

Research Article

FACTOR INFLUENCING THE RESURGENCE OF BROWN PLANTHOPPER IN BANGLADESH

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ABSTRACT

Possible causes of brown plant hopper resurgence were determined at the net-house of Entomology Division of Bangladesh Rice Research Institute (BRRRI) during 2015. Causes of resurgence in the form of resurgence ratios were higher with acetamiprid, acephate, chlorpyrifos, cypermethrin, deltamethrin, fenvalerate, lambda cyhalothrin, thiamethoxam insecticides compared to imidacloprid, cartap, dinotefuran, isoprocarb /MIPC, phenthoate, pymetrozine when even applied at recommended dose. However, thiamethoxam, imidacloprid, isoprocarb / MIPC and cartap applied at sub-lethal dose produced higher resurgence ratio of BPH than others. Isoprocarb / MIPC, a commonly used recommended insecticide was found to have a higher resurgence ratio with the insecticide treatment at the egg stage (1.71) and combination of all stages (0.82). These insecticides influenced on the growth and reproductive physiology of rice brown planthopper and consequently resurgence ratio ranged increased.

Keywords: Brown planthopper, Dose, Insecticide, Resurgence, Stage

INTRODUCTION

Rice (*Oryza sativa* L.) is one of the world's most important crops providing a staple food for more than half of the global population (Muthayya et al., 2014). A large complex of pest organisms, consisting of insects, vertebrates, disease and weeds, have been associated with rice for hundreds of years (Islam and Catling, 2012). There are more than 232 insects (Ali et al., 2017) and several vertebrate pest species, which cause damage to the rice plants (Islam et al., 2003). Out of this large complex, about 20-30 species may be considered as the major pests and these have the potential to cause significant yield loss (Krishnaiah et al., 2008). Among the pests infesting rice, the brown planthopper (BPH), *Nilaparvata lugens* (Stal.) (Homoptera: Delphacidae)

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has gained major importance in several Asian countries including Bangladesh. The loss in grain yield ranges from 10 to 70% (Liu and Sun, 2016). It is also a vector in transmitting viral diseases such as grassy stunt, ragged stunt and wilted stunt (Cabauatan et al., 2009). In Bangladesh rice crops worth at least US\$ 8.1 million were lost due to BPH attack during the last three widespread outbreaks in 1976, 1978 and 1983 (Alam and Karim, 1986). Nowadays its outbreak is occurring frequently in different areas of Bangladesh (Ali et al., 2014), Sri Lanka (Sivasubramaniam and Imthiyas, 2018) and many other countries.

The control of this insect pest has always been emphasized and largely relied on insecticides in most rice producing countries (Ali et al., 2019) especially in countries where commercial, resistant varieties are not available. All the pesticides have different types of effect on the pest, which may lead to the differential development of the next generation of the pest (Volodymyr et al., 2018). Heavy uses of broad-spectrum chemicals also reduce the biodiversity of natural enemies, lift the natural control, induce outbreak of secondary pests and contaminate ecosystem (Ali et al., 2019). Continuous use of insecticides has resulted in BPH resistance to insecticides (Wu et al., 2018) and its outbreaks.

Resurgence is one of the major causes for BPH outbreak. After application of insecticides, its resurgence was reported in Bangladesh (Alam, 2013), India (Ghosal and Chatterjee, 2018), the Philippines (Heong and Hardy, 2009) and the Poland (Wojciechowska et al., 2016). In insecticide trials in experimental stations and in farmer's fields, hopper burn commonly occurs in treated plots while untreated areas remain relatively free of infestation (FAO, 2006). Existing literatures suggest that resurgence of BPH take place after application of some insecticides (Alam, 2013; Wojciechowska et al., 2016) that killed natural enemies (Bommarco et al., 2011) while on the other hand, it could be because of stimulated fecundity of certain pests after the applications of some insecticides (Wang et al., 2005). Improper methods of application of some insecticides also caused resurgence. To find out the reasons for resurgence of BPH, an experiment was planned, designed and conducted in the net house conditions.

