

Binding Studies and Effect of Light on the Conductance of Intercalated Curcumin into DNA

M. A. Subhan^{*}, M. M. Islam, and M. R. U. Chowdhury

Department of Chemistry, Shah Jalal University of Science and Technology, Sylhet, Bangladesh.

Received 25 August 2011, accepted in final revised form 16 February 2012

Abstract

Curcumin is one of the most valuable ingredients for scientific research in view of medicinal and therapeutic importance. Curcumin is reported to have antitumor and anticancer activities. It can easily be extracted from Turmeric by solvent extraction methods. At first, dye was extracted through reflux and trituration process. Then curcumin was isolated from dye using column chromatography, which was investigated by infrared spectroscopy and melting point. Curcumin formed a complex with Ce^{3+} , $Ce(C_{21}H_{20}O_6)_n$, which was confirmed by comparing the IR spectrum of complex with that of ligand. We observed the effect of light on the curcumin intercalated into DNA and found that the electrons in curcumin excited by absorption of light and the conductance of the solution increased gradually when irradiated by UV-light. The result may indicate the electron conducting property of DNA in solution. We focused on the binding of curcumin with DNA. Our investigations also showed that both curcumin and Ce^{3+} complex have luminescence properties upon irradiation by UV-light.

Keywords: Curcumin; Ce^{3+} -Complex; Luminescence; DNA; Antioxidant.

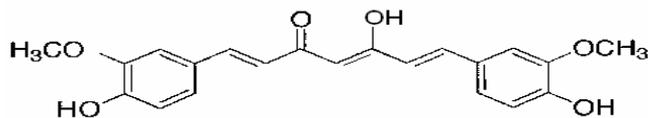
© 2012 JSR Publications. ISSN: 2070-0237 (Print); 2070-0245 (Online). All rights reserved.
doi: <http://dx.doi.org/10.3329/jsr.v4i2.8482> J. Sci. Res. 4 (2), 411-418 (2012)

1. Introduction

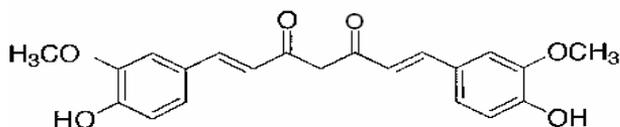
Turmeric is a member of the curcuma botanical group, which is a part of the ginger family of herbs, the *zingiberaceae*. The root and rhizome (underground stem) of the curcuma longa plant is crushed and powdered into ground turmeric. Ground Turmeric is used as a source of curcumin for its therapeutic effect. Turmeric is an ancient spice and a traditional remedy that has been used as medicine, condiment and flavoring. Turmeric contains a variety of bioactive substances called curcumoids. The major curcumoids are curcumin, demethoxycurcumin and bis-demethoxycurcumin [1]. The most active component is curcumin. These substances comprise 3 to 6% of curcuma longa. Curcumin makes up 70 to 75% of the curcuminoids, demethoxycurcumin 15 to 20% and bis-demethoxycurcumin about 3% [2].

^{*} Corresponding author: subhan_che@yahoo.com

Curcuminoids are obtained from turmeric by column (Silica gel, 20-270 mesh) extraction with dichloromethane. Curcumin is the most studied of the curcuminoids. In its pure form, it is an orange-yellow crystalline powder that is insoluble in water. Its chemical name is (E, E)-1,7-bis (4-hydroxy-3-methoxyphenol)-1,6-heptadiene-3,5-dione. The structure of the curcumin was elucidated in 1910 and it was first known as diarylheptanoid [3]. The structures of curcumin in enol and keto forms are shown as:



Enol form



Keto form

^{13}C NMR spectral data show that curcumin exists primarily in the enol form and not as the diketone. Curcumin exhibits a wide range of biological activities, e.g. anti-bacterial, anti-inflammatory, hypolipidemic, hepatoprotective, lipoxygenase, cyclooxygenase, protease inhibitory effects, besides being effective active oxygen species scavengers and lipid peroxidase (a class of oxidoreductase enzymes) inhibitors [4, 5]. Curcumin and the curcuminoids also lower cholesterol, reduce platelet aggregation, inhibit proliferation of cancer cells and improve digestion by increasing the flow of bile from the gallbladder [6]. Extracts of *curcuma longa* rhizomes exhibit good preventive activity against carbon tetrachloride induced liver injury *in vivo* and *in vitro*. Curcumin inhibits intestinal gas formation by *Clostridium perfringens* at 0.05% concentration [7]. Its effect was evaluated at 0.005, 0.013, 0.025 and 0.05% on gas formation by *C. perfringens* of intestinal origin. Gas formation decreased gradually as the curcumin concentration increased and there was no gas when curcumin concentration was 0.05%, the level at which bacterial growth was inhibited completely. Oral administration of curcumin and curcuminoids (750 mg/kg) has been reported to prevent the formation and dissolution of urinary calculi. Turmeric powder extracts and curcumin also exhibit antioxidant property [8, 9].

