

Available Online

JOURNAL OF SCIENTIFIC RESEARCH www.banglajol.info/index.php/JSR

J. Sci. Res. **11** (1), 83-88 (2019)

Lethal Effects of Toxic Plants (Neem, Datura, Acanthus) on Artemia Species

T. A. Jihad, M. S. Bhuyan*, M. S. Islam

Institute of Marine Sciences and Fisheries, University of Chittagong, Chittagong, Bangladesh

Received 17 April 2018, accepted in final revised form 14 October 2018

Abstract

Effects of the water-soluble extracts (dry leaf powder of *Azadirachta indica, Datura stramonium* and *Acanthus ilicifolius*) were studied on the behavioral pattern and mortality of *Artemia* species. The extracts of all the plants brought significant change in the behavioral pattern of *Artemia*. The *Artemia* become agitated just offer exposure to the extract and started to move upwards and downwards vigorously. The *Artemia* exposed to the various concentration of water-soluble plant extracts were unable to maintain dorsoventral position and dropped to the bottom finally. The mortality rates recorded after 24 h exposure to the extract *Azadirachta indica* were 30% at 1500 mg/L and 70% at 3000 mg/L. Similarly, after 24 h exposure to the extracts of *Datura stremonium*, the mortality rates were 40% at 2000 mg/L and 75% at 3500 mg/L. Again, after 24 h has exposure to the extracts of *Acanthus ilicifolius*, the mortality rates were 35% at 2500 mg/L and rose to 80% at 4000 mg/L. The 24 h LC₅₀ values of the water-soluble extracts of Neem, Datura and Acanthus were 2331.930, 2357.948 and 2922.957. *Artemia* was more sensitive to the water-soluble extract of Neem than Datura and Acanthus.

Keywords: Lethal effects; Toxic plants; Mortality; Behavioral pattern; Artemia.

© 2019 JSR Publications. ISSN: 2070-0237 (Print); 2070-0245 (Online). All rights reserved. doi: <u>http://dx.doi.org/10.3329/jsr.v11i1.36424</u> J. Sci. Res. **11** (1), 83-88 (2019)

1. Introduction

Artemia is one kind of zooplankton popularly known as brine shrimp [1]. It is widely distributed on the five continents in hyper saline biotopes including salt lakes coastal or inland waters rich in chloride, sulphate or carbonate and especially in coastal salinas [2]. It is about 12 mm in length. *Artemia* is a high saline organism (20-300 ppt) [3,4]. They can survive short periods of time in the fresh water but cannot reproduce. The optimum temperature is 20 to 55 °C for *Artemia* to survive. They have two modes of reproduction [5]. *Artemia* life cycle begins by the hatching of dormant cysts, which are encased embryos that are metabolically inactive. The cysts can remain dormant for many years as long as they are kept in dry and oxygen-free condition [6]. Surgeloss [7] reported that *Artemia* has a very efficient ecological defense mechanism. It has great practical value

^{*} Corresponding author: <u>simulbhuyan@gmail.com</u>

because of its availability as day cysts. However, since the demand perennially treats, it is always in short supply and thus very expensive.

Artemia used as feed in the hatchery and they can be used as feed mixed with artificial diets as a source of protein as an attractant [8]. Artemia is an excellent food for newly hatched fish [9] and shrimp larvae [10]. The nauplii of Artemia are readily taken by aquatic animals in nursery stage.

There are many kinds of toxic plants in the natural environment [11]. These toxic plants have a great impact on the brine shrimp *Artemia* [12]. Toxicity of plants to plants to plants fauna (*Artemia salina*) is different from plants to plants [13]. In the present experiment *Azadirachta indica* (Neem), *Datura stremonium* (Datura) and *Acanthus ilicifolius* (Acanthus) are used as toxic plants to assess the extent of toxicity of these plants on *Artemia*.

2. Materials and Methods

The experiment was conducted in the laboratory of the Institute of Marine Sciences and Fisheries, the University of Chittagong from May to August 2016. The fresh leaves of Neem, Datura, and Acanthus were collected from the south campus of the area of Chittagong University and Coastal area of Cox's Bazar. The collected leaves of the plants were dried at room temperature and then heated in an oven at (50-60)°C until a constant weight of the samples were reached. The dried samples were powdered with the help of a powder driven grinder.

Three water-soluble extracts were prepared from the dried powdered leaves of Neem, Datura, and Acanthus. For the preparation of a stock solution, the dried powders were mixed thoroughly with distilled water at a ratio 1:10, that means one gram powder was mixed with 10 mL of distilled water. The mixture was left at room temperature for 24 h. The mixture was then filtered with fine mesh clothing. Thus a crude extract was obtained. It was collected in a conical flask and stored in a refrigerator at 4 °C as a stock solution.

2.1. Saline water preparation

Concentrated brine was collected from the salt bed of Chakaria coast, Cox's Bazar. The brine was mixed with tap water. 1 kg of mud was mixed with ten liters of the tap water in an aquarium for the better adaptation. Gradually, 5 kg mud was mixed with 50 L of the tap water.

