

Xanthone from the Fruits of *Terminalia Arjuna*

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Abstract

2,7-dimethoxyxanthone (**1**) along with a well known compound β -sitosterol-3-O- β -D-glucopyranoside (**2**) were isolated from the methanolic extract of the fruits of *Terminalia arjuna*. Their structures were elucidated by spectroscopic (NMR and mass) analysis.

Keywords: *Terminalia arjuna*; Combretaceae; 2,7-Dimethoxyxanthone.

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1. Introduction

Terminalia arjuna Wight & Arn. (Combretaceae) is distributed in tropical and subtropical regions of the world. Its bark decoction is being used in the Indian subcontinent for anginal pain, hypertension, congestive heart failure, and dyslipidemia, based on the observations of ancient physicians for centuries [1-3]. Fruits and Bark of *T. arjuna* have been used as diuretic [1], tonic [4] an antioxidant [5], anticancer [6,7] hypocholesterolaemic, hypolipidemic [8-10], antiulcer [11], antiviral, antifungal and antibacterial [12-16] agents. A number of triterpenoids, saponines, tannins, cardenolides and flavonoids were reported from earlier phytochemical investigations of *T. arjuna* [17-24]. In the present study, we have isolated one xanthone identified as 2,7-dimethoxyxanthone (**1**) [Fig. 1]. Herein we report the isolation for the first time and structure elucidation of a known compound from *Terminalia arjuna*. The structure was identified on the basis of ¹H and ¹³C NMR experiments and mass spectroscopic data, UV, FTIR and by comparison with the authentic data of the compound published in the literatures [25,26].

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2. Experimental

2.1. Instruments

NMR Spectra: *Bruker spectrometer* (^1H 600 & 500 MHz and ^{13}C 150 & 100 MHz) and residual protons used as internal standard. EI mass: *JEOL* (JMS 600H) mass spectroscopy. IR: *SIMADZU* FTIR (8900). UV: *HITACHI* (U-3200) spectrophotometer. Optical rotation: *JASCO* (P-2000) polarimeter. Melting point: *BUCHI* 535 digital Melting Point apparatus and were uncorrected. TLC spots were detected by UV light and fixed upon spraying ceric sulfate reagent followed by heating.

2.2. Plant material

The fruits of *Terminalia arjuna* W. & A. were collected from the campus of University of Rajshahi, Bangladesh, during September 2007 and was authenticated by Bangladesh National Herbarium, Dhaka, Bangladesh. A voucher specimen has been deposited (Acc. No. 32612, DACB) in the Bangladesh National Herbarium, Dhaka, Bangladesh.

2.3. Extraction and isolation

Air dried fruits of *T. arjuna* was powdered (18 kg) and extracted with methanol (MeOH) (60 Lx3) at room temperature for 3 days. MeOH was evaporated under vacuum to give a solid product (850 g), which was dissolved again in MeOH and partitioned with n-hexane to remove sticky and gummy materials. MeOH was evaporated under vacuum to give a residue (590 g). 200 g from this residue was suspended in H_2O and partitioned with dichloromethane (5 Lx3), ethyl acetate (5 Lx3) and n-butanol (5 Lx3) successively. The Ethyl acetate fraction (E, 14.5 g) was subjected to column chromatography over silica gel (30x7 cm) eluted with ethyl acetate : n-hexane and the fractions were collected at 20% ethyl acetate (E-1), 50% ethyl acetate (E-2), 75% ethyl acetate (E-3), 100% ethyl acetate (E-4) and 100% methanol (E-5). Fraction E-1 (0.6 g) was subjected to column chromatography with silica gel (25x2.5 cm) and eluted with 20% ethyl acetate : n-hexane to give a mixture which on further purification by preparative TLC over silica gel with 40% acetone : n-hexane as eluent gave a compound 2,7-dimethoxyxanthone (**1**) [Fig. 1] (2.8 mg). Known compound β -sitosterol-3-O- β -D-glucopyranoside (**2**) [Fig. 3] (21.4 mg) [27-29] was obtained from fraction E-3 by reversed-phase C18 silica gel and Sephadex LH 20 column chromatography.

