Short Communication REPELLENT ACTIVITY OF DERRIS INDICA EXTRACTS AGAINST TRIBOLIUM CASTANEUM (HERBST) ADULTS

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mivsk: Putji #Kuo †cukv (Tribolium castaneum)-Gi Ici Derris indica Bennet. (KiÄy)-Gi dtji #Luny, cuZy, gtji evKj, gtji KW, evR, Kutži evKj I Kutži Kutvi #Kutiudg@bh@n cüputMG_yi usZvob KuhRuiZv chfe¶Y Kivnq, hutZ dtji #Luny, cuZy, evR, Kutži evKj ubh@nmuqZv † utqtQ- hLb ubh@yi uscintZ F-ratios (df 7, 28) utj h_yutg 5.08, 12.97, 15.27 Ges 11.08 Aek" cuZy, evR I Kutži evKj ubh@ ZjbugjKfute fyi mpuqZv† utqtQ (P<0.001), dtji #Lunni t¶tĨ hvuQj Atc¶KZ.`efP (P<0.05), Ges gtji evKj I gtji KW †KubB KuhRuiZv † Lupub; Avi KuhRuiZvi gulibuhupubh@yi#K evR > cuZv> Kutži evKj > dtji #LunvµgViputi mRub huq)

Key words: Repellent activity, Derris indica, Tribolium castaneum.

The Pongam tree *Derris indica* Bennet. (Family Fabaceae) is cultivated mainly for two purposes: an ornamental plant in gardens and along avenues and roadsides, for its fragrant wisteria like flowers, and a host plant for lac insects (Rama Murti and Seshadri 1944). The gardeners use well-decomposed flowers as compost for plants requiring rich nutrients. It is appreciated as an ornamental plant throughout coastal India and all of Polynesia. This tree is an inhabitant of SriLanka, Myanmar, Thailand, Laos, Cambodia, Vietnam, Malaysia, Indonesia and North Australia, and occurs in Bangladesh mainly in the tidal forests and also often along river and canal banks in all districts (Kirtikar and Basu 1935).

D. indica is a medicinal plant, the dried flowers of which are used in decoction to quench thirst in diabetes; and the seeds are mainly valued for the oil obtained from them, which has many industrial and medicinal uses (Kirtikar and Basu 1935). The Pongam oil is nonedible and used for tanning leather, as an ingredient in soap; as a liniment to treat scabies, herpes and rheumatism and as an illuminating oil (Burkill 1966). Powdered seed is valued as a febrifuge and tonic, and used also in bronchitis and whooping cough (Kirtikar and Basu 1935). It is also used to lubricate cattle cartwheels by the native people of Bangladesh. In addition, the juice of the leaves is prescribed in flatulence dyspepsia, diarrhoea and cough; and also is considered as a remedy for leprosy and gonorrhoea. A hot infusion of the leaves is used as a medicated bath for relieving rheumatic pains, and for cleansing foul ulcers and sores. The stem-bark is fibrous and is used for cordage. The fresh bark has a feebly sweetish and mucilaginous taste at first but soon becomes bitter combined with a sort of pungency. It is given internally in bleeding piles (haemorrhoids). A decoction of the bark is used for beriberi. It shows positive tests for the presence of alkaloids, triterpenoid and saponin. A paste of the roots is used for local application in scrofulous enlargements. Extracts of the leaves were active against Micrococcus pyogenes var. aureus (Anon 1969). The juice of the roots is used for cleansing foul ulcers and closing fistulous sores and for cleaning teeth and strengthening gums. Roots are used as fish-poison by the aborigines of Australia (Kirtikar and Basu 1935). Insecticidal activity of this plant against the pulse beetle Callosobruchus maculatus had been traced (Mondal and Islam 2008). Moreover, both antibacterial and larvicidal potentials of this plant had also been reported (Mondal et al. 2010). Being a member of the rotenone containing family Fabaceae this plant deserves an especial attention, however, still the information on biological activities of D. indica is scanty, but it requires a thorough investigation of its properties to step up for their possible use in the pest control technology. This is just a part of a research endeavor entitled characterization of Karanja, where biological activities and phytochemical investigation of D. indica components have been targeted.

The fresh leaves, fruit shell, root bark, root wood, seeds, stem bark, and stem wood of *D. indica* were collected from the campus of the University of Rajshahi,

Bangladesh. After drying the plant parts were powdered in a grinder machine separately avoiding excess heat during grinding. Chloroform was selected to extract seven different parts of *D. indica* separately. The ground dried materials were extracted with sufficient amount of chloroform ($500g \times 1500ml \times 3$ times) for each of the items. Separate extracts were collected by the cool method after 72h of plunging for each of the materials. Extracts thus obtained as residue after filtration and evaporation of the solvent were kept in a refrigerator with proper labeling.

