

Molecular Diagnostics versus Traditional Culture Method for Diagnosis of Infectious Diseases

Mizanur Rahman¹

1. Sr. Consultant,
Molecular Diagnostics,
Evercare Hospital Dhaka.

Address for Correspondence :

Dr. Mizanur Rahman
Sr. Consultant
Molecular Diagnostics Lab,
Evercare Hospital Dhaka.
mizanur.rahman@evercarebd.com

Submitted: 20 – Mar - 2024

Accepted: 25 – Jun - 2024

ABSTRACT

Molecular diagnostic (MDx) techniques have revolutionized the field of clinical diagnostics, providing several advantages over traditional microbial culture methods. The ability to perform multiplex molecular panel tests is a significant advancement that allows for the simultaneous detection of multiple pathogens from a single specimen. Multiplex panels are available for various types of infections, including respiratory, gastrointestinal, urinary, and sexually transmitted infections. This versatility makes them valuable tools in different clinical scenarios. The integration of these advanced diagnostic tools into routine clinical practice holds great promise for improving patient outcomes and public health.

Keywords: Culture, Molecular diagnostics, PCR, Infectious disease.

INTRODUCTION

Microbial culture is traditionally the gold standard for the detection of many infectious pathogens¹⁻⁶. However, there are limitations. Limitations of culture methods include long turnaround times, labor-intensive processes, and difficulty in identifying difficult-to-culture microorganisms. Molecular diagnostics (MDx), especially nucleic acid amplification tests (NAATs), offer several advantages over traditional culture methods. MDx technologies are faster, more sensitive, and more specific in detecting infectious pathogens. NAATs can detect organisms that may be missed by routine culture. Worldwide infectious disease experts already have acknowledged the advantages of NAAT-based diagnostics. The Infectious Diseases Society of America (IDSA) and the American Society for Microbiology recommend the use of NAATs, stating that they have largely replaced rapid antigen tests and culture for respiratory virus detection⁷. MDx, particularly NAATs, have played a pivotal role in the detection of infectious agents during the coronavirus crisis. The US Centers for Disease Control and Prevention (CDC) notes that laboratory-based NAATs are considered the most sensitive tests for detecting SARS-CoV-2, the virus

responsible for COVID-19⁸. The World Health Organization (WHO) states that confirmation of monkeypox infection relies on NAAT, specifically using real-time or conventional polymerase chain reaction (PCR) to detect unique sequences of viral DNA⁹.

ADVANTAGES OF MOLECULAR DIAGNOSTICS

Besides being quicker and more accurate than microbial culture, MDx has facilitated the development of multiplex molecular panels, enabling comprehensive testing for a range of potential pathogens in a single test from a single specimen. This advancement has implications for various clinical scenarios:

1. Comprehensive Testing with Multiplex Molecular Panels:
 - Multiplex molecular panels can test for a breadth of potential pathogens in a single test, providing a more comprehensive view of the microbial landscape in a given specimen.
 - In cases where the dominant symptom is non-specific, such as diarrhea, clinicians can order a single test that covers multiple

pathogens, avoiding the need for multiple individual tests.

2. Challenges in Traditional Testing:

- Traditional testing methods, such as pathogen-specific cultures, may be less effective in identifying causative agents, especially when symptoms are ambiguous or multiple pathogens are involved.
- Despite repeated testing using individual tests, the causative agents of certain symptoms may remain unknown, making diagnosis difficult and treatment nonspecific.

3. Applications Across Different Infections:

- Multiplex molecular panel tests are available for various types of infections, including gastrointestinal, respiratory, urinary, and sexually transmitted infections.
- These tests offer a streamlined approach for diagnosing a range of conditions, from upper respiratory tract infections to urinary tract infections, vaginal discharge, and fevers of unknown origin.

4. Applications Across Different Infections:

- The faster and more accurate diagnosis provided by multiplex molecular panels allows for earlier and more specific treatments.
- This streamlined approach has the potential to lead to better clinical outcomes by enabling targeted interventions based on accurate identification of the causative agents.

5. Reduction in Inappropriate Antibiotic Usage:

- By facilitating a more specific diagnosis including resistance gene information, multiplex molecular panels may contribute to a reduction in inappropriate antibiotic usage.
- Clinicians can tailor treatments based on the identified pathogens, avoiding unnecessary and broad-spectrum antibiotic prescriptions.

