Investigation on Spectroscopic, Thermal and Antimicrobial Activity of Newly Synthesized Binuclear Cr(III) Metal Ion Complex

M. R. Islam, J. A. Shampa, M. Kudrat-E-Zahan[*], M. M. Haque, Y. Reza

Department of Chemistry, University of Rajshahi, Rajshahi-6205, Bangladesh

Abstract

Binuclear Cr(III) metal ion complex was synthesized using diphenylacetic acid as primary ligand and 2-methyl pyridine (2-picoline) as secondary ligand. The synthesized complex was characterized by conductivity, FTIR, UV-Vis spectroscopy, magnetic moment and thermogravimetric analysis (TGA). FTIR spectra indicated the coordination of deprotonated diphenylacetic acid and 2-picoline through nitrogen and oxygen. The presence of water molecules inside the coordination sphere of the complex was confirmed from IR spectrum and TGA analysis. Binuclear and octahedral structure of the Cr(III) complex confirmed by TGA, UV-Vis spectroscopy and magnetic moment measurement. The complex showed moderate antibacterial activity with no antifungal activity.

Keywords: Mixed ligand; Binuclear complex; Cr(III); TGA analysis.

1. Introduction

Cr(III) complexes have played an important historical role in the development of transition metal chemistry, primarily due to their relative kinetic inertness [1]. Cr(III) is an essential trace element for humans. Together with insulin, it removes glucose from the blood and plays a vital role in fat metabolism. Biological function of chromium is not fully known yet. The diabetes relevant interaction of Cr(III) is with the hormone insulin and its receptors. This suggests that Cr(III) acts with insulin on the first step in the metabolism of sugar entry into the cell, and facilitates the interaction of insulin with its receptor and the cell surface [2,3]. Chromium increases insulin binding to cells, insulin receptor number and activates insulin receptor kinase leading to increased insulin sensitivity [4]. Thus, chromium is an essential element

[*] *Corresponding author*: kudrat.chem@ru.ac.bd

involved in glucose metabolism and diabetes [5]. Chromium is also known to activate enzymes, maintain protein stability. Organic Cr(III) sources have been shown to enhance the availability of chromium [6]. However, long exposure to Cr(III) compounds may result in dermatitis and liver or kidney damage in humans and is thereby toxic in large doses [7,8].

The metal complexes with the ligand containing heterocyclic ring systems are very promising candidates for practical applications [9]. Synthesis, characterization and biological activities of mixed ligand complexes of some transition elements with Phthaliimides as primary and 2-picoline as secondary ligands have already been published [10,11]. Manimekala reported the synthesis and thermal stability of some mixed ligand complexes of transition metals with diphenylacetic acid and neutral hydrazine [12]. Reza *et al.* disclosed the crystal structure of mixed ligand copper complex formed with 2,2-bipyridine and diphenylacetic acid [13]. To the best of our knowledge, the research work on the mixed ligand complex of transition metals with diphenylacetic acid and 2-picoline is not reported yet.

Here, we have investigated the preparation and characterization of binuclear Cr(III) metal ion complex using diphenylacetic acid as primary ligand and 2-methyl pyridine (2-picoline) as secondary ligand. We also included the antibacterial and antifungal activity of the synthesized complex.

2. Experimental

2.1. *Instrumentation*

The weighing operation was performed on a METTLER PM 200 electronic balance. Infrared spectra were recorded on a FTIR-8400, SHIMADZU, Japan using a KBr disc, in Central Science Lab of Rajshahi University. The Sherwood Scientific susceptibility balance was used for magnetic susceptibility measurement. The melting point or decomposition temperature of the prepared metal complex was observed with the Tlectrothermal® Melting Point Apparatus. It was however, not possible to measure the melting points beyond 300°C. The thermogravimetric analysis (TGA) was performed on Perkin Elmer Simultaneous Thermal Analyzer, STA-8000. All conductivity readings were measured by Horiba B-173 compact conductivity meter. The electronic spectrum of the complex in solution phase (9.82×10^{-4} M) was recorded on a Shimadzu Double Beam spectrophotometer model UV-1200 and UV-1650PC.