Self-assembled dye-DNA network and its photo induced electrical conductivity have been investigated [10]. Design of successive ion conduction path in DNA films with ionic liquids has also been studied [11]. In our work we attempted to investigate the dye-DNA network and its photo induced electrical conductivity, because electrons in curcumin may be excited by absorption of UV-light and may influence the conductance of the system.

There is a wide range of applications of fluorescence in the field of biochemistry and medicine. Large biological molecules can have a fluorescent chemical group attached by a chemical reaction, and the fluorescence characteristic of the attached tag enables very sensitive detection of the molecule. Aquarian, from the jellyfish *Aequorea victoria*, produces a blue glow in the presence of Ca^{2+} ions [12]. It has been used to image calcium flow in cells in real time. The success with aequorin spurred further investigation of *A. victoria* and led to the discovery of Green Fluorescent Protein (GFP), which has become an extremely important research tool. GFP and related proteins are used as reporters for many biological events including sub-cellular localization. Levels of gene expression are sometimes measured by linking a gene for GFP production to another gene. Binding and cleavage of DNA is at the heart of cellular transcription and translation, it is an obvious goal for therapeutic intervention and the development of diagnostic structural probes [13, 14]. In this paper, we report on the electrical properties, binding studies of intercalated curcumin into DNA and also the synthesis, characterization and luminescent properties of cerium-curcumin complex, $\text{Ce}(\text{C}_{21}\text{H}_{20}\text{O}_6)_n$.

2. Experimental

2.1. Dye extraction

Ground turmeric (40 g) was crushed and mixed with dichloromethane (100 mL) in a conical flask, which was stirred with a magnetic stirrer on a hot plate. It was then transferred into a 500 mL round-bottom flask and heated to reflux for 2 h. The mixture was filtered and the filtrate was concentrated in a hot water bath at 50°C. The reddish-yellow oily residue was triturated with hexane (30 mL). The resulting solid was collected by filtration. TLC analysis was carried out with preparative TLC plates coated with silica gel by using a solvent mixture (3% methanol: 97% dichloromethane), which showed the presence of three components.

2.2. Isolation of curcumin by column chromatography

The crude material obtained after trituraion with hexane was dissolved in a minimum amount of 99% dichloromethane and 1% methanol (v/v) mixture and loaded on to a column packed with silica gel (30 g). The column was eluted with the same solvent. Elutents were taken into various conical flasks successively, 5 mL in each case. Then TLC analysis was carried out for each collection. Some elute which was collected at the beginning of the elution process showed the presence of only one component. Other collections showed the presence of two components. The elutants, which showed the presence of only one component in TLC analysis, were collected and transferred into a 100 mL round-bottom flask and then evaporated in a rotatory evaporator to dryness. The melting point of the solid sample was found to be 165-169°C.

2.3 IR analysis

The IR spectrum of the isolated and dried curcumin ($C_{21}H_{20}O_6$) was recorded by SHIMADZU, IP Prestige-21, and FTIR Spectrophotometer with KBr pellet.

2.4. Synthesis of Ce ($C_{21}H_{20}O_6$)_n complex

0.001 M solution of $Ce(NO_3)_3$ was prepared by dissolving 0.04342 g into 100 mL of absolute ethanol in a beaker. 0.003 M solution of curcumin was prepared by dissolving 0.04416 g into 100 mL of absolute ethanol in a separate beaker. Then curcumin solution was added to metal solution slowly with stirring. Orange color precipitate formed was separated by filtration and air-dried. The IR spectrum of the complex was recorded.

