SCREENING OF *GLYCOSMIS PENTAPHYLLA* FOR CYTOTOXIC AND REPELLENT POTENTIALS AGAINST *ARTEMIA SALINA* NAUPLII AND *TRIBOLIUM CASTANEUM* (HERBST) ADULTS

M. S. Pramanik, M. Y. Akter, A. E. Ekram¹, H. Islam, A. R. Khan and N. Islam*

Department of Zoology, University of Rajshahi, Rajshahi 6205, Bangladesh; ¹Department of Genetic Engineering and Biotechnology, University of Rajshahi, Rajshahi 6205, Bangladesh

Abstract: Chloroform extracts of the leaf, stem bark, stem wood and roots of *Glycosmis pentaphylla* (Retz.) were tested against the brine shrimp, *Artemia salina* nauplii for cytotoxic activity in which the dose-mortality assay revealed LC_{50} values of 28.579, 28.659, 57.213 and 84.111 ppm respectively, for the plant parts efficacy of which could be arranged in the order of leaf > stem bark > stem wood > root. The same extracts were tested for repellency against *Tribolium castaneum* (Herbst) adults where the root bark and root wood were separately extracted and used. The F values of the arcsin transformed data were 60.983, 14.177, 19.437, 15.429 and 1.082 respectively for the leaf, stem bark, stem wood, root bark and root wood extracts. Except for the root wood extract, strong repellent activity was observed for the rest of the extracts (P<0.001).

Key words: Artemia salina, Tribolium castaneum, Glycosmis pentaphylla, cytotoxic activity, repellent activity.

miusk: eBb ukt=ui bluz-i (Artemia salina nauplii) Ici Glycosmis pentaphylla (Aultikuivev ful) NuQi cuZy, Kuži euKj, Kuži KW I tkKtoi tKutivdg@bh@n ctqutW tWR-gUğiW cix[]Y c×uZtZ mBtUUW K cuZupav ch&e[]Y Kivnq huZ LC₅₀ cul qv huq h_utg 28.579, 28.659, 57.213 I 84.111 ususGg; Ges KuhRuiZui wK witq gubi ubgpgubynti mRutj djudj cul qv huh cuZu> Kuži euKj> Kuži KW> tkKo | H GKB ubm@ yj Aului tcuKv Tribolium castaneum (Herbst)-Gi Dci wZvob KuhRuiZul ch&e[]Y Kivnq (Gtq]tî tkKtoi euKj I tkKtoi KW Ayiv v Ayiv ufute cix[]v Kivnq) | AK@b ifcušuiZ Dcuži Analysis of variance Kti F-gub cul qv huq cuZy, Kuži euKj, Kuži KW, tkKtoi euKj I tkKtoi KuVi Rb" h_utg 60.983, 14.177, 19.437, 15.429 I 1.082 | tkKtoi KW e ZuZ meK@U ubh@mi wZvob guÎv ulj D'P Zvch@V@(P <0.001)]

Introduction

Glycosmis pentaphylla (Retz.) is a wild, oriental and an unarmed shrub or small tree distributed throughout the roadsides, under large trees or on uncultivated lands in Bangladesh, Sri Lanka, Eastern part of India, Southern Indochina, Malaysia, Indonesia and in Eastern Australia. In Bangladesh and India it is popularly known as Ash-sheura, Bon-nimbu, Bon-nebu, Atishoti, Vat and Dantan-gaas (Rastogi et al., 1980). In Ayurvedic medicine the plant has been used traditionally in bilious complaints, cough, worms, jaundice and fever (Anon, 1952). Its medicinal virtues came into light when it was found that people who were cleansing their teeth with little sticks of G. pentaphylla do not suffer from Kala-azar (Kirtikar and Basu, 1935). Its growth-inhibiting activity against larvae of Diaprepes abbreviatus is also known (Shapiro et al., 1997). Different species of Glycosmis contain a wide variety of compounds with potential biological activity (Stone, 1985). These include terpenoids (Chakravarty et al., 1996), amides (Greger et al., 1996; Hofer et al., 1995a, 1998), imides (Hofer et al., 1995b), alkaloids (Wu et al., 1983; Wurz et al., 1993; Ono et al., 1995), coumarins (Rahmani et al., 1998), and flavonoids (Wu et al., 1983). Compounds exhibiting antifungal and insecticidal activities have already been isolated from