2.2. *Synthesis of binuclear Cr(III) complex, [Cr$_2$(DiPhAc)$_2$(2-pic)$_2$(H$_2$O)$_5$]Cl$_2$*

2 mmol (0.436 g) of diphenylacetic acid was dissolved in absolute alcohol to which 2 mmol (0.187 g) 2-picoline was added. Then 2 mmol (0.202 g) triethylamine was added and the mixture was stirred for 10 min. After that an ethanolic solution of 2 mmol (0.574 g) of $CrCl_3 \cdot 6H_2O$ was added to the mixture while stirring with magnetic stirrer.

The stirring was continued for 6 h at 60°C and then cooled to room temperature. Thus, the ppt. formed was filtered, washed several times with alcohol and dried in a vacuum desiccator over anhydrous $CaCl_2$.

3. Results and Discussions

3.1. *Decomposition temperature/melting point*

Decomposition temperature/melting point give an approximate idea about the nature of the complexes and can suggest whether it is covalent or ionic. Also with comparing initial materials m.p./decomposition temperature can predict preliminary whether any reaction occurs or not. We found that the synthesized complex $[Cr_2(DiPhAc)_2(2-Pic)_2(H_2O)_5]Cl_2$ is having decomposition temperature above 300°C which indicated the formation of a stable coordination compound of Cr(III) [14,15].

3.2. *Magnetic moment*

The measurements of magnetic susceptibilities were made at constant temperature 304 K, Curie-law was used and was calculated from the equation. $\mu_{eff} = 2.828 \sqrt{\chi_m^{corr}.T}$ B.M. Thus μ_{eff} obtained is known as effective magnetic moment. The magnetic moment of this complex was determined on the solid as $\mu_{eff} = 3.79$ B.M., which corresponds to three unpaired electrons and indicate that the synthesized Cr(III) complex is paramagnetic and might be having octahedral geometry [16-18].

3.3. *Molar conductance*

The value of molar conductance in ohm^{-1} cm^2 mol^{-1} indicates the charge type of electrolyte. The molar conductance of 9.82×10^{-4} M solution of the complex in DMSO was measured at 29°C. The molar conductance values 115 ohm^{-1} cm^2 mol^{-1} indicated that the complex is good 1:1 electrolyte [19].

3.4. *Infrared spectra*

The spectrum of free diphenylacetic acid shows a broad absorption near 3568-3106 cm^{-1} which indicated the presence of υ(O-H) group. There was also a C-O single bond band at 1215 cm^{-1} and the υ(C=O) band at 1705 cm^{-1}. The spectra of free 2-picoline shows band at 1693 cm^{-1} assigned to υ(C=N) [20]. The complex displays bands at 1563 cm^{-1} and 1600 cm^{-1} due to υ_{sym}(C=O) and υ_{asy}(C-O) respectively, significantly lower than those of free diphenylacetic acid indicating the coordination of acid through its carboxylate anions. A broad band observe at 3200-3600 cm^{-1} is due to υ(O-H) for free acid. The disappearance of the υ(O-H) mode observe in the free acid molecule clearly indicate the loss of the protons from both O-H groups upon

coordination, revealing that acid are negative bidentate ligand coordinating through carboxylate anions. Several absorptions of similar intensity at 3068 cm^{-1} to 2950 cm^{-1} indicate the presence of two υ(M-O) in the complex [21] and an absorption near 3606 cm^{-1} indicate the presence of a bridged water ligand [22].

Further the presence of M-O bonding is evident from the appearance of υ(M-O) modes at 431-585 cm^{-1} in the spectra of the complex. The in-plane and out of plane ring deformation modes of the heterocyclic amines observe at ~700 cm^{-1} undergo a positive shift in mixed ligand complexes confirming thereby coordination through nitrogen. The presence of υ(M-N) bonding in the complex is also evident from appearance of υ(M-N) modes at 747 cm^{-1} in the spectrum of the complex.