Gastrointestinal System

There are challenges associated with the traditional culture-based diagnosis of *Campylobacter* infections in the gastrointestinal system and the advantages of using molecular methods, such as PCR and quantitative PCR (qPCR), for the detection of *Campylobacter* and other gastrointestinal pathogens¹⁰. *Campylobacter* is a leading cause of human gastroenteritis worldwide, particularly affecting vulnerable populations. Difficulties in culturing *Campylobacter* contribute to its underdiagnosis. *Campylobacter* species are fastidious, requiring specific growth conditions, including a microaerobic environment and a temperature of 42°C. Culture-based diagnosis involves stool sample implantation into a selective medium and a lengthy incubation period (about 72 hours), followed by an additional time for bacterial identification (up to 7 days). Molecular tests are found to be superior to culture-based methods in terms of sensitivity, specificity, and positive predictive value for *Campylobacter* detection. In a study comparing culture and culture-independent tests, PCR identified more *Campylobacter*-positive specimens than culture (sensitivity of 51.2% for culture)¹¹. The Global Enteric Multicenter Study (GEMS) showed higher attributable incidences of *Campylobacter* and other gastrointestinal pathogens with qPCR compared to traditional microbiological methods¹². qPCR demonstrated higher sensitivity for various pathogens, including adenovirus, *Shigella* spp / Enteroinvasive *Escherichia coli* (EIEC), and heat-stable enterotoxin-producing *Escherichia coli*. There has been a marked increase in the use of culture-independent diagnostic tests (CIDTs) for detecting *Campylobacter*, *Salmonella*, *Shigella*, and other gastrointestinal pathogens¹³⁻¹⁶. Healthcare providers in developed countries are more likely to order CIDTs and DNA-based syndromic panels due to their speed and ease of use compared to traditional culture methods. Centers for Disease Control (CDC) and Prevention confirms the trend of increased use of CIDTs over the last decade.

In summary, the shift towards molecular methods, particularly PCR and qPCR, for the diagnosis of

Campylobacter and other gastrointestinal pathogens is driven by their faster turnaround time, higher sensitivity, and ease of use compared to traditional culture-based methods. This transition has implications for improving the accuracy and efficiency of diagnosing and managing gastrointestinal infections.

Respiratory System

The utility of multiplex molecular panels, particularly in the context of detecting causative agents in atypical pneumonia is underscored. Atypical pneumonias historically posed challenges due to their varied symptoms, distinct chest X-ray appearances, and poor response to standard antibiotics. The term "atypical" now refers to pneumonias that are difficult to detect through traditional bacterial methods like culture¹⁷. The findings from a study highlight the effectiveness of multiplex molecular panels, specifically multiplex reverse transcription polymerase chain reaction (RT-PCR), in identifying bacterial pathogens causing atypical pneumonia¹⁸. Historically, atypical pneumonias were characterized by different symptoms, distinctive chest X-ray appearances, and poor response to standard antibiotics. Currently, the term "atypical" is associated with pneumonias that are challenging to detect through standard bacterial methods like culture. The study¹⁸ focused on patients with symptoms of atypical pneumonia who tested negative for typical pneumonia agents through both culture and viral PCR. Researchers used multiplex RT-PCR to detect key bacterial pathogens causing atypical pneumonia in these cases. Among 368 samples that were culture- and viral PCR-negative, multiplex RT-PCR identified specific bacterial pathogens. Positive results were observed for *Bordetella parapertussis*, *Bordetella pertussis*, *Chlamydia pneumoniae*, *Chlamydia psittaci*, *Legionella pneumophila*, *Legionella* spp., and *Mycoplasma pneumoniae*. Co-infection of *Bordetella pertussis* and *Bordetella parapertussis* was observed in two patients. Multiplex molecular panels, by accurately identifying causative pathogens, have the potential to improve the diagnosis of atypical pneumonia. Accurate

identification of pathogens is crucial for guiding appropriate patient management and treatment strategies. This technology enables the identification of specific bacterial pathogens that may go undetected using traditional culture and viral PCR methods, leading to improved diagnostic accuracy and subsequent patient management.

Urinary System

Culture and MDx differ in their ability to detect co-infection or polymicrobial infection. A study involving 582 patients with lower urinary tract infections, where PCR demonstrated significantly higher sensitivity compared to urine culture in detecting polymicrobial infections¹⁹. Among the 175 patients with polymicrobial infections, PCR reported 95% of cases, whereas culture only reported 22%. PCR revealed polymicrobial infections in an additional 67 patients who had negative culture results. This indicates that PCR has the ability to detect infections that may be missed by traditional culture methods. Simultaneous detection of various pathogens through PCR can have implications for clinical management, enabling more specific treatments and reducing the likelihood of recurrent infections resulting from inadequate or inappropriate treatments. Additionally, the combination of multiplex PCR with pooled antibiotic sensitivity testing enhances the microbiological data obtained from standard urine culture methods²⁰.