2.3.1. 2,7-Dimethoxyxanthone (1)

Physical state: pale brown amorphous solid; Yield: 2.8 mg (4.74×10^{-5} %); M.P.: 174-176°C (Lit²⁵ mp: 172-174 °C); EI MS (rel. int. %) *m/z*: 256(6) [M^+], 221(6), 169(9), 168(84), 167(7), 149(10), 138(16), 137(100), 121(14), 109(43); FAB MS (+ve) *m/z*: 257 [$\text{M} + \text{H}]^+$; FAB MS (-ve) *m/z*: 255 [$\text{M} - \text{H}]^+$; UV λ_{max} (MeOH) (log ϵ) nm: 216 (2.4), 249

(2.3), 302 (2.5); IR ν_{\max} (CHCl₃) cm⁻¹: 1660, 1588, 1455, 900-650; ¹H NMR (600 MHz, solvent: CDCl₃, δ in ppm): Shown in Table-1; ¹³C NMR (150 MHz, solvent: CDCl₃, δ in ppm): Shown in Table-1.

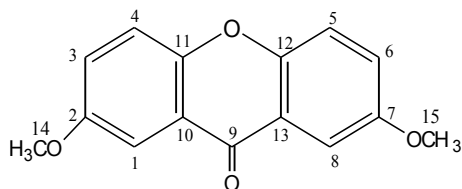
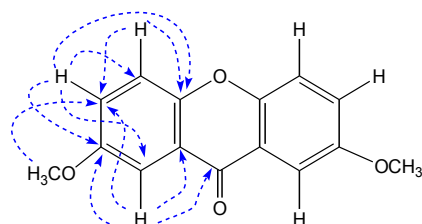
2.3.2. β -sitosterol-3-O- β -D-glucopyranoside (2)

Physical state: White amorphous solid; Yield: 21.4 mg (3.63 x 10⁻⁴ %); M.P.: 274-277°C (Lit.²⁷ : 272-274°C); [α]_D²⁵: - 18.7° (c 0.4, MeOH); IR ν_{\max} (KBr) cm⁻¹: 3408, 3050, 1690, 1618, 1595, 1550; EI MS (rel. int. %) m/z : 414 (7), 396 (100), 381 (13), 288 (10), 255 (17), 161 (14), 147 (21), 95 (16) 57 (14), 43 (16); FAB MS (-ve) m/z : 575 [M - H]⁺; HR FAB MS m/z : 575.0210 (calcd 575.0199 for C₃₅H₆₀O₆); ¹H-NMR (C₅D₅N, 500 MHz) δ_H : see Table-2; ¹³C-NMR (C₅D₅N, 100 MHz) δ_C : see Table-2.

3. Results and Discussion

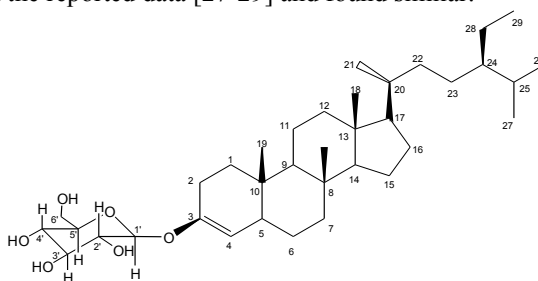
Compound **1** [Fig. 1] was isolated as a brown amorphous solid with [α]_D²⁵: -9.6° (c 2.5, CH₃OH). The molecular formula C₁₅H₁₂O₄ with ten degrees of unsaturation was determined by EI MS at m/z 256 [M⁺], FAB MS (positive ion mode) at m/z 257, FAB MS (negative ion mode) at m/z 255 and ¹³C NMR (BB), DEPT 90° and DEPT 135° experiment data. IR spectrum has displayed bands for carbonyl group (1660 cm⁻¹) with aromatic ring (1588, 1455, 900-650 cm⁻¹) functionalities [30]. UV spectrum (MeOH) exhibited absorptions at 216, 249 and 302 nm characteristic of a furanocoumarin moiety [31]. ¹³C NMR (BB), DEPT 135° and DEPT 90° experiments showed eight signals for 15 carbon atoms, which includes two methyl, six methylene and seven quaternary carbons. One quaternary carbon has showed resonance at δ_C 168.8 (C-9) for a carbonyl group. Due to plane of symmetry, all others carbons and protons in compound **1** appeared as one signal for every two symmetric carbons and protons in ¹H and ¹³C NMR spectra, respectively. Two oxygenated quaternary carbons showed resonance at δ_C 146.2 (C-11/12), two quaternary carbons at δ_C 122.5 (C-10/13) and two oxygenated aromatic carbons at δ_C 151.7 (C-2/7). Three signals appeared at δ_C 123.6, 117.3 and 115.8 (C-3/6, C-1/8, C-4/5 res.) in aromatic region and another resonance at δ_C 52.3 (2x-OCH₃), integrated for two carbons. Two methoxy groups [δ_H/δ_C 3.82/52.3] were identified at identical positions 2 and 7, confirmed by HMBC correlations [Fig. 2] of H-1 with C-2, C-10, C-9, C-3, C-11; H-3 with C-2, C-4, C-11 and H-4 with C-3, C-2, C-11. Methoxy protons have exhibited HMBC correlation with C-3 atoms.