3.5. *UV-Vis spectra*

The UV-Vis spectra of the complex [Cr$_2$(DiPhAc)$_2$(2-Pic)$_2$(H$_2$O)$_5$]Cl$_2$, CrCl$_3$·6H$_2$O, diphenylacetic acid and 2-methyl pyridine (2-picoline) are shown in Fig. 1. The ligand 2-picoline shows transition at 204 nm for the presence of (C=N) as $\pi\rightarrow\pi^*$ and transitions occurred at 256 nm to 270 nm for the presence of conjugated double bonds. Diphenylacetic acid shows transitions at, 260 nm and 266 nm for $n\rightarrow\sigma^*$ and $n\rightarrow\pi^*$, The assignment of the geometrical configuration of some Cr(III) complexes with mixed ligands can be suggested by inspection of the d−d absorption spectra [23,24]. The UV spectrum of CrCl$_3$.6H$_2$O in DMSO shows two d-d transition at 14970 cm^{-1} (668 nm) and 21322 cm^{-1}(469 nm). The reflectance spectra of Cr(III) complex consist of three transition bands at 260 nm, 410 nm and 574 nm assignable to the transitions $^4A_{2g}\rightarrow^4T_{2g}$, $^4A_{2g}\rightarrow^4T_{1g}(F)$ and $^4A_{2g}\rightarrow^4T_{1g}(P)$ respectively. These transitions of the complex have occurred at significantly higher energy state (lower wavelength). The spectrum of the complex is further supported by the magnetic moment of the complex and predicted the octahedral structure of the complex [Cr$_2$(DiPhAc)$_2$(2-Pic)$_2$(H$_2$O)$_5$]Cl$_2$ [25].
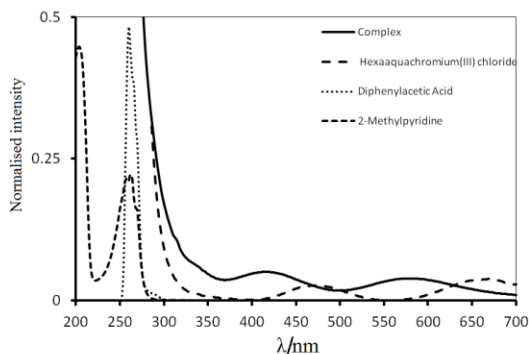
Fig. 1. UV spectra of complex [Cr$_2$(DiPhAc)$_2$(2-Pic)$_2$(H$_2$O)$_5$]Cl$_2$, CrCl$_3$.6H$_2$O, diphenylacetic acid and 2-methyl pyridine (2-picoline).

3.6. *Thermogravimetric analysis*

TGA was carried out for solid complex $[Cr_2(DiPhAc)_2(2\text{-Pic})_2(H_2O)_5]Cl_2$, under N_2 flow and heating rate was suitably controlled at 30°C min^{-1} and the weight loss was measured from the ambient temperature upto 800°C. The TGA curve of the complex is shown in Fig. 2. Tables 1-2 show the maximum temperature values $T_{max}/°C$, species lost together with the corresponding weight loss for each step of the decomposition reaction [26,27]. The data obtained strongly supported the proposed formulas of the complex. The data from the TGA clearly indicated that the decomposition of the complex proceeds in four or five steps. In the first step of decomposition two chlorine atoms were lost between 30°C to ~331°C (calculated 8.10%, experimental 7.75% of weight) [28,29]. Four molecules of coordinated water molecules are gradually decomposed between temperatures 332°C to 384.4°C (calculated 8.508%, experimental 8.85% of weight) in second step of decomposition [30]. Monodentate two 2-picoline ligands were decomposed in third step at 385°C to 658°C (calculated 21.29%, experimental 24.69% of weight). Bridging diphenylacetic acid and water molecules are to be decomposed as biphenyl ketone and hydrogen atom respectively at >650 °C in fourth or fifth stage of decomposition and Cr_2O_3 assumed to be formed at high temperature [31]. The proposed degradation pathway, supported by TGA curve and data in Fig. 3 showing order and steps of possible elimination of atoms and groups from the synthesized Cr(III) complex.

