDR and PDR *Acinetobacter* spp. (n=32)

Categories of <i>Acinetobacter</i>		Definition	No.	Percentage
MDR	Resistant to 1. All cephalosporins and inhibitor combination 2. Fluroquinolones, 3. Aminoglycosides.	12	37.5%	
XDR	MDR+ Resistant to carbapenems	09	28.12%	
PDR	XDR + Resistant to polymixin E(colistin)	01	3.12%	
Unclassified	Miscellaneous pattern of sensitivity	10	31.25%	

Table-V
Acinetobacter spp. isolated from different article in NICU, Obstetrics & Gynaecology OT (OBG-OT)

Site	Susceptibility pattern
1. Incubator door handle (NICU) R- CTR / CAZ/ CXM /AMC/	1. S- GEN/ IMP/ CIP/ LEV/ PT/ AK/ CL
2. Suction tube (NICU)	2. S-CL IS- LEV
3. Suction water (NICU)	3. S-CL IS-LEV
4. Body surface of baby	4. S- CL / IMP
5. OT environment(OBG-OT)	5. S-CL / IMP

Note: S(Sensitive), IS- Intermediate Sensitive, NICU(Neonatal ICU), GEN (gentamycin), IMP(imipenem), CIP (ciprofloxacin), LEV (levofloxacin), PT(piperacillin-tazobactum), AK (amikacin), CL (colistin).

Discussion

Acinetobacter spp. has emerged as an important pathogen causing life threatening infections both in community and hospital. Rapid emergence of multidrug-resistant *Acinetobacter* spp. has further made the situation critical². Multidrug resistant nosocomial *Acinetobacter* spp. septicemia may cause severe clinical diseases in neonate that is associated with a high mortality¹⁵. From January to December 2011 eighty seven organisms were isolated from the blood of neonates. Out of 87 blood culture positive cases *Acinetobacter* spp. was the most prevalent organism in NICU patients (34.48%). Isolation rate of *Acinetobacter* spp. from blood samples of neonates in various study varies from 8.3% to 15.2%^{6,16}.

NICU of Ad-din women's Medical Collage Hospital, Dhaka, is a referral centre for babies born outside of this hospital. In the present study, we found that *Acinetobacter* spp.

septicemia was more common for the babies in the first 7 days of life (early onset) having birth weight less than 2kg, baby took born before 37weeks of pregnancy. Among twenty four neonates having *Acinetobacter* septicemia, 20 of them delivered in the same hospital by the method of LUCS and four by NVD. *Acinetobacter* also affected the average gestational age (30%) and average birth weight babies (23.3%). *Acinetobacter* spp. blood infection can even occur in the babies without being under intensive care or antibiotics, mechanical ventilation or having indwelling catheters¹⁷. Institutional birth, preterm birth, and low birth weight were identified the most frequent risk factors^{6,16,17}. This might be because of prevalence of multi-drug resistant strains in the hospital environment⁶. Mortality rate is lowered (3.33%) in our study compare with others¹⁸. It might be due to involvement of babies more than higher birth weight and without prior severe illness.

Acinetobacter spp. has been implicated in many outbreaks in the neonatal sepsis in NICU¹³. From January to May of 2011, 15 *Acinetobacter* spp. isolated from blood of suspected cases of septicemia patients in Neonatal intensive care (NICU) in Ad-din women's medical college hospital, but in the month of June 2011, 12 *Acinetobacter* spp. were isolated from blood of neonates, rate was surprisingly more than other months, so we took it as an outbreak, and nine of them are XDR (MDR + resistance to carbapenam).

Surveillance study was done and samples were taken from several sites in NICU and operation theatre of OBG department. *Acinetobacter* spp. was isolated from incubator door handle, suction tube, suction water and body surface of the baby in NICU and it was also isolated from OT environment during the outbreak time. Organisms isolated from suction water and suction tube are XDR, these susceptibility pattern correlates with the susceptibility pattern of organisms isolated from the patients at the time of outbreak. This time some of the sucker machines in NICU were unserviceable and a single sucker machine being used for several babies. Enforcement of hospital infection prevention control measure had been done, in the following six months only five *Acinetobacter* sp. were isolated from the blood of newborn.