MATERIALS AND METHODS

The trial was conducted at entomology net house of Bangladesh Rice Research Institute during 2015. The trial cage consisted of a steel frame covered with fine mesh wire screen. The size of the cage was 152 × 66 × 84 cm. It consisted of three chambers. The size of each chamber was 51 × 65 × 84cm. The cage bottom was open and placed on a tray made of steel (183 × 91 × 15 cm). Standing water was maintained in the tray. The tray with cage was placed in an iron frame (183 × 76 × 76 cm). Twenty-five to 30-day-old plants were used as experimental units. Each pot contained three hills and the number of tillers per pot ranged from 40-50. The plants in the pot were cleaned by removing dried, diseased and insect infested leaves.

Before imposing treatments, it was also confirmed that plants were free from eggs of any insects. Each pot was covered by Mylar cage of height 76.2 cm and diameter 50.4 cm. Ten gravid BPH female were introduced in each Mylar cage followed by closing the top with net. The gravid female within the pots were allowed to lay egg for 48 hours. Egg bearing pot was kept in chamber. Three pots were introduced in each chamber, which was considered as a replication. In all growth stages, BPH were collected from rearing chamber and released into experiment chamber (*ca.* 100/pot). Burned tillers bearing pots were replaced by fresh tiller bearing pots during study period as and when necessary. The partition of each chamber was covered by thick polythene sheet in order to keep the original condition of each chamber. This experiment was laid out in a Randomized Complete Block Design (RCBD) with three replications. Each chamber with three pots was considered as one replication of a treatment, and each room treated as one block.

Effect of different insecticides on the development of BPH resurgence

Fourteen groups of single molecule containing chemical insecticides were applied at recommended dose (Table 1). Fresh water was applied on the plants of control treatments. Hand sprayer (Seesa hand pressure sprayer) was used to spray insecticide.

Table 1. Name of the insecticides with concentration used in the treatments

Treatment	Name of insecticide	Recommended dose (g or ml/L water)
T ₁	Acetamiprid (Tundra 20 SP)	0.25 g
T ₂	Acephate (Mimpahte 75 SP)	1.50 g
T ₃	Imidacloprid (Admire 20 SL)	0.25 ml
T ₄	Cartap (Suntap 50 SP)	2.40 g
T ₅	Chlorpyrifos (Dursban 20 EC)	2.00 ml
T ₆	Cypermethrin (Cymbaz 10 EC)	1.00 ml
T ₇	Deltamethrin (Decis 2.5 EC)	1.00 ml
T ₈	Dinotefuran (Token 20SG)	0.30 g
T ₉	Fenvalerate (Fenfen 20 EC)	0.50 ml
T ₁₀	Isoprocarb/MIPC (Chabi 75 WP)	2.60 g
T ₁₁	Lambda cyhalothrin (Karate 2.5 EC)	1.00 ml
T ₁₂	Phenthoate (Kiron 50 EC)	2.00 ml
T ₁₃	Pymetrozine (Plenum 50 WG)	0.60 g
T ₁₄	Thiamethoxam (Spike 25 WG)	0.12 g
T ₁₅	Control	Only water

Effect of different doses of selected insecticides on resurgence development

Three doses *viz.* lower, recommended and higher dose of five selected insecticides were sprayed in an experimental arena as treatment with the help of a hand sprayer. A control experimental site was also maintained which was sprayed with fresh water only (Table 2).

Table 2. Name of the treatments of insecticides with different doses

Treatment	Insecticide	Low, recommended and higher dose (g or ml/L water)
T ₁	Imidacloprid (Admire 20 SL)	0.20 ml
T ₂	Imidacloprid (Admire 20 SL)	0.25 ml
T ₃	Imidacloprid (Admire 20 SL)	0.30 ml
T ₄	Cartap (Suntap 50 SP)	2.20 g
T ₅	Cartap (Suntap 50 SP)	2.40 g
T ₆	Cartap (Suntap 50 SP)	2.80 g
T ₇	Isoprocarb/MIPC (Chabi 75 WP)	2.20 g
T ₈	Isoprocarb/MIPC (Chabi 75 WP)	2.60 g
T ₉	Isoprocarb/MIPC (Chabi 75 WP)	2.80 g
T ₁₀	Pymetrozine (Plenum 50 WG)	0.30 g
T ₁₁	Pymetrozine (Plenum 50 WG)	0.60 g
T ₁₂	Pymetrozine (Plenum 50 WG)	0.90 g
T ₁₃	Thiamethoxam (Spike 25 WG)	0.10g
T ₁₄	Thiamethoxam (Spike 25 WG)	0.12 g
T ₁₅	Thiamethoxam (Spike 25 WG)	0.14 g
T ₁₆	Control	Only water