3. Results and Discussions

3.1. IR study of curcumin and Ce ($C_{21}H_{20}O_6$)_n

Curcumin can form strong chelate with metal ions[15]. There have been recent studies on metal complexes of curcumin[16]. We have synthesized $Ce(C_{21}H_{20}O_6)_n$ and characterized by IR spectra. IR spectrum of curcumin, $C_{21}H_{20}O_6$ showed peaks at 751 cm^{-1} , which was due to C-H deformation of disubstituted (ortho) aromatic ring; at 805 cm^{-1} for C-H deformation of para disubstituted aromatic ring; at 958 cm^{-1} for C-H deformation of disubstituted (trans) alkene; peaks at 1072 cm^{-1} for C-O stretching of secondary alcohol; at 1122 cm^{-1} for C-O stretching of ether; at 1278 cm^{-1} for C-O stretching of C=C-O-C group; at 1200 cm^{-1} for phenolic C-O stretching and 1457 cm^{-1} for alcohol C-O-H bending; at 1656 cm^{-1} for conjugation of double bond C=C-C stretching and 1782 cm^{-1} for unsaturated ketone; at 2940 cm^{-1} for C-H stretching in CH_2 ; at 2969 cm^{-1} for Ar-H stretching, at 2862 cm^{-1} for C-H stretching of C-O- CH_3 group. For the complex, $Ce(C_{21}H_{20}O_6)_n$ the IR peaks observed were at 805 cm^{-1} , 1026 cm^{-1} , 1095 cm^{-1} , 1278 cm^{-1} , 1734 cm^{-1} , 2343 cm^{-1} , 2358 cm^{-1} , 2862 cm^{-1} , 2923 cm^{-1} , 2970 cm^{-1} and 3740 cm^{-1} . IR showed characteristic peaks for the curcumin ligand and $Ce(C_{21}H_{20}O_6)_n$ complex also showed similar IR spectra with shift in some peaks. A clear observation of the spectral changes in finger print region of $Ce(C_{21}H_{20}O_6)_n$ complex compared to that of curcumin ligand also indicates the formation of Ce^{3+} -complex.

3.2. Effect of light on conductance of intercalated curcumin into DNA

DNA was extracted from brinjal (*Solanum Melongena*) by solvent extraction method [17]. The intercalated curcumin into DNA was irradiated either by mercury lamp or tungsten filament bulb at room temperature to study the effect of light. A 30 mL solution of DNA was taken in a beaker and conductance was measured to be 0.018. Curcumin solutions

were then added, 1 ml each time. After addition of 3 mL of curcumin solution, the solution was irradiated with UV-visible light. Results are shown in Figs. 1 and 2.

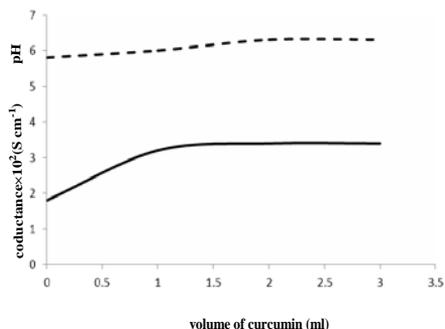


Fig. 1. Effect on conductance (—) and pH (·····) of the DNA solution due to the addition of 3 mL curcumin solution.

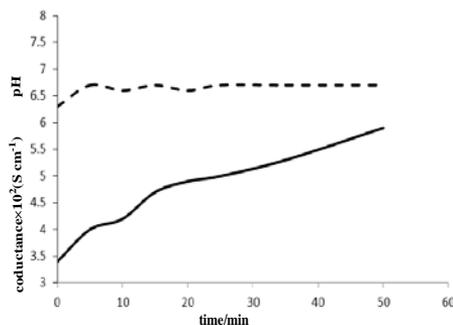


Fig. 2. Effect of light on the conductance (—) and pH (·····) of the solution of curcumin intercalated into DNA.

With the addition of curcumin into DNA solution up to 3 mL, no or little change in pH and/or conductance were observed at around neutral pH values as shown in Fig. 1. While irradiation took place, an increase in conductance was observed with little or no change in pH values of the solution as shown in Fig. 2.

3.3. DNA–curcumin binding studies

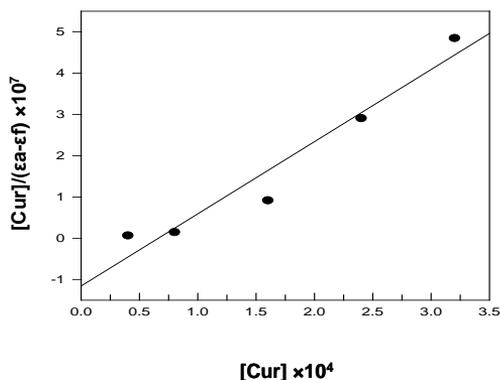
DNA was extracted [17] from brinjal for the binding studies with curcumin. DNA contains four kinds of smaller building blocks or monomers called deoxyribotides or deoxyribonucleotides [18]. Hundreds of thousand of nucleotides are hooked to form a chain, and two chains are paired together and twisted into a double helix to form the finished DNA molecule. The interaction of the DNA and curcumin can be examined from the UV-Vis spectra (UV-Vis spectrophotometer, Model: UV-9100, China) of curcumin-DNA intercalated complexes with the successive addition of curcumin. By monitoring the change in absorbance with increasing concentration of curcumin, the intrinsic binding constant can be evaluated by Eq. (1) [19, 20]. The intrinsic binding constant, K_b was determined from the plot of $[\text{Cur}]/(\epsilon_a - \epsilon_f)$ vs $[\text{Cur}]$ according to the equation (1) where $[\text{Cur}]$ is the concentration of curcumin, ϵ_a , the apparent extinction coefficient is obtained by calculating $A_{\text{obsd}}/[\text{DNA}]$, ϵ_f corresponds to the extinction coefficient of the DNA in its free form and ϵ_b refers to the extinction coefficient of the curcumin-DNA complex in the fully bound form.