Sexually Transmitted Diseases and Other Infections

The emphasis on NAATs and MDx in the recommendations by CDC, and WHO highlighted already the importance of these advanced diagnostic methods in the detection, confirmation, and monitoring of infectious diseases, particularly those with implications for public health, such as sexually transmitted diseases such as *Chlamydia trachomatis* and *Neisseria gonorrhoeae* and viral infections like HIV and Hepatitis^{7, 21}. The third WHO Model List of Essential In Vitro Diagnostics includes PCR for *Pneumocystis jirovecii*, the fungus causing *Pneumocystis pneumonia*, and a

human measles reverse transcription-PCR (RT-PCR) test, which accurately confirms cases and helps prevent outbreaks. The ability to provide rapid and accurate results, along with the noninvasive nature of specimen collection, makes these methods invaluable in diverse clinical settings.

BENEFITS OF CULTURE

Here are the key points:

1. **Role of Microbial Culture:** Microbial culture serves as a benchmark for diagnosing many infectious diseases. This method is particularly relevant in areas where MDx technologies are underdeveloped, costly, require skilled personnel, or may yield results that are clinically insignificant.
2. **Antimicrobial Susceptibility Testing:** Culture is crucial for determining antimicrobial susceptibility, which is essential for tailoring effective treatment plans. Examples include assessing susceptibility in patients with Mycobacterium tuberculosis infections and pregnant women with penicillin allergy who have Group B Streptococcus Colonization²²⁻²³.
3. **Molecular Subtyping:** In addition to antimicrobial susceptibility data, culture may be necessary for molecular subtyping. This information is vital for identifying antibiotic resistance, tracking outbreaks, and monitoring disease trends.
4. **Public Health Surveillance:** The CDC encourages the use of reflex culture in laboratories for bacteria of public health importance. Reflex culture involves culturing specimens that have tested positive using CIDT (Culture-Independent Diagnostic Tests). This is particularly relevant for bacteria like Campylobacter, Salmonella, Shigella, Shiga toxin-producing Escherichia coli (STEC), Vibrio, and Yersinia infections⁽¹³⁻¹⁶⁾.
5. **Importance of Information for Surveillance:** The information obtained through culture is crucial for public health surveillance. It aids in identifying and monitoring the prevalence of specific pathogens, tracking antibiotic

resistance patterns, and responding to outbreaks effectively.

CONCLUSION

The decision to supplement culture with Molecular Diagnostics (MDx) for infectious diseases indeed involves various factors such as, technology performance, regulatory considerations, financial aspects, accessibility, and awareness. Laboratories that have embraced MDx infrastructure can leverage their capabilities not only for the recent pandemic but also for a broader range of infectious diseases, contributing to more effective disease management and public health responses.

REFERENCES

1. Kostyusheva A, Brezgin S, Babin Y, Vasilyeva I, Glebe D, Kostyushev D, Chulanov V. Methods. CRISPR-Cas systems for diagnosing infectious diseases. 2022 Jul;203:431-446. doi: 10.1016/j.ymeth.2021.04.007. Epub 2021 Apr 9. PMID: 33839288
2. Strich JR, Chertow DS. CRISPR-Cas Biology and Its Application to Infectious Diseases. J Clin Microbiol. 2019 Mar 28;57(4):e01307-18. doi: 10.1128/JCM.01307-18.
3. Lau SK, Sridhar S, Ho CC, Chow WN, Lee KC, Lam CW, Yuen KY, Woo PC. Laboratory diagnosis of melioidosis: past, present and future. Exp Biol Med (Maywood). 2015 Jun;240(6):742-51. doi: 10.1177/1535370215583801. Epub 2015 Apr 22. PMID: 25908634
4. Limmathurotsakul D, Jamsen K, Arayawichanont A, Simpson JA, White LJ, Lee SJ, Wuthiekanun V, Defining the true sensitivity of culture for the diagnosis of melioidosis using Bayesian latent class models. PLoS One. 2010 Aug 30;5(8):e12485. doi: 10.1371/journal.pone.0012485.
5. Suárez I, Fünfer SM, Kröger S, Rademacher J, Fätkenheuer G, Rybniker J. The Diagnosis and Treatment of Tuberculosis. Dtsch Arztebl Int. 2019 Oct 25;116(43):729-735. doi: 10.3238/arztebl.2019.0729. PMID: 31755407.
6. Skevaki CL, Kafetzis DA. Tuberculosis in neonates and infants: epidemiology, pathogenesis, clinical manifestations, diagnosis, and management issues. Paediatr Drugs. 2005;7(4):219-34. doi:10.2165/00148581-200507040-00002. PMID: 16117559.
7. Miller JM, Binnicker MJ, Campbell S, Carroll KC, Chapin KC, Gilligan PH, A Guide to Utilization of the Microbiology Laboratory for Diagnosis of Infectious Diseases: 2018 Update by the Infectious Diseases Society of America and the American Society for Microbiology. Clin Infect Dis. 2018 Aug 31;67(6):e1-e94. doi: 10.1093/cid/ciy381. PMID: 29955859; PMCID: PMC7108105.