Compound **1** showed two aromatic protons at δ_H 7.41 as a doublet ($J=2.4$ Hz, H-1 and H-8), other two aromatic protons at δ_H 7.39 as a doublet of doublet ($J=2.4, 9.0$ Hz, H-3 and H-6), and another two aromatic protons at δ_H 6.79 as a doublet ($J=9.0$ Hz, H-4 and H-5). A signal appeared as singlet at δ_H 3.82 integrated for six protons were assigned for two methoxy groups. Thus, the structure of compound **1** was established as 2,7-dimethoxyxanthone which was synthesized by Menéndez and Mahfouz and their co-worker [25,26]. Compound **1** was isolated first time from any natural source.

Fig. 1. 2,7-Dimehoxyxanthone (**1**).Fig. 2. HMBC correlation of **1**.Table 1. ^1H and ^{13}C NMR data (600 & 150 MHz) of compound **1** (solvent: CDCl_3 , δ in ppm).

C No.	Mult.	δ_{C}	δ_{H} (m, J Hz)
C1/C8	CH	117.3	7.41 (d, J = 2.4 Hz)
C2/C7	C	151.7	-
C3/C6	CH	123.6	7.39 (dd, J = 2.4, 9.0 Hz)
C4/C5	CH	115.8	6.79 (d, J = 9.0 Hz)
C9	C	168.8	-
C10/C13	C	122.5	-
C11/C12	C	146.2	-
C14/C15	CH_3	52.3	3.82 (s)

Known compound β -sitosterol-3-O- β -D-glucopyranoside (**2**) [Fig. 3] showed melting point 274-277°C (in Lit.²⁷: 272-274°C), and FAB MS (-ve ion mode) showed peak at m/z 575 $[\text{M}-\text{H}]^+$, HR FAB MS showed peak at m/z 575.0210 (calcd 575.0199 for $\text{C}_{35}\text{H}_{60}\text{O}_6$) was compared with the reported data [27-29] and found similar.

Fig. 3. β -Sitosterol-3-O- β -D-glucopyranoside (**2**).Table 2. ^1H and ^{13}C NMR data of compound **2** (solvent: $\text{C}_5\text{D}_5\text{N}$, δ in ppm).

C No.	Mult.	2 †		Ref ²⁷ ‡	
		δ_{C}	δ_{H} (m, J Hz)	δ_{C}	δ_{H} (m, J Hz)
1	CH_2	37.4	2.72 (dd, 11.5, 2.5 Hz)	36.85	1.25 (m, 2H)
2	CH_2	30.1	2.21, 1.72 (d, 11.5 Hz)	29.12	1.33 (m, 2H)
3	CH	78.5	4.29, 3.99 (dd, 8.0 Hz)	78.61	3.13 (m, 1H)
4	CH_2	39.3	2.46 (dd, 11.0 Hz)	42.12	2.14 (m, 2H)
5	C	140.8	-	139.98	-
6	CH	138.8	5.05 (tbr, 2.5 Hz)	121.54	5.09 (brs, 1H)
7	CH_2	32.1	1.52, 1.88 (overlap)	31.41	1.73 (m, 2H)
8	CH	32.0	1.35 (overlap)	31.46	1.22 (m, 1H)
9	CH	50.2	0.89 (s)	49.83	1.22 (m, 1H)