$$[\text{Cur}]/(\epsilon_a - \epsilon_f) = [\text{Cur}]/(\epsilon_b - \epsilon_f) + 1/K_b(\epsilon_b - \epsilon_f) \quad (1)$$

Each set of data, gave a straight line with a slope of $1/(\epsilon_b - \epsilon_f)$ and a y -intercept of $1/K_b(\epsilon_b - \epsilon_f)$. K_b was determined from the ratio of the slope to intercept. The absorbance of [DNA] solution was 0.080 and molar extinction coefficient of DNA solution, ϵ was 6600. According to Beer-Lambert law, $C = A/\epsilon$, and the initial concentration of DNA solution was [DNA], 1.212×10^{-5} . By using Table 1 and Fig. 3 the intrinsic binding constant of DNA-curcumin complex, K_b , was found to be 1.51.

Table 1. Binding studies of curcumin with DNA.

Conc. of curcumin solution, [Cur]	Observed absorbance of DNA-curcumin solu. at 420 nm (A_{obs})	Apparent extinction coeff. (ϵ_a) = $A_{obs}/[DNA]$	Extinction coefficient of the DNA in its free form (ϵ_f)	$ (\epsilon_a - \epsilon_f) $	$\frac{[Cur]}{ \epsilon_a - \epsilon_f }$
3.2×10^{-4}	0.072	5940.60		695.41	4.85×10^{-7}
2.4×10^{-4}	0.070	5775.57		824.43	2.91×10^{-7}
1.6×10^{-4}	0.059	4860.87	6600	1739.13	0.92×10^{-7}
0.8×10^{-4}	0.019	1567.65		5032.35	0.15×10^{-7}
0.4×10^{-4}	0.010	825.08		5774.9	0.069×10^{-7}

Fig. 3. A plot of $[Cur]/(\epsilon_a - \epsilon_f) \times 10^7$ vs $[Cur] \times 10^4$ for curcumin intercalated with DNA.

3.4. Curcumin: a potent complexing agent for metals

The antioxidant activity of curcumin derivatives are greater than that of vitamin E, and particularly identical to that of curcumin in a Fe(II)-linoleic acid peroxidation test. Curcumin is a very interesting substance because it generates phototoxic oxidizing species including $HO\cdot$ and H_2O_2 , when exposed to light, but it also protects against lipid peroxidation as radical scavenger [21]. This compound is a potent complexing agent for

metal ions such as iron (III) [21], it can act as phototoxicant and protective agent. Curcumin, like other β -diketones, exists partly in the enol form, which may also help to explain many of its unusual anti- and prooxidant properties [21]. Curcumin forms a complex with cerium, $\text{Ce}(\text{C}_{21}\text{H}_{20}\text{O}_6)_n$, whose structure was confirmed by comparing the IR spectrum of both complex and ligand in solid form. Curcumin is a β -diketone, can act as uninegative bidentate ligand. Coordination number of Ce^{3+} would be eight, but for the formation of neutral complex and also because of the bulky substituent in the curcumin, in the complex, $\text{Ce}(\text{C}_{21}\text{H}_{20}\text{O}_6)_n$, $n = 3$ might be suitable for complexation.

3.5. Luminescent properties of curcumin and its metal complex

When irradiated with UV-light, a mercury lamp at 380 nm both curcumin (orange) and $\text{Ce}(\text{C}_{21}\text{H}_{20}\text{O}_6)_n$ (deep orange) showed green fluorescence emission in CH_2Cl_2 solution at room temperature, which is disappeared when removed from light source. Both curcumin and Ce^{3+} complex had luminescence properties. It was clearly observed that the fluorescence is greatly enhanced when Ce^{3+} formed a complex with curcumin.