In the current study isolated *Acinetobacter* spp. showed high level of resistance to most of the antibiotic compared with other studies done in different places^{16,17}. In present study more than 96% of isolates were resistant to 3rd generation cephalosporin, followed by gentamycin 84.37%, ciproflaxacin 78.12%, amikacin 75%, piperacillin-tazobactam 59.37%, imipenem and cotrimoxazole 53.12%, levofloxacin 40.62% and colistin 3.12%. Ampicillin and 1st and 2nd generation cephalosporin found 100% resistant. In a surveillance study of the antibiotic susceptibility pattern of the ICUs of five European countries had done and the prevalence of resistance of *Acinetobacter* spp. to gentamycin was 0-81%, amikacin 10-51%, ciprofloxacin 19-81%, ceftazidime 0-81%, piperacillin-tazobactam 36-75% and imipenem 5-19%¹⁹. In present study

38% of *Acinetobacter* spp. were MDR, 28.12% XDR, 3.12% PDR and miscellaneous pattern of sensitivity 31.25%. *A. baumannii* was isolated most in a study were multidrug resistant (MDR)²⁰. According to Abbo et al., 2005, MDR criteria were defined when resistant to all of studied antibiotics, that is, piperacillin-tazobactam, cefepime, ceftazidime, aztreonam, ciprofloxacin, gentamicin, and tobramycin, but could be sensitive to amikacin, ampicillin-sulbactam, imipenem, meropenem, and minocycline²¹. Several reports showed that the usage of broad spectrum antibiotics affected normal flora and induced MDR *A. baumannii*^{22,23}. *Acinetobacter* spp. are rapidly spreading with emergence of extended resistance to even newer antimicrobials. They have the ability to acquire resistance at a much faster than other gram-negative organism⁹. Due to their ease of survival in the hospital environment, they have immense potential to cause nosocomial outbreaks. In addition to antibiotic resistance, their biofilm forming ability plays a crucial role in their *in-vitro* and *in-vivo* survival⁹.

Conclusion

In the present study, we concluded that *Acinetobacter* spp. is one of the major pathogens in NICU, can reach in outbreak proportions and may involve babies with normal birth weight. Most of the *Acinetobacter* spp. are multidrug resistant. Colistin is the drug of choice for blood stream infection of ICU patient caused by *Acinetobacter* spp. To decrease the spread of *Acinetobacter* infections and reduce the pace of emergence of resistance in MDR *Acinetobacter*, it is important to promote the rational use of antimicrobials, with implementation and monitoring of the Antibiotics Stewardship Program in hospitals. Hospital infection prevention committee can play a key role to prevent nosocomial infection.

References

1. Paterson DL. The epidemiological profile of infections with multidrug-resistant *Pseudomonas aeruginosa* and *Acinetobacter* species. Clin Infect Dis 2006; 43: S43-8.
2. Bergongne-Berezin E, Towner JK. *Acinetobacter* spp. as nosocomial pathogens: Microbiological, clinical