Effect of insecticides on different stages of BPH for the possible resurgence development

Different stages of BPH, such as T₁= egg, T₂= 1st-2nd instar nymph, T₃= 3rd-4th instar nymph, T₄= adult and T₅= combination of all stages was used as the treatment. Specific arena was developed according to treatment. A control chamber (T₆) was also maintained consisting all growth stages of BPH. Isoprocarb / MIPC insecticide (Chabi 75 WP) was applied in each chamber at the rate of 2.6 g/L of water. The control chamber was sprayed with fresh water.

Data Collection: The BPH populations were recorded before imposing treatments through counting. The BPH populations were recorded after 72 hours and 30 days of

spraying. The resurgence ratio of BPH was calculated by the following equation (Heinrichs et al., 1981):

$$\text{Resurgence ratio} = \frac{\text{Population after application of insecticide}}{\text{Population in check field at the same interval}}$$

Statistical analysis: The collected data were arranged as required for statistical analysis. The software program STATISTIX 10 was selected to analysis the data as it is reported to be more efficient in analyzing entomological data. The mean differences among the treatments were determined by LSD test.

RESULTS AND DISCUSSION

Effect of different insecticides on the development of BPH resurgence

The number of BPH/pot was not significantly different among the treatments before insecticide spray (Table 3), although it varied significantly after 72 hours and one month of spraying. The data of table 3 clearly showed that the resurgence ratio was significantly higher in T₆ (2.06) followed by T₇, T₁, T₁₁ and T₉. Moderate resurgence ratio was found in T₁₄ (1.07) and it was not significantly different with resurgence ratio of T₂ (1.03) and T₅ (1.02). The lowest resurgence ratio was found in T₈ and T₁₃ (0.25 to 0.26) followed by T₁₂ (0.71), T₄ (0.82), T₁₀ (0.83) and T₃ (0.96). T₈ and T₁₃ were almost identical and T₄, T₁₀ were not significantly different among them.

Table 3. Effect of single molecule containing insecticides on resurgence development

Treatment	Recommended dose (g or ml/L water)	No. of BPH/pot at different time intervals			Resurgence ratio
		Before spray (pretreatment)	After 72 hours of spray	After one month of spray	
T ₁ Acetamiprid (Tundra 20 SP)	0.25 g	109.33a	50.00b	212.67bc	1.38bc
T ₂ Acephate (Mimpahte 75 SP)	1.50 g	109.33a	17.33f	159.00de	1.03d
T ₃ Imidacloprid (Admire 20 SL)	0.25 ml	110.67a	37.00	149.00f	0.96e
T ₄ Cartap (Suntap 50 SP)	2.40 g	110.33a	10.67g	126.67g	0.82f
T ₅ Chlorpyrifos (Dursban 20 EC)	2.00 ml	110.33a	10.67g	157.33e	1.02d
T ₆ Cypermethrin (Cymbaz 10 EC)	1.00 ml	109.67a	49.00b	318.33a	2.06a