2.2. Experimental setting

Three aquaria of equal size $(30'' \times 18'' \times 16'')$ washed with tap water were set up in a ventilated and damp proof room. Necessary equipment and chemicals were collected and set in a laboratory for measuring the water parameters of the aquaria. Each saline water provided with a mechanical aerator. Experimental setup for the toxicity test with different plants is shown in Fig. 1.



Fig. 1. Experimental setting.

2.3. Artemia collection and acclimatization

Artemia was collected from local market of Cox's Bazar [14]. After collection, *Artemia* was put in a plastic bowl. Then a jar was taken and three liters of saline water (20 ppt) was put in the jar. Then *Artemia* cysts were poured in the jar. Then an aerator was linked in the jar for aeration and waited 2 days for hatching. After hatching, the cysts of *Artemia* was poured in the aquarium containing saline water (20 ppt). Gradually after hatching, *Artemia* started to grow large [15]. Then egg sperm was given as a feed for newly hatched *Artemia*. Salinity, temperature, pH were measured with the help of refractometer, thermometer and pH meter. Then waited for three weeks until the *Artemia* gained matured size. After gaining matured size, the toxicity test was executed.

2.4. Observation of physico-chemical variables

Salinity, temperature and pH of the aquaria water were measured with the help of refractometer (Model: DR103, The United Kingdom), thermometers, and Hanna pH meter (HI 9124 HI 9125, China) respectively. Dissolved oxygen is determined by the titrimetric method developed by Winkler [16].

2.4.1. Calculation

Dissolved oxygen ppm = volume of sodium thiosulphate consumed in mL × its normality $\times 0.008 \times 10^6$ /volume of sample in mL.

2.5. Toxicity test

For the toxicity test, four jars were taken with four liters saline water. Then 20 species of *Artemia* was given into each jar. After that, different concentration (e.g. 1500, 2000, 2500, 3000, 3500 mg/L) of leaf powder was given to the *Artemia* species in each jar with micro pipette. Then reading was taken after 6, 12, 18 and 24 h. After that, the mortality rate was calculated. The measured water parameter of the aquarium was (salinity 20-70) ppt, temperature (28-30)°C, pH (6.8 ± 0.13) and dissolved oxygen (8.5 ± 0.20) mg/L. The experiment was repeated twice for the three plants extracts.

2.6. Statistical analysis

All the data were analyzed statistically. Statistical analyses were done by Probit analysis [17]. Probit analysis suggested by Fisher and Yates [18] was followed graphically and for determining the LC_{50} values mathematically. The values for empirical Probit, working Probit were taken from the tables given by Finney [19].

3. Results and Discussion

From the present study it was found that the water-soluble extracts of the leaf powder of the indigenous plant's species (Neem, Datura and Acanthus) were effective in killing the *Artemia* at low concentration 1500 mg/L in Neem, 2000 mg/L in Datura and 2500 mg/L in Acanthus. Behavioral changes become gradually apparent with the increase in the exposures of time and concentration. At medium concentrations 2000 mg/L to 2500 mg/L in Acanthus, neem and 2500 mg/L to 3000 mg/L Datura and 3000 mg/L to 3500 mg/L in Acanthus, *Artemia* subjected to change in their behavioral pattern. The *Artemia* began to move up and down vigorously soon after they were released into the higher concentration and came up to the surface repeatedly. After a considerable period of time, the *Artemia* lost their stability to swim. Their movement becomes slow down and closed their mouths, consequently, the whole body becomes paralyzed. Finally, mortality occurred at a different interval of time.

Different concentrations of the water extracts caused varying rates of *Artemia* mortality during the 24 h exposure period. In all cases, the mortality rate was low at low concentration and it increased with the increased concentration and exposure time. The mortality recorded after 24 h exposures to the extracts of *Azadirachta indica* were 30% at 1500 mg/L and increased to 70% at 3000 mg/L (Table 1).

Name of the plant extract	Artemia (Species)		Numb	er of Ar	temia s	pecies d	ied after	r exposu	re (h)				
		Concentration (mg/L)	6		12		18		24				
			No. of died	M %	No. of died	M %	No. of died	M %	No. of died	M %			
Neem Leaf	20	Control	0	0	0	0	0	0	0	0			
	20	1500	0	0	1	5%	3	15%	6	30%			
	20	2000	0	0	2	10%	4	20%	7	35%			
	20	2500	1	5%	3	15%	6	30%	10	50%			
	20	3000	2	10%	5	25%	9	45%	14	70%			

Table 1. Mortality of Artemia exposed to the water-soluble extracts of Azadirachta indica.

After 24 h exposure to the extracts of *Datura stremonium* mortality was 40% at 2000 mg/L and raised to 75% at 3500 mg/L (Table 2).

Table 2. Mortality of Artemia exposed to the water-soluble extracts of Datura stremonium.