Table 1. Ranges in percent repulsion (PR) of chloroform extracts assessed for various parts of D. indica against adult T. castaneum.

Doses	Fruit shell	Leaves	Root bark	Root wood	Stem bark	Stem wood	Seed
µg cm ⁻²	(PR ranges)	(PR ranges)	(PR ranges)	(PR ranges)	(PR ranges)	(PR ranges)	(PR ranges)
12	33.32-80.00	33.32-46.66	20.00-80.00	13.32-66.66	20.00-80.00	26.60-73.20	20.00-60.00
25	20.00-80.00	33.32-66.66	6.66-80.00	0.00-53.32	40.00-66.66	33.20-80.20	40.00-73.32
49	46.66-60.00	80.00-93.32	6.66-80.00	6.66-80.00	33.32-53.32	46.60-73.20	0.00-20.00
98	6.66-53.32	66.66-100.00	6.66-66.66	13.32-60.00	46.66-73.32	53.20-86.60	33.32-53.32
197	0.00-6.66	46.66-93.32	20.00-80.00	0.00-60.00	0.00-13.32	40.00-86.60	26.66-66.66
393	20.00-26.66	86.66-100.00	33.32-80.00	0.00-40.00	0.00-33.32	6.60-80.00	13.32-53.32
786	0.00-60.00	33.32-80.00	33.32-66.66	0.00-53.32	0.00-13.32	0.00-53.20	33.32-66.66
1573	20.00-53-32	60.00-93.32	46.66-80.00	6.66-66.66	0.00-33.32	53.33-86.66	66.66-93.32

Table 2. ANOVA components showing the effects of various parts of *D. indica* extracts on *T. castaneum* adult by repellency tests.

Test	Sources of	Df	F-ratios
materials	variance		
Fruit shell	Doses	7/28	5.08***
	Time intervals	4/28	0.67ns
Leaf	Doses	7/28	12.97***
	Time intervals	4/28	5.75**
Seed	Doses	7/28	15.27***
	Time intervals	4/28	1.21ns
Stem bark	Doses	7/28	11.08***
	Time intervals	4/28	2.83*

Df = degrees of freedom; * = P <0.05; ** = P <0.01; *** = P <0.001; *** = P <0.001; ns = not significant.

A general concentration for each of the extracts as stock dose was prepared to make other successive doses by serial dilution that gave 1573, 787, 393, 197, 98, 49, 25 and 12 μ gcm⁻² concentrations for surface film application. The repellency test used was adopted from the method (No. 3) of McDonald et al. (1970) with some modifications by Talukder and Howse (1993, 1994). Half filter paper discs (Whatman No. 40, 9cm diameter) were prepared and selected doses of all extracts were applied separately onto each of the halfdiscs and allowed to dry out as exposed in the air for 10 min. Each treated half-disc was then attached lengthwise, edge-to-edge, to a control half-disc with adhesive tape and placed in Petri dishes. The orientation of the same was changed in the replica to avoid the effects of any external directional stimulus affecting the distribution of the test insects. Ten adult insects were released in the middle of each of the filter paper circles. Each concentration was tested five times. Insects that settled on each half of the filter paper discs were counted after 1h and then at hourly intervals for 5 hrs. The average of the counts was converted to percent repellency (PR) using the formula of Talukder and Howse (1993, 1995): PR = (Nc – 5) × 20, where Nc was the number of insects present on the control half of the disc. Positive values expressed for repellency and negative values for attractant activities. The data recorded as PR were again converted by *arcsin* transformation for the calculation of ANOVA.

Among the seven extracts the fruit shell, leaves, seed, and stem bark extracts showed repellent activity (Table 1), while the ANOVA results (Table 2) offered F-values 5.08, 12.97, 15.27 and 11.08 respectively for 7 degrees of freedom with an error df of 28 to reveal that the leaves, seeds and the stem bark contain stronger repellent properties (P<0.001) than the fruit shell extract (P<0.05). However, the stem wood, root bark and root wood extracts showed no repellency at all. The P values were found 2.602E-07, 0.000814223, 4.759E-08, 1.235E-06 for the leaf, fruit shell, seed, stem bark extracts respectively and thus the intensity of repellent activity of the test materials it could be arranged in descending order seed > leaf > stem bark > fruit shell. This finding is similar to that of Khan (1983) who also reported the repellent activity of D. indica against stored product insect pests. Sighamony et al. (1984) reported that the oils of cedar wood, karanja and acetone extract of black pepper were more potent than other standard

repellents and the repellency of karanja oil persist strongly over the 8 weeks of experimental period. However, this plant possesses efficacy other than insect repulsion, while antibacterial activity of 3 *Derris* species' extractives collected in different organic solvents revealed by Khan *et al.* (2006), and this work confers no antifungal activity. Biswall *et al.* (2011) confirmed presence of flavonoids, alkaloids, saponins and steroids through phytochemical screenings.

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