



Bangladesh. After drying the plant parts were powdered in a grinder machine separately avoiding excess heat during grinding. Chloroform was selected to extract seven different parts of *D. indica* separately. The ground dried materials were extracted with sufficient amount of chloroform (500g × 1500ml × 3 times) for each of the

items. Separate extracts were collected by the cool method after 72h of plunging for each of the materials. Extracts thus obtained as residue after filtration and evaporation of the solvent were kept in a refrigerator with proper labeling.

**Table 1.** Ranges in percent repulsion (PR) of chloroform extracts assessed for various parts of *D. indica* against adult *T. castaneum*.

Doses μg cm <sup>-2</sup>	Fruit shell (PR ranges)	Leaves (PR ranges)	Root bark (PR ranges)	Root wood (PR ranges)	Stem bark (PR ranges)	Stem wood (PR ranges)	Seed (PR ranges)
12	33.32-80.00	33.32-46.66	20.00-80.00	13.32-66.66	20.00-80.00	26.60-73.20	20.00-60.00
25	20.00-80.00	33.32-66.66	6.66-80.00	0.00-53.32	40.00-66.66	33.20-80.20	40.00-73.32
49	46.66-60.00	80.00-93.32	6.66-80.00	6.66-80.00	33.32-53.32	46.60-73.20	0.00-20.00
98	6.66-53.32	66.66-100.00	6.66-66.66	13.32-60.00	46.66-73.32	53.20-86.60	33.32-53.32
197	0.00-6.66	46.66-93.32	20.00-80.00	0.00-60.00	0.00-13.32	40.00-86.60	26.66-66.66
393	20.00-26.66	86.66-100.00	33.32-80.00	0.00-40.00	0.00-33.32	6.60-80.00	13.32-53.32
786	0.00-60.00	33.32-80.00	33.32-66.66	0.00-53.32	0.00-13.32	0.00-53.20	33.32-66.66
1573	20.00-53-32	60.00-93.32	46.66-80.00	6.66-66.66	0.00-33.32	53.33-86.66	66.66-93.32

**Table 2.** ANOVA components showing the effects of various parts of *D. indica* extracts on *T. castaneum* adult by repellency tests.

Test materials	Sources of variance	Df	F-ratios
Fruit shell	Doses	7/28	5.08***
	Time intervals	4/28	0.67ns
Leaf	Doses	7/28	12.97***
	Time intervals	4/28	5.75**
Seed	Doses	7/28	15.27***
	Time intervals	4/28	1.21ns
Stem bark	Doses	7/28	11.08***
	Time intervals	4/28	2.83*

Df = degrees of freedom; \* = P < 0.05; \*\* = P < 0.01; \*\*\* = P < 0.001; ns = not significant.

A general concentration for each of the extracts as stock dose was prepared to make other successive doses by serial dilution that gave 1573, 787, 393, 197, 98, 49, 25 and 12 μgcm<sup>-2</sup> concentrations for surface film application. The repellency test used was adopted from the method (No. 3) of McDonald *et al.* (1970) with some modifications by Talukder and Howse (1993, 1994). Half filter paper discs (Whatman No. 40, 9cm diameter) were prepared and selected doses of all extracts were applied separately onto each of the half-discs and allowed to dry out as exposed in the air for 10 min. Each treated half-disc was then attached lengthwise, edge-to-edge, to a control half-disc with adhesive tape and placed in Petri dishes. The orientation of the same was changed in the replica to avoid the effects of any external directional stimulus affecting the distribution of the test insects. Ten adult insects were

released in the middle of each of the filter paper circles. Each concentration was tested five times. Insects that settled on each half of the filter paper discs were counted after 1h and then at hourly intervals for 5 hrs. The average of the counts was converted to percent repellency (PR) using the formula of Talukder and Howse (1993, 1995):  $PR = (Nc - 5) \times 20$ , where Nc was the number of insects present on the control half of the disc. Positive values expressed for repellency and negative values for attractant activities. The data recorded as PR were again converted by *arcsin* transformation for the calculation of ANOVA.

Among the seven extracts the fruit shell, leaves, seed, and stem bark extracts showed repellent activity (Table 1), while the ANOVA results (Table 2) offered F-values 5.08, 12.97, 15.27 and 11.08 respectively for 7 degrees of freedom with an error df of 28 to reveal that the leaves, seeds and the stem bark contain stronger repellent properties (P < 0.001) than the fruit shell extract (P < 0.05). However, the stem wood, root bark and root wood extracts showed no repellency at all. The P values were found 2.602E-07, 0.000814223, 4.759E-08, 1.235E-06 for the leaf, fruit shell, seed, stem bark extracts respectively and thus the intensity of repellent activity of the test materials it could be arranged in descending order seed > leaf > stem bark > fruit shell. This finding is similar to that of Khan (1983) who also reported the repellent activity of *D. indica* against stored product insect pests. Sighamony *et al.* (1984) reported that the oils of cedar wood, karanja and acetone extract of black pepper were more potent than other standard

repellents and the repellency of karanja oil persist strongly over the 8 weeks of experimental period. However, this plant possesses efficacy other than insect repulsion, while antibacterial activity of 3 *Derris* species' extractives collected in different organic solvents revealed by Khan *et al.* (2006), and this work confers no antifungal activity. Biswall *et al.* (2011) confirmed presence of flavonoids, alkaloids, saponins and steroids through phytochemical screenings.

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