8. COVID-19: Nucleic Acid Amplification Tests (NAATs) <https://www.cdc.gov/coronavirus/2019-ncov/lab/naats.html>
9. Laboratory Testing for the Monkeypox Virus: Interim Guidance <https://www.who.int/publications/i/item/WHO-MPX-laboratory-2022.1>
10. Campylobacter: Key Facts <https://www.who.int/news-room/fact-sheets/detail/campylobacter>
11. Özcan N, Bacalan F, Çakır F, Bilden A, Genişel N, Dal T. Culture and culture-independent diagnostic tests in Campylobacter enteritis. *J Infect Dev Ctries*. 2022 Apr 30;16(4):616-621. doi: 10.3855/jidc.14902. PMID: 35544622. <https://pubmed.ncbi.nlm.nih.gov/35544622/>
12. Liu J, Platts-Mills JA, Juma J, et al. Use of quantitative molecular diagnostic methods to identify causes of diarrhoea in children: a reanalysis of the GEMS case-control study. *Lancet*. 2016 Sep 24;388(10051):1291-301. doi: 10.1016/S0140-6736(16)31529-X. PMID: 27673470; PMCID: PMC5471845. <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC5471845/>
13. Marder EP, Cieslak PR, Cronquist AB, et al. Incidence and Trends of Infections with Pathogens Transmitted Commonly Through Food and the Effect of Increasing Use of Culture-Independent Diagnostic Tests on Surveillance — Foodborne Diseases Active Surveillance Network, 10 U.S. Sites, 2013–2016. *MMWR Morb Mortal Wkly Rep* 2017; 66:397–403. DOI: <http://dx.doi.org/10.15585/mmwr.mm6615a1> <https://www.cdc.gov/mmwr/volumes/66/wr/mm6615a1.htm>
14. Campylobacter (Campylobacteriosis): Information for Health Professionals <https://www.cdc.gov/campylobacter/technical.html>
15. Salmonella: Diagnostic and Public Health Testing <https://www.cdc.gov/salmonella/general/diagnosis-treatment.html>
16. Foodborne Illness and Culture-Independent Diagnostic Tests (CIDTs) <https://www.cdc.gov/foodnet/reports/cidt-questions-and-answers-2015.html>
17. Atypical Pneumonia <https://www.cdc.gov/pneumonia/atypical/index.html>
18. Wagner K, Springer B, Imkamp F, et al. Detection of respiratory bacterial pathogens causing atypical pneumonia by multiplex Lightmix® RT-PCR. *Int J Med Microbiol*. 2018 Apr;308(3):317-323. doi: 10.1016/j.ijmm.2018.01.010. Epub 2018 Jan 31. PMID: 29397298. <https://pubmed.ncbi.nlm.nih.gov/29397298/>
19. Wojno KJ, Baunoch D, Luke N, et al. Multiplex PCR Based Urinary Tract Infection (UTI) Analysis Compared to Traditional Urine Culture in Identifying Significant Pathogens in Symptomatic Patients. *Urology*. 2020 Feb;136:119-126. doi: 10.1016/j.urology.2019.10.018. Epub 2019 Nov 9. PMID: 31715272. <https://pubmed.ncbi.nlm.nih.gov/31715272/>
20. Luke, D. Baunoch. Baunoch. After 180 Years, Is It Time for Something Better for Diagnosing UTI's? . *JOJ Urology & Nephrology*, 2020; 7(2): 555714. DOI: 10.19080/JOJUN.2020.07.555714 <https://juniperpublishers.com/jojun/JOJUN.MS.ID.555714.php>
21. Centers for Disease Control and Prevention. Sexually transmitted disease treatment guidelines. *MMWR Morb Mortal Wkly Rep* 2015; 64:1–135. <https://www.cdc.gov/mmwr/preview/mmwrhtml/rr6403a1.htm>
22. Report of an Expert Consultation on the Uses of Nucleic Acid Amplification Tests for the Diagnosis of Tuberculosis https://www.cdc.gov/tb/publications/guidelines/amplification_tests/default.htm
23. Shin JH, Pride DT. Comparison of Three Nucleic Acid Amplification Tests and Culture for Detection of Group B Streptococcus from Enrichment Broth. *J Clin Microbiol*. 2019 May 24;57(6):e01958-18. doi: 10.1128/JCM.01958-18. PMID: 30944190; PMCID: PMC6535594. <https://pubmed.ncbi.nlm.nih.gov/30944190/>