4. Conclusion

Yellow dye was extracted from dried turmeric herb and curcumin was separated from this dye and characterized by IR and UV-visible spectra. Complex of curcumin with cerium, Ce^{3+} may be used for medicinal purpose. Intercalated curcumin into DNA showed conductivity when irradiated either by mercury lamp or tungsten filament bulb at room temperature. This may be due to the conducting DNA-curcumin solution. Binding studies of curcumin with DNA was performed to calculate binding constant, K_b , which was found to be 1.51. This might be an important parameter for the application of DNA-curcumin intercalated compound in medicine. The cerium-curcumin complex has enhanced luminescence; showed green luminescence upon UV-irradiation in solution at room temperature.

References

1. S. Gilda, M. Kanitkar, R. Bhonde, and A. Paradkar, *LWT- Food Sci. Technol.* **43**, 59 (2010).
2. T. J. Zachariah, Indian Spices and Exploration of Its Intrinsic Quality: http://www.biocircle-project.eu/media/7080/dr_john_z_calicut.pdf
3. A. M Anderson, M. S. Mitchell, and R.S. Mohan, *J. Chem. Edu.* **77**, 359 (2000). <http://dx.doi.org/10.1021/ed077p359>
4. V. P. Menon and A. R. Sudheer, *Adv. Exp. Med Biol.* **595**, 105 (2007). http://dx.doi.org/10.1007/978-0-387-46401-5_3
5. A. K. Tuba and I. Gülçin, *Chemo-Biological Interactions*, **174**, 27 (2008). <http://dx.doi.org/10.1016/j.cbi.2008.05.003>
6. R. Wilken, M. S. Veena, M. B. Wang, and E. S. Srivatsan, *Molecular Cancer* **10** (12), 1 (2011). PMID:21205300; PMCID:3024301
7. B. B. Aggarwal, Y-J. Surh, and S. Shishodia, *Advances in Experimental Medicine and Biology* (Springer, New York, **595**, 2007).

8. N. K. Khanna, *Current Sci.* **76**, 1351 (1999).
9. A. K. Nadkarni, *Indian Materia Medica* (Popular Prakashani, Bombay, 1976).
10. G. Jianhu, T. Shinnichi, O. Youchi, T. Hitoshi, and K. Tomoji, *Appl. Phys. Lett.* **80**, 688 (2002). <http://dx.doi.org/10.1063/1.1435805>
11. N. Naomi and O. Hiroyuki, *J. Mater. Chem.* **12**, 2299 (2002).
<http://dx.doi.org/10.1039/b202972c>
12. F. Prendergast and K. Mann, *Biochemistry* **17**, 3448 (1978).
<http://dx.doi.org/10.1021/bi00610a004>
13. R. Tsien, *Annu.Rev. Biochem.* **67**, 509 (1998).
<http://dx.doi.org/10.1146/annurev.biochem.67.1.509>
14. L. J. Boerner and J. M. Zaleski, *Curr. Opin. Chem. Biol.* **9**, 135 (2005).
<http://dx.doi.org/10.1016/j.cbpa.2005.02.010>
15. M. Borsari, E. Ferrari, and R. Grandi, *Inorg. Chim. Acta*, **328**, 61 (2002).
[http://dx.doi.org/10.1016/S0020-1693\(01\)00687-9](http://dx.doi.org/10.1016/S0020-1693(01)00687-9)
16. Y. M. Song, J. P. Xu, L. Ding, Q. Hou, J. W. Liu, and Z. L. Zhu, *J. Inorg. Biochem.* **103**, 396 (2009). <http://dx.doi.org/10.1016/j.jinorgbio.2008.12.001>
17. M. A. Subhan, M. K. H. Rashed, B. Ahmed, and M. R. U. Chowdhury, *Proc. Pakistan Acad. Sci.* **45**, 171 (2008).
18. P. S. Verma and V. K. Aggarwal, *Cell Biology, Genetics, Molecular Biology, Evolution as well as Ecology*, 4th Edition (S. Chand & Company Ltd., New Delhi, 2005).
19. H. L.Chan, H. Q. Liu, B. C. Tzeng, Y.S. You, S.M. Peng, M. Yang, and C. M. Che, *Inorg. Chem.* **41**, 161 (2002). <http://dx.doi.org/10.1021/ic0112802>
20. S. Arounaguirri, D. Easwaramoorthy, A. Ashokkumar, A. Dattagupta, and B. G. Maiya, *Proc. Indian Acad. Sci.* **112**, 1 (2000). <http://dx.doi.org/10.1007/BF02704295>
21. A. L. Richard, *Naturally occurring antioxidants* (Lewis Publishers, New York, 1997).