C No.	Mult.	2 †		Ref ²⁷ ‡	
		δ_C	δ_H (m, J Hz)	δ_C	δ_H (m, J Hz)
10	C	36.9	-	36.27	-
11	CH ₂	21.2	1.38, 1.05 (overlap)	20.21	1.33 (m,2H)
12	CH ₂	39.9	1.94, 1.05 (overlap)	38.20	1.33 (m, 2H)
13	C	42.4	-	41.88	-
14	CH	56.7	0.92 (overlap)	56.36	1.22 (m,1H)
15	CH ₂	24.5	1.55, 1.03 (overlap)	23.79	1.73 (m, 2H)
16	CH ₂	28.5	1.81, 1.23 (overlap)	27.76	1.73 (m, 2H)
17	CH	56.1	1.07	55.66	1.73 (m, 2H)
18	CH ₃	11.9	0.63 (s)	11.27	0.62 (s, 3H)
19	CH ₃	19.4	0.88 (d)	19.10	0.94 (s, 3H)
20	CH	36.3	1.36 (overlap)	35.70	1.32 (m, 1H)
21	CH ₃	18.9	0.92 (d, 6.5 Hz)	18.69	0.84 (d, 3H, J=6.3 Hz)
22	CH ₂	34.1	1.38,1.06 (o)	33.51	1.73 (m, 2H)
23	CH ₂	26.2	1.21 (o)	25.64	1.73 (m, 2H)
24	CH	45.9	0.97 (o)	45.49	1.12 (m, 1H)
25	CH	29.3	1.70 (o)	28.74	2.14 (m, 1H)
26	CH ₃	19.1	0.83 (s)	18.69	0.75 (d, 3H, J=7.7 Hz)
27	CH ₃	19.9	0.89 (s)	18.35	0.73 (d, 3H, J=1.6 Hz)
28	CH ₂	23.3	1.26 (o)	22.60	1.33 (m, 2H)
29	CH ₃	12.1	0.87 (o)	12.29	0.77 (t, 3H, J=6.9 Hz)
1'	CH	102.5	5.33 (d, 7.5 Hz)	100.74	4.11 (d, 1H, J=7.8 Hz)
2'	CH	78.6	4.31 (d, 9.0 Hz)	73.21	3.41 (m, 1H)
3'	CH	78.0	3.99 (o)	76.18	3.41 (m, 1H)
4'	CH	71.6	4.29 (d, 8.0 Hz)	69.90	3.41 (m, 1H)
5'	CH	75.3	4.06 (t, 8 Hz)	75.62	3.06 (m, 1H)
6'	CH ₂	62.7	4.41, 4.56 (d, 9.5, 5.0Hz)	61.36	2.94 (m, 2H)

¹H & ¹³C: † 500 & 100 MHz, Solvent C₅D₅N; ‡ 400 & 100 MHz, Solvent: CDCl₃

4. Conclusion

Synthesized compound **1** was isolated from the methanolic extract of the fruits of *Terminalia arjuna* and the structure was identified as 2,7-dimethoxyxanthone which was isolated first time from any natural source, confirmed by SciFinder[®]. Compound **2** showed the physical and spectral data similar to the reported.

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References

1. R. N. Chopra, I. C. Chopra, K. L. Handa, and L. D. Kapur, *Indigenous Drugs of India*, U.N. (Dhur and Sons Pvt. Ltd., Calcutta, 1958) pp. 421.
2. S. Dwivedi and D. Chopra, *J. Tradit. Complement Med.* **4**, 224 (2014).
<http://dx.doi.org/10.4103/2225-4110.139103>