*Corresponding author, email: n_islamm@yahoo.com

several *Glycosmis* species (Greger *et al.*, 1996). However, previous workers investigated this plant giving emphasis mostly on the chemical constituents and their medicinal profile but information on its various biological activities is still scanty. Since this plant is being used as a toothbrush, bioactive potentials other than killing of the test insects were taken into consideration and accordingly, cytotoxic and repellent activity tests were carried out on the brine shrimp, *Artemia salina* nauplii and the red flour beetle, *Tribolium castaneum* (Herbst).

Materials and Methods

Brine shrimp lethality assay: It is a bench top bioassay method recently developed in the arena of bioassay for the bioactive compounds, which indicates cytotoxicity, as well as, a wide range of pharmacological activities, *e.g.* anticancer, antiviral, pesticidal, AIDS, etc. of the compounds. Brine shrimp eggs were purchased and kept in aerated seawater (3.8% sodium chloride or 38gm salt/1000ml) of water at room (25-30 °C) temperature. It normally takes 30 – 48 h to give nauplii under these conditions. For the leaf and stem bark, samples (2 mg) were initially dissolved in 100 µl of dimethyl sulfoxide (DMSO) to make it hydrophilic before adding 19.98 ml of water and diluted up to 20 ml with brine water to obtain a series of concentration, viz. 100, 50, 25 and 12.5 ppm. For the stem wood and root samples 5 mg of each were initially dissolved in 200 μ l of DMSO before adding 19.98 ml of water and diluted up to 20 ml with brine water to obtain a series of concentration, e.g. 250, 125, 62.5, 31.25 and 15.625 ppm. After 48 h, 10 freshly hatched nauplii were added to each test tube with different concentration of the extracts of *G. pentaphylla* samples. After 24 h the number of nauplii killed in each tube was counted and compared to controls. LC₅₀ values were calculated by probit analysis.

Repellent activity test: The repellency test used in this investigation was adopted from the method (No. 3) of McDonald et al. (1970) with some modifications by Talukder and Howse (1993, 1994) and the test insect was T. castaneum adults. For the continuous supply of the adult beetles the test insect was mass-cultured. A standard mixture of whole wheat flour with powdered dry yeast (19:1) was used as food medium throughout the experimental period (Park and Frank, 1948). Both the flour and the powdered dry yeast were sterilized at 60° C for six hours in an oven. Food was not used until at least 15 days after sterilization to allow its moisture content to equilibrate with the environment (Khan and Selman, 1981). Half filter paper discs (Whatman No. 40, diameter 9 cm) were prepared and selected doses of all the CHCl₃ extracts were separately applied onto each of the half-discs and allowed to dry out as exposed to the air for 10 minutes. Each treated half-disc was then attached lengthwise, edge-to-edge, to a control half-disc with adhesive tape and placed in petri dishes. The orientation of the same was changed in the replica to avoid the effects of any external directional stimulus affecting the distribution of the test insects. Ten adult insects were released in the middle of each of the filterpaper circles. Each concentration was tested for five times. Insects that settled on each half of the filter paper discs were counted after 1 h and then at hourly intervals for 5 hours. The average of the counts was converted to percent repellency (*PR*) using the formula of Talukder and Howse (1993, 1995): PR = 2(C - 50), where, C is the percentage of insects on the untreated half of the disc. Positive values expressed repellency and negative values for attractant activity. The data obtained as percent repellency, was again developed by arcsin transformation for the calculation of analysis of variance (ANOVA).