Name of the plant extract	Artemia (Species)		Number of Artemia species died after exposure (h)								
		Concentration (mg/L)	6		12		18		24		
			No. of	Μ	No. of	Μ	No. of	Μ	No. of	М	
			mortality	%	mortality	%	mortality	%	mortality	%	
ıf	20	Control	0	0	0	0	0	0	0	0	
Leaf	20	2000	0	0	2	10%	5	25%	8	40%	
ra	20	2500	1	5%	3	15%	6	30%	10	50%	
Datura	20	3000	2	10%	5	25%	9	45%	14	70%	
	20	3500	2	10%	5	25%	10	50%	15	75%	

After 24 h exposure to the extracts of *Acanthus ilicifolius* mortality was 35% at 2500 mg/L and raised to 80% at 4000 mg/L. The 24 h LC_{50} values for the different concentrations of the extract of *Azadirachta indica*, *Datura stremonium* and *Acanthus ilicifolius* are given in Table 3.

Table 3. Mortality of Artemia exposed to the water-soluble extracts of Acanthus ilicifolius.

Name of the plant extract	Artomia	Conc. (mg/L)	Number of Artemia species died after exposure (h)								
			6		12		18		24		
			No. of M		No. of	М	No. of	М	No. of	М	
			mortality	%	mortality	%	mortality	%	mortality	%	
Acanthus Leaf	20	Control	0	0	0	0	0	0	0	0	
	20	2500	0	0	1	5%	3	15%	7	35%	
	20	3000	1	5%	3	15%	6	30%	10	50%	
	20	3500	2	10%	5	25%	9	45%	14	70%	
	20	4000	3	15%	6	30%	10	50%	16	80%	

4. Conclusion

Toxic plants have a great impact to brine shrimp *Artemia* species. After giving the concentration of leaf powder, *Artemia* species gradually lose their surviving attitude. Then mortality rates recorded after 24 h exposure. Lethal effects of toxic plants on *Artemia* is an exceptional task and it will bring new dimensions for the researcher in the future.

References

- 1. M. H. Al-Seria, R. R. Al-Ani, and A. G. Anwer, Baghdad Sci. J. 8, 66 (2011).
- 2. W. Tackaert and P. Sorgeloos, The Use of Brine Shrimp *Artemia* in Biological Management of Solar Salt Works (1993) pp. 617.
- 3. G. Van Stappen, G. Fayazi, and P. Sorgeloos. Hydrobiologia **466**, 133 (2001). <u>https://doi.org/10.1023/A:1014510730467</u>
- 4. W. D. Williams, Environ. Conservation **29**, 154 (2002). https://doi.org/10.1017/S0376892902000103
- 5. G. D. Treece, *Artemia* Production for Marine Larval Fish Culture, Southern Sorgeloos, ed. D. A. Bengtson et al. (*Artemia* Research and its Wettern, Belgium, 2000) p. 201.
- 6. G. V. V. Stappen, Introduction, Biology and Ecology of *Artemia* (Laboratory of Aquaculture and *Artemia* Ref. cent. University of Gent, Belgium, 1996).
- P. Sorgeloos, The Use of Brine Shrimp Artemia in Aquaculture, the Brine shrimp Artemia. Ecology, Culturing, Use in Aquaculture, ed. G. Persoone et al. (Universa Press, Wetteren, Belgium, 1980) 3, pp. 25.
- P. Leger, D. A. Bengtson, P. Sorgeloos, K. L. Simpson, and A. D. Beck, The Nutritional Value of *Artemia*: A Review In: *Artemia* Research and its Applications. Ecology, Culturing, Use in Aquaculture, ed. P. Sorgeloos et al. (Universa Press, Wetteren, Belgium 556, 1987) 3, pp. 357.
- 9. A. Seale, Trans. American Fisher. Soci. 63, 129 (1933).
- 10. M. Hudinaga, National Res. Coun. Japan 10, 305 (1942).
- 11. A. Ghani, Asiat. Soc. Bangladesh (2003).
- 12. A. Aktar, M. Sc Thesis, University of Dhaka, Bangladesh (1996).
- T. W. Sam, Toxicity Testing Using the Vrine Shrimp: *Artemia salina*. In Bioactive Natural Products Detection, Isolation and Structural Determination, ed. S. M. Colegate et al. (CRC press, Baca Raton, FL, 1993) pp. 442.
- 14. M. R. Chowdhury, M.Sc. thesis, IMSF, Chittagong University, Bangladesh (1986).
- 15. I. A. Chowdhury, Study on *Artemia* Biomass Production in Outdoor Culture at a Hatchery in Cox's Bazar (IMSF. Chittagong University, Bangladesh, 2005).
- 16. L. W. Winkler, The Determination of Dissolved Oxygen in Water, Berlin. Deut. Chem. yeas, 21.2845 Nov, 133 (1981).
- 17. D. J. Finney, Probit Analysis, 3rd Edition (Cambridge University Press, London, 1971) pp. 333.
- R. A. Fisher and F. Yates, Statistical Tables for Biological, Agricultural and Medicinal Research, 6th Edition (Oliver and Boyd Ltd., Edinburgh, 1963) pp. 34.
- 19. D. J. Finney, Probit Analysis (Cambridge University Press, London, 1949) pp.137.