3. S. K. Maulik and K. K. Talwar, *Am. J. Cardiovasc. Drugs* **12**, 157 (2012).
<http://dx.doi.org/10.2165/11598990-000000000-00000>
4. G. Balamurugan, P. Muralidharan, and S. Selvarajan, *J. Sci. Res.* **1**, 153 (2009).
<http://dx.doi.org/10.3329/jsr.v1i1.1057>
5. P. P. Singh and S. M. Chauhan, *Nat. Prod. Res.* **28**, 760 (2014).
<http://dx.doi.org/10.1080/14786419.2014.919287>
6. V. Gangadevi and J. Muthumary, *Appl. Biochem. Biotechnol.* **158**, 675 (2009).
<http://dx.doi.org/10.1007/s12010-009-8532-0>
7. P. L. Kuo, Y. L. Hsu, T. C. Lin, L. T. Lin, J. K. Chang, and C. C. Lin, *Anti-Cancer Drugs* **16**, 409 (2005). <http://dx.doi.org/10.1097/00001813-200504000-00007>
8. R. Gupta, S. Singhal, A. Goyle, and V. N. Sharma, *J. Assoc. Physicians India* **49**, 231 (2001).
9. A. Ram, P. Lauria, R. Gupta, P. Kumar, and V. N. Sharma, *J. Ethnopharmacol.* **55**, 165 (1997).
[http://dx.doi.org/10.1016/S0378-8741\(96\)01493-6](http://dx.doi.org/10.1016/S0378-8741(96)01493-6)
10. S. Subramaniam, S. Ramachandran, S. Uthrapathi, V.R. Gnamanickam, and G. P. Dubey, *Ind. J. Exp. Biol.* **49**, 282 (2011).
11. R. S. Devi, S. Narayan, G. Vani, P. Srinivasan, K. V. Mohan, K. E. Sabitha, and C. S. Devi, *Phytother. Res.* **21**, 762 (2007). <http://dx.doi.org/10.1002/ptr.2160>
12. H. Y. Cheng, C. C. Lin, and T. C. Lin, *Antiviral Res.* **55**, (2002).
13. K. R. Aneja, C. Sharma, and R. Joshi, *Braz. J. Otorhinolaryngol.* **78**, 68 (2012).
<http://dx.doi.org/10.1590/S1808-86942012000100011>
14. S. L. Shinde, S. B. Junne, S. S. Wadje, and M. M. Baig, *Pak. J. Bio. Sci.* **12**, 1483 (2009).
<http://dx.doi.org/10.3923/pjbs.2009.1483.1486>
15. M. Saxena, S. Yadav, D. U. Bawankule, S. K. Srivastava, A. Pal, R. Mishra, M. M. Gupta, M. P. Darokar, Priyanka, and S. S. P. Khanuja, *Nat. Prod. Commun.* **3**, 891 (2008).
16. M. Fakruddin, K. M. A. Alam, R. M. Mazumdar, S. Islam, M. N. Nipa, A. Iqbal, and H. R. Bhuiyan, *J. Sci. Res.* **3**, 129 (2011). <http://dx.doi.org/10.3329/jsr.v3i1.6094>
17. S. Jain, P. P. Yadav, V. Gill, N. Vasudeva, and N. Singla, *Phytochem. Rev.* **8**, 491 (2009).
<http://dx.doi.org/10.1007/s11101-009-9134-8>
18. S. Dwivedi, *J. Ethnopharmacol.* **114**, 114 (2007). <http://dx.doi.org/10.1016/j.jep.2007.08.003>
19. V. K. Tripathi and B. Singh, *Orient. J. Chem.* **12**, 1 (1996).
20. R. Hossain, R. Sultana, A. Adhikari, M.I. Choudhary, Y. Ali, and S. Zaman, *Nat. Prod. Commun.* **9**, 371 (2014).
21. W. Wang, Z. Ali, Y. Shen, X-C. Li, and I. A. Khan, *Fitoterapia*, **81**, 480 (2010).
[doi:10.1016/j.fitote.2010.01.006](http://dx.doi.org/10.1016/j.fitote.2010.01.006)
22. W. Wei, Z. Ali, X-C. Li, Y. Shen, and I. A. Khan, *Planta Med.* **76**, 1751 (2010).
<http://dx.doi.org/10.1055/s-0030-1249809>
23. W. Wang, Z. Ali, X-C. Li, Y. Shen, and I. A. Khan, *Planta Med.* **76**, 903 (2010).
<http://dx.doi.org/10.1055/s-0029-1240841>
24. R.N. Yadava and K. Rathore, *J. Asian Nat. Prod. Res.* **2**, 97 (2002).
<http://dx.doi.org/10.1080/10286020008039898>
25. C. A. Menéndez, F. Nador, G. Radivoy, and D. C. Gerbino, *Org. Lett.* **16**, 2846 (2014).
dx.doi.org/10.1021/ol500964e.
26. N. M. A. Mahfouz, H. Hambloch, N. M. Omar, A. W. Frahm, *Archiv der Pharmazie in (Weinheim, Germany)*, 1990) **323(3)**, pp. 163-169.
27. M. Khatun, M. Billah, and M. A. Quader. *Dhaka Univ. J. Sci.* **60**, 5, (2012).
28. F. M. Moghaddam, M. M. Farimani, S. Salahvarzi, and G. Amin, *eCAM*, **4**, 95 (2007).
[doi: 10.1093/ecam/nel065](http://dx.doi.org/10.1093/ecam/nel065)
29. M. Arora and A.N. Kalia, *Int. J. Pharm. Sci.* **5**, 245 (2013).
30. O. Purev, K. Oyun, G. Odontuya, A. M. Tankhaeva, G. G. Nikolaeva, K. M. Khan, S. T. A. Shah, and W. Voelter, *Z. Naturforsch.* **57b**, 331 (2002).
31. Atta-Ur-Rahman, A. B. Reitz, M. I. Choudhary, *Frontiers of Medicinal Chemistry (Bentham Scientific Publishers Ltd., 2002)* **4**, pp. 23-85.