Result and Discussion

For the leaf, stem bark, stem wood and root extracts the LC₅₀ values established were 28.579, 28.659, 57.213 and 84.111 ppm respectively (Table 1). The findings of Muthukrishnan et al. (1999) resembles corroborate with the results. The quinazolone of arborine isolated from the ethyl acetate fraction of G. pentaphylla leaf extract to water resulted in 83 to 100% mortality of Culex quinquefasciatus larvae. Except the root wood extract strong repellent activity was found. The repellency due to differences between doses were highly significant (P<0.001). The F values have been established through ANOVA with the arcsine transformed data being 60.983, 14.177, 19.437, 15.429 and 1.082 for leaf, stem bark, stem wood, root bark and root wood extracts respectively (Table 2 and 3). The P values were established as 2.03E-11, 5.41E-6, 4.62E-7, 2.48E-6 and 0.4002 for the analysis between doses for the leaf, stem bark, stem wood, root bark and root wood extracts respectively, and the degree of repellent activity could be arranged in a descending order: leaves> stem wood> root bark> stem bark. This finding has been supported by Chopra et al. (1956) who mentioned that the leaves of this plant are used to keep insects away from sweets and other edible items that are taken by natives in India and Australia.

Plant parts	LC ₅₀ value	95% confid	lence limits	Regression equations	χ^2 value (Df)	
	(ppm)	Upper	Lower	Regression equations		
Leaves	28.579	33.840	24.136	Y = -0.901 + 4.053 X	4.056 (2)	
Stem bark	28.659	41.419	19.831	Y = 1.639 + 2.306 X	1.402 (1)	
Stem wood	57.213	68.863	47.535	Y = -1.436 + 3.662 X	5.870 (2)	
Root	84.111	109.118	64.836	$Y = 1.007 \ + 2.074 \ X$	3.013 (3)	

Table 1. Dose mortality effect of G. pentaphylla extracts against A. salina nauplii after 24 hours of exposure

Type of extracts	Dose (µg/cm ²)	Average of hourly observations (Nc)				Percent repulsion (PR) PR = $(Nc - 5) \times 20\%$					
C T		1h	2h	3h	4h	5h	1h	2h	3h	4h	5h
Leaf	78.634	9.33	9.33	9.00	9.66	9.66	86.6	86.6	80.0	93.2	93.2
	39.317	9.00	9.66	7.33	7.33	7.00	80.0	93.2	46.6	46.6	40.0
	19.659	6.00	6.66	7.00	7.33	6.66	20.0	33.2	40.0	46.6	33.2
	9.829	8.00	7.00	7.00	6.33	6.00	60.0	40.0	40.0	26.6	20.0
	4.915	5.33	3.66	4.33	3.66	3.66	6.6	-26.8	-13.4	-26.8	-26.8
	2.457	4.66	3.66	4.00	3.00	1.33	-6.8	-26.8	-20.0	-40.0	-73.4
Stem bark	78.634	7.00	7.00	4.33	4.00	7.66	40.0	40.0	-13.4	-20.0	53.2
	39.317	8.33	8.00	4.33	7.66	6.33	66.6	60.0	-13.4	53.2	26.6
	19.659	7.00	7.00	6.66	5.66	6.66	40.0	40.0	33.2	13.2	33.2
	9.829	3.66	3.00	3.33	4.00	4.66	-26.8	-40.0	-33.4	-20.0	-6.8
	4.915	4.00	4.66	4.00	3.00	2.66	-20.0	-6.8	-20.0	-26.8	-46.8
	2.457	2.66	3.00	3.00	3.66	4.00	-46.8	-40.0	-40.0	-26.8	-20.0
Stem Wood	78.634	7.66	7.33	6.66	7.66	7.33	53.2	46.6	33.2	53.2	46.6
	39.317	5.33	3.66	4.33	3.66	3.66	6.6	-26.8	-13.4	-26.8	-26.8
≥	19.659	4.66	5.33	5.66	4.00	1.66	-6.8	6.6	13.2	-20.0	-66.8
Е	9.829	7.00	7.33	7.00	6.00	6.33	40.0	46.6	40.0	20.0	26.6
ite	4.915	3.66	3.33	4.00	3.33	4.33	-26.8	-33.4	-20.0	-33.4	-13.4
S	2.457	5.00	4.00	3.66	4.33	4.33	00.0	-20.0	-26.8	-13.4	-13.4
	78.634	10.0	10.0	10.0	10.0	10.0	100	100	100	100	100
Root bark	39.317	9.00	9.00	8.66	5.66	5.66	80.0	80.0	73.2	13.2	13.2
	19.659	9.00	8.33	7.00	7.33	7.00	80.0	66.6	40.0	46.6	40.0
	9.829	6.00	5.33	5.33	4.66	4.33	20.0	6.6	6.6	-6.8	-13.4
	4.915	7.00	7.33	6.00	4.66	4.33	40.0	46.6	20.0	-6.8	-13.4
	2.457	3.66	3.33	4.00	3.33	5.00	-26.8	-33.4	-20.0	-33.4	00
	78.634	8.33	9.66	9.33	10.00	9.66	66.6	93.2	86.6	100.0	93.2
ŏ	39.317	9.00	9.66	9.66	9.66	9.33	80.0	93.2	93.2	93.2	86.6
Root wood	19.659	8.00	9.66	9.66	9.66	10.00	60.0	93.2	93.2	100.0	100.0
ot	9.829	7.00	7.66	8.66	8.66	8.66	40.0	53.2	73.2	80.0	73.2
Ro	4.915	8.33	7.33	9.00	9.00	9.00	66.6	46.6	80.0	66.6	80.0
—	2.457	6.33	6.66	8.66	8.66	8.33	26.6	33.2	73.2	53.2	66.6