- and epidemiological features. Clin Microbiol Rev 1996; 9: 148-51.
3. Bartlett, JG. *Acinetobacter baumannii*: Johns Hopkins Medicine POC-IT Guides. Last updated: November 15, 2011 http://www.hopkinsguide.com/hopkins/ud/view/Johns_Hopkins_ABX_Guide/54003/all/A.
 4. Touati A, Achour W, Cherif A, et al. Out break of *Acinetobacter baumannii* in neonatal intensive care unit: antimicrobial susceptibility and genotyping analysis. Ann Epidemiol 2009; 19:372-8.
 5. Jarousha AA, El Qouqa IA, Jadba AE, Al Afifi AS. *Acinetobacter baumannii* infection in the neonatal intensive care unit. Iranian J Health 2008; 37:107-12.
 6. Arora U, Jaitwani J. *Acinetobacter* spp.: an emerging pathogen in neonatal septicaemia in Amritsar. Indian J Med Microbiol 2006;24:81.
 7. Falagas ME, Koletsi PK, Bliziotis IA. The diversity of definitions of multidrug-resistant (MDR) and pandrug-resistant (PDR) *Acinetobacter baumannii* and *Pseudomonas aeruginosa*. J Med Microbiol 2006;55: 1619-29.
 8. Falagas ME, Karageorgopoulos DE. Pandrug resistance (PDR), extensive drug resistance (XDR), and multidrug resistance (MDR) among Gram-negative bacilli: need for international harmonization in terminology. Clin Infect Dis 2008;46: 1121-2.
 9. Manchanda V, Sanchaita S, Singh NP. Multidrug resistant *Acinetobacter*. J Global Infect Dis 2010; 2(3): 291-304.
 10. Kirby AW, Sherris JC, Truck M. Antibiotic susceptibility testing by a standardized single disk method. Am J Clin Pathol 1996; 36(3): 493-6.
 11. Collee JG. Laboratory strategy in the diagnosis of infective syndrome. In: Duguid JP, Fraser AG, Marimon BP. eds. Mackie and McCartney Practical Medical Microbiology. 14th ed. New York: Churchill Livingstone; 1996.
 12. Chessbrough M. Medical laboratory manual for tropical countries. Vol. II. Cambridge: ELBS; 1984.
 13. Von Dolinger de Brito D, Oliveira EJ, Abdullah VO, et al. An outbreak of *Acinetobacter baumannii* septicemia in a neonatal intensive care unit of a university hospital in Brazil. Brazil J Infect Dis 2005; 9:301-9.
 14. Mohiuddin M, Haq A, Haq M, Huq F. Microbiology of nosocomial infection in tertiary hospital of Dhaka city and its impact. Bangladesh J Med Microbiol 2010; 4: 32-7.
 15. Cisneros JM, Reyes MJ, Pachon J, et al. Bacteremia due to *Acinetobacter baumannii*: epidemiology, clinical findings and prognostic features. Clin Infect Dis 1996;22:1026-32.
 16. Shete VB, Ghadage DP, Muley VA, Bhore AV. *Acinetobacter* septicemia in neonate admitted to intensive care unit. J Lab Phys 2009;1: 73-6.
 17. Ashit M, Shudhir M, Geetha J, et al. *Acinetobacter* Sepsis in Newborns. Indian Pediatr 1998;35:27-32.
 18. Seifert H, Strate A, Pulverer G. Nosocomial bacteremia due to *Acinetobacter baumannii*: clinical features, epidemiology and predictors of mortality. Medicine 1995; 74: 340-9.
 19. Hanberger H, Garcia-Rodriguez JA, Gobernado M, et al. Antibiotic susceptibility among aerobic gram-negative bacilli in intensive care units in 5 European countries. French and Portuguese ICU Study Groups. JAMA 1999;281: 67-71.
 20. Tjoa E, Moehario LH, Rukmana A, Rohsiswarno R. *Acinetobacter baumannii*: role in blood stream infection in neonatal unit, Dr. Cipto Mangunkusumo Hospital, Jakarta, Indonesia. Int J Microbiol 2013;
 21. A Abbo S, Navon-Venezia O, Hammer-Muntz T, et al. Multidrug-resistant *Acinetobacter baumannii*. Emerg Infect Dis 2005; 11(1): 22-9.
 22. Agustí CM, Pujol MJ, Argerich MJ, et al. Short-term effect of the application of selective decontamination of the digestive tract on different body site reservoir ICU patients colonized by multi-resistant *Acinetobacter baumannii*. J Antimicrob Chemother 2002; 49: 205-8.
 23. Camp, Tatum OL. A review of *Acinetobacter baumannii* as a highly successful pathogen in times of war. Lab Med 2010; 41: 649-57.