Table 1. Thermogravimetric data of synthesized complex.

Entry	Step	Temperature range (°C)	Weight loss found (calculated) (%)
$[Cr_2(DiPhAc)_2(2\text{-}pic)_2(H_2O)_5]Cl_2$	First	30-331	7.75 (8.10)
	Second	332-384.41	8.85 (8.508)
	Third	385-658	24.69 (21.29)
	Fourth	>650	(44.58)
	Fifth	>650	(0.23)

Table 2. The TGA data of the complex (thermal decomposition).

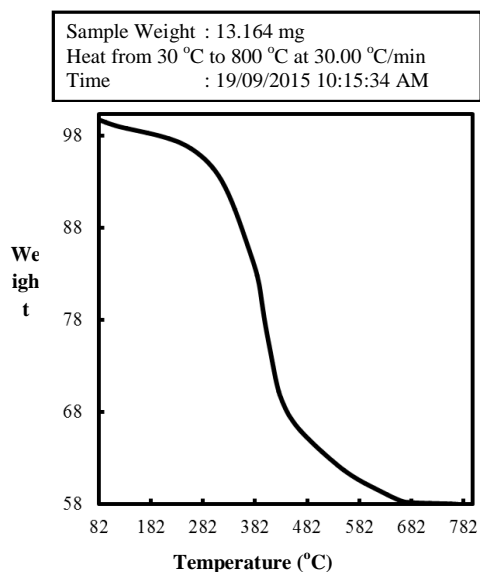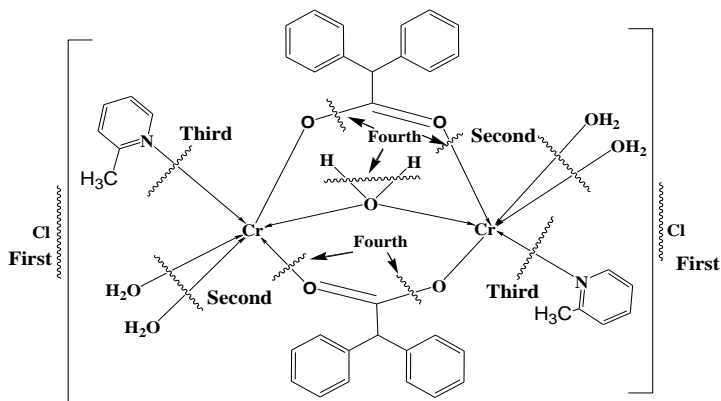| Temperature (°C)	100 & 150	200 & 250	300 & 350	400 & 450	500 & 550	600 & 650	700 & 750
Complex	Weight loss (%)						
$[Cr_2(DiPhAc)_2(2\text{-}pic)_2(H_2O)_5]Cl_2$	99.62 & 98.88	98.23 & 97.17	94.83 & 89.20	78.26 & 67.64	64.68 & 62.05	60.31 & 58.90	58.38 & 58.30

Fig. 2. TGA pattern of complex $[Cr_2(DiPhAc)_2(2\text{-pic})_2(H_2O)_5]Cl_2$.

Fig. 3. A possible degradation pathway of complex $[Cr_2(DiPhAc)_2(2\text{-Pic})_2(H_2O)_5]Cl_2$.

3.7. *Chloride (Cl⁻) test*

The reaction of the synthesized complex with $AgNO_3$ gave white precipitate, and this result showed presence of Chloride (Cl⁻) in outer coordination sphere. This result is also consistent with the molar conductance values 115 ohm^{-1} cm^2 mol^{-1}.