Table 2. Repellency of *T. castaneum* adults by chloroform extract of leaf, stem bark, stem wood, root bark and root wood of *G. pentaphylla*

Table 3. ANOVA results of repellency by chloroform extract of leaf, stem bark, stem wood, root bark and root wood of *G. pentaphylla* against *T. castaneum* adults.

Sources of variation		SS	Df	MS	F-values
Leaf					
Between doses	42904.49		5	8580.897	60.98325***
Between hourly observations	1419.20		4	354.800	2.521516ns
Error	2814.18		20	140.709	
Total	47137.87		29		
Stem bark					
Between doses	269	948.51	5	5389.703	14.17715***
Between hourly observations	27	745.83	4	686.457	1.805666ns
Error	70	503.37	20	380.168	
Total	37297.71		29		
Stem wood					
Between doses	24779.38	5		4955.876	19.43674***
Between hourly observations	1339.77	4		334.943	1.313633ns
Error	5099.49	20		254.975	
Total	31218.64	29			
Root bark					
Between doses	20117.29	5		4023.457	15.42936***
Between hourly observations	3617.81	4		904.452	3.46844*
Error	5215.33	20		260.766	
Total	28950.42	29			
Root wood					
Between doses	2576.38	5		515.276	1.081782ns
Between hourly observations	2423.46	4		605.864	1.271965ns
Error	9526.43	20		476.322	
Total	14526.27	29			

*=P<0.05; *** =P<0.001; ns= not significant

Since types or category of compounds of this plant is already known it is also known that this plant might have some bioactive potential. The result of this investigation has been added a clue in this connection showing cytotoxic and repellent activity of the chloroform extract of *G. pentaphylla*. Chinese researchers reviewed the plants used for pest insect control in China and found that there was a strong connection between medicinal and pesticidal plants (Jembere *et al.*, 1995). So, being a medicinal plant and originally being confined to a certain area of Asia and Australia this plant deserves attention to be investigated especially for its bioactive potentials.

Acknowledgements

The authors are thankful to the University of Rajshahi for a Research Grant and also to the Chairman, Department of Zoology, University of Rajshahi, Rajshahi 6205, Bangladesh for providing laboratory facilities.

References

- Anon, 1952. "The wealth of India" A dictionary of Indian raw materials and industrial products, CSIR, New Delhi, 3, 69.
- Chakravarty AK, Das B, Masuda K, Ageta H. 1996. Tetracyclic triterpenoids from *Glycosmis arborea*. Phytochemistry 42(4), 1109-1113.
- Chopra RN, Nayar SL, Chopra IC. 1956. Glossary of Indian Medicinal PlantsCSIR. pub., New Delhi (India) pp. 21.
- Greger H, Zechner G, Hofer O, Vajrodaya S. 1996. Bioactive amides from *Glycosmis* species. J Nat Prod 59, 1163-1168.
- Hofer O, Vajrodaya S, Greger H. 1998. Phenethylamides with an unusual 4-oxo-2-oxolenyl terpenoid side chain from *Glycosmis* species. Monatsh Chem 129 (2), 213-219.
- Hofer O, Zechner G, Vajrodaya S, Lutz G, Greger H. 1995a. New anthranilic and methylsulfonylpropenoic acid amides from Thai *Glycosmis* species. Liebigs Ann Chem 1789-1794.
- Hofer O, Zechner G, Wurz G, Hadacek F, Greger H. 1995b. Ritigalin, a new thiocarbonic acid imide from *Glycosmis* species. Monatsh Chem 126, 365-368.
- Jembere B, Obeng-Ofori D, Hassanali A, Nyamasyo GNN. 1995. Products derived from the leaves of *Ocimum kilimanndscharicum* (Labiatae) as post-harvest grain protectants against the infestation of three major stored product insect pests. Bull Entomol Res 85, 361-367.
- Khan AR, Selman BJ. 1981. Some techniques for minimizing the difficulties in egg counting of *Tribolium castaneum* (Herbst). Ent Rec Var 93, 36-37.
- Kirtikar KR, Basu BD. 1935. Indian Medicinal Plants. Lalit Mohan Basu Pub. Allahabad (India), 1, 551.

- McDonald LL, Guy RH, Speirs RD. 1970. Preliminary evaluation of new candidate materials as toxicants, repellents and attractants against stored-product insects. Agricultural Research Service, U. S. Department of Agriculture, Washington D.C., Marketing Research Report No. 882.
- Muthukrishnan J, Seifert K, Hoffmann H, Lorenz MW. 1999. Inhibition of juvenile hormone biosynthesis in *Gryllus bimaculatus* by *Glycosmis pentaphylla* leaf compounds. Phytochemistry 50, 249-254.
- Ono T, Ito C, Furukawa H, Wu TS, Kuoh CS, Hsu KS. 1995. Two new acridone alkaloids from *Glycosmis* species. J Nat Prods 58, 1629-1631.
- Park T, Frank MB. 1948. The fecundity and development of the flour beetles, *Tribolium castaneum* and *Tribolium confusum* at three constant temperatures. Ecology 29, 386-375.
- Rahmani M, Ling CY, Sukari MA, Ismail HBM, Meon S, Aimi N. 1998. 7-methoxyglycomaurin: A new carbazole alkaloid from *Glycosmis rupestris*. Planta Med 64, 780-780.
- Rastogi K, Kapil PS, Popli SP. 1980. New alkaloids from leaves of *Glycosmis mauritiana*, Phytochemistry 19, 945-948.
- Shapiro JP, Bowman KD, Smith H. 1997. Resistance of citrus rootstocks and *Glycosmis pentaphylla* against larval citrus root weevils *Diaprepes abbreviatus*, in live root or diet incorporation assays. Fla Entomol 80, 471-477.
- Stone BC. 1985. A conspectus of the genus *Glycosmis* Correa: Studies in Malesian Rutaceae, III. Proc Acad Natl Sci Philadelphia 137, 1-27.
- Talukder FA, Howse PE. 1995. Evaluation of *Aphanamixis* polystachya as a source of repellant, antifeedants, toxicants and protectants in storage against *Tribolium castaneum* (Herbst). J Stored Prod Res 31(1), 55-61.
- Talukder FA, Howse PE. 1993. Deterrent and insecticidal effects of extracts of pithraj, *Aphanamixis polystachya* (Mcliaceae), against *Tribolium castaneum* in storage. J Chem Ecol 19, 2463-2471.
- Talukder FA, Howse PE. 1994. Efficacy of pithraj (*Aphanamixis polystachya*) seed extracts against storedproduct pests. Proc Int Working Conf on Stored-prod Protec 2, 848-852.
- Wu TS, Furukawa H, Kuoh C-S, Hsu K-S. 1983. Acridone alkaloids. Part 9. Chemical constituents of *Glycosmis citrifolia* (Willd.) Lindl. Structures of novel linear pyranoacridones, furoacridones, and other new acridone alkaloids. J Chem Soc Perkin Trans 1, 1681-1688.
- Wurz G, Hofer O, Greger H. 1993. Structure and synthesis of phenaglydon, a new quinolone derived phenanthridine alkaloid from *Glycosmis cyanocarpa*. Nat Prod Let 3(3), 177-182.