3.8. *Antimicrobial activity*

Any chemical or biological agent that either destroys or inhibits the growth of microorganisms is called antimicrobial agent. The susceptibility of microorganism to antimicrobial agent can be determined in vitro by a number of methods. The disc diffusion technique [32] is widely acceptable for preliminary investigations of materials which are suspected to possess antimicrobial properties. Diffusion procedure, as normally used in essentially a qualitative test, which allocates organism of the susceptible, intermediate (moderately susceptible) or resistant categories. The antibacterial activities of the test complexes were determined by using the dose of 20 µg/disc. The complexes showed strong sensitivity against both gram positive and gram negative bacteria and the results were compared with antibiotic disc of Streptomycin. The synthesized Cr(III) complex showed antibacterial activity in zone of inhibition diameter as 13 mm, 12 mm, and 12 mm against Bacillus subtilis, Escherichia coli and Proteus volgaris respectively. The complex hasn't shown any antifungal activity against Fusarium filamentous fungi.

4. Conclusion

Magnetic moment data indicated that the complex is paramagnetic in nature having three unpaired electrons. Conductivity measurement indicated that the complex is good 1:1 electrolyte in nature. IR spectral data showed the ligands coordinate with metal atom through O and N atoms. TGA showed the thermal stability of complex and the data obtained from TGA strongly supported the proposed formula of the complex [33,34] . UV-Vis spectra of the synthesized Cr(III) complex confirmed the formation and octahedral structure of synthesized $[Cr_2(DiPhAc)_2(2\text{-Pic})_2(H_2O)_5]Cl_2$ complex where two molecules of diphenylacetic acid and one molecule of water coordinated through μ_2 bridging. Based on these facts a structure of complex $[Cr_2(DiPhAc)_2(2\text{-Pic})_2(H_2O)_5]Cl_2$ has been proposed as shown in Fig. 4.
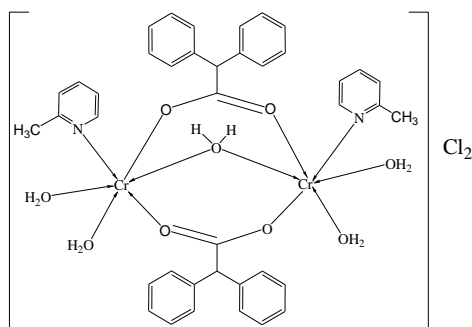

Fig. 4. Proposed structure of the synthesized $[Cr_2(DiPhAc)_2(2\text{-Pic})_2(H_2O)_3]Cl_2$.

References

1. F. A. Cotton and G. Wilkinson, Advanced Inorganic Chemistry, 5th edition (Wiley, New York, 1988).
2. Z. Krejpcio, Polish J. Environ. Studies **10(6)**, 399 (2001).
3. J. B. Vincent, A History of Chromium Studies (1955−1995), The Nutritional Biochemistry of Chromium(III) (Elsevier, New York, 2007) .
4. R.A. Anderson, Chromium in the prevention and control of diabetes. Diabetes and Metabolism, **26**, 22-27 (2000).
5. A. R. Kumar A, P. Riyazuddin, Microchem. J. **93**, 236 (2009).
6. J. B. Vincent, Acc. Chem. Res. **33**, 503 (2000). http://dx.doi.org/10.1021/ar990073r
7. J. E. Wahlberg and G. Wennersten, Br. J. Dermatol. **97**, 411 (1977). http://dx.doi.org/10.1111/j.1365-2133.1977.tb14250.x
8. V. Murphy, S. A. M. Tofail, H. Hughes, and P. McLoughlin, Chem. Eng. J. **148**, 425 (2009). http://dx.doi.org/10.1016/j.cej.2008.09.029
9. M. A. Subhan, F. Ahmed, M. S. Rahaman, A. K. Azad, and K. Begum, J. Sci. Res. **7(3)**,113 (2015). http://dx.doi.org/10.3329/jsr.v7i3.23270
10. L. A. Banu, M. S. Islam, M. A. A. Al-Bari and M. Kudrat-E-Zahan, Int. J. Adv. Multi. Res. **2(1)**, 145 (2015).
11. M. Kudrat-E-Zahan, M. M. Haque, L. Ahmmed, M. S. Ali, and M. S. Islam, Int. J. Mater. Sci. Appl. **4(2)**, 120 (2015).
12. R. Manimekalai, Int. J. Appl. Biol. Pharmaceut. Technol. **2(3)**, 268 (2011).
13. M. Y. Reza, L. A. Banu, M. S. Islam, S. W. Ng, and E. R. T. Tiekink, Acta Cryst. **E67**, m399 (2011). http://dx.doi.org/10.1107/S160053681100729X
14. M. Kudrat-E-Zahan, M. A. Bashar, M. F. Hossen, and M. S. Islam, AJRC **8(2)**, 74 (2015).
15. H. A. Bayoumi, A-N. M.A. Alaghaz, and M. S. Aljahdali, Int. J. Electrochem. Sci. **8**, 9399 (2013).
16. D. L. Maples, R. D. Maples, W. A. Hoffert, T. H. Parsell, A. V. Asselt, J. D. Silversides, S. J. Archibald, and T. J. Hubin, Inorganica Chimica Acta **362**, 2084 (2009). http://dx.doi.org/10.1016/j.ica.2008.09.034
17. F. A. Cotton and G. Wilkinson, Advanced Inorganic Chemistry, 3rd edition (John Wiley and Sons, Canada, 1972) 838.
18. R.L. Carlin, Magnetochemistry (Springer-Verlag, Berlin, 1986). http://dx.doi.org/10.1007/978-3-642-70733-9
19. M. B. Hossain, M. S. Islam, M. R. Islam, M. A. Salamd, and M. A. Yousuf, J. Bangl. Chem. Soc. **25(2)**, 139 (2012).
20. N. N. Greenwood and K. Wade, J. Chem. Soc. **232**, 1130 (1960). http://dx.doi.org/10.1039/jr9600001130
21. B. Šopotrajanov, V. Stefov, M. Žugić, V. M. Petruševski, J. Mol. Struc. **482**,109 (1999).
22. J. R. Carney, A. V. Fedorov, J. R. Cable, and T. S. Zwier, J. Phys. Chem. A **105**, 3487 (2001). http://dx.doi.org/10.1021/jp003375f
23. J. -H. Choi, M. A. Subhan, and S. W. Ng, J. Inorg. Gen. Chem. **638(2)**, 433 (2012).
24. S. A. Sadeek, M. S. El-Attar, and N. S. Abd El-Lattif, Bull. Chem. Soc. Ethiop. **28(1)**, 53 (2014). http://dx.doi.org/10.4314/bcse.v28i1.7
25. A. Debnath, F. Hussain, and D. T. Masram, Bioinorg. Chem. Appl. Article ID 457478, (2014).
26. D. Gürbüz, A. Çinarli, A. Tavman, and A. S. B. Tan, Bull. Chem. Soc. Ethiop. **29(1)**, 63 (2015). http://dx.doi.org/10.4314/bcse.v29i1.6
27. V. K. Sharma and S. Srivastava, Turk. J. Chem. **30**, 755 (2006).
28. M. Shabbir, Z. Akhter, A. Gul, and M. Bolte, J. Chem. Soc. Pak. **36(1)**, 56 (2014).
29. S. M. El-Megharbel, J. Microb. Biochem. Technol. **7(2)**, 65 (2015).
30. A. W. Bauer, D. M. Perry, and W. M. M. Kirby, A. M. A. Arch. Intern. Med. **104**, 208 (1959). http://dx.doi.org/10.1001/archinte.1959.00270080034004

31. M. S. Masoud, S. A. A. EI-Enein, and H. M. Kamel, Ind. J. Chem. **41A**, 297 (2002).
32. A. Sharma, T, Mehta, and M. K. Shah, Der Chemica Sinica **4(1)**, 141 (2013).