Treatment	Recommended dose (g or ml/L water)	No. of BPH/pot at different time intervals			Resurgence ratio	
		Before spray (pretreatment)	After 72 hours of spray	After one month of spray		
T ₇	Deltamethrin (Decis 2.5 EC)	1.00 ml	109.67a	48.00b	215.67b	1.40b
T ₈	Dinotefuran (Token 20SG)	0.30 g	110.00a	3.00i	39.00i	0.25h
T ₉	Fenvalerate (Fenfen 20 EC)	0.50 ml	109.33a	79.00a	206.67c	1.34c
T ₁₀	Isoprocarb/MIPC (Chabi 75 WP)	2.60 g	110.00a	10.67g	129.00g	0.83f
T ₁₁	Lambda cyhalothrin (Karate 2.5 EC)	1.00 ml	111.33a	25.67e	210.00bc	1.36bc
T ₁₂	Phenthoate (Kiron 50 EC)	2.00 ml	108.00a	17.33f	109.33h	0.71g
T ₁₃	Pymetrozine (Plenum 50 WG)	0.60 g	109.67a	6.67h	40.00i	0.26h
T ₁₄	Thiamethoxam (Spike 25 WG)	0.12 g	107.67a	32.00d	165.00d	1.07d
T ₁₅	Control	Only water	108.00a	79.67a	154.67ef	-
Level of significance			NS	**	**	**
LSD _{0.05}			9.92	3.29	6.98	0.05
CV%			5.41	6.19	2.62	2.97

**= Significant at 1% level of probability, *= Significant at 5% level of probability

NS= Non-Significant, Values in a column followed by different letters are significantly different

With increased adoption of new high yielding varieties, use of insecticides were also increased and the destruction of predators and parasitoids that followed insecticide misuse resulted in resurgence of several rice pests including the BPH, *Nilaparvata lugens* (Heinrichs and Mochida, 1984). Anand et al. (2019) conducted an experiment with seven insecticides viz., chlorpyrifos, profenophos, cypermethrin, deltamethrin, bifenthrin, lambda cyhalothrin and imidacloprid on the growth and reproductive physiology of rice brown planthopper. They reported that bifenthrin, cypermethrin, lambda cyhalothrin and deltamethrin resulted in enhancement of fecundity of brown planthopper (227.67, 218.33, 199.00 and 191.00 nymph's vs. 131.00 nymphs in control) and consequently resurgence ratio ranged from 1.18 to 1.74. Chlorpyrifos, cypermethrin, deltamethrin, bifenthrin and lambda cyhalothrin also significantly

increased the nymphal survival (86.67 per cent to 96.00 per cent against 80.67 per cent in control) and growth index (6.34 to 7.11 vs 5.63 in control).

Effect of different doses of selected insecticides on development of BPH resurgence

The data clearly showed that the number of BPH/pot was not significantly different (104 to 107) among the treatment before spray (Table 4) that differed significantly after 72 h and one month of spraying. The resurgence ratio was highest in T₁₃ (1.61) followed by T₁ (1.39) and then T₁₄ (1.30), T₇ (1.26) and T₄ (1.22). T₁₃ was significantly different with other treatments but T₁₄ and T₇ were insignificant. Resurgence ratio in T₁₅, T₁₀, T₂, T₉, T₃ and T₅ were 1.08, 1.02, 0.98, 0.97, 0.92, 0.91 and 0.91, respectively. Significantly the lowest resurgence ratio was found in T₁₂ (0.65) and T₁₁ (0.69) followed by T₆ (0.84). T₁₂ and T₁₁ were insignificantly different among them but significantly different with T₆.

Use of insecticide at lower dose is common in farmers' practice as it saves some money apparently. The practice of using low dose combined with short residual toxicity of many commercial insecticides may often cause the sub-lethal effect to the pest. The result is in conformity with the earlier findings (Heinrichs et al., 1982; Chelliah and Uthamasamy, 1986; Karns and Stewart, 2003. Heinrichs et al. 1982) and reported high BPH populations (40-fold) with lower application rates of FMC 3500 (0.2 kg a.i./ha) as compared to high rate (1.0 kg a.i./ha). Chelliah (1979) reported that low doses of resurgence-inducing insecticides increased the reproductive rate of the BPH and reduced the nymphal duration, eventually leading to resurgence. Heinrichs and Mochida (1984) reported that dose rates had a distinct effect on the degree of resurgence in both the decamethrin and methyl parathion treatments with the higher rates permitting the higher BPH populations. There were 850 BPH per hill at the high and 210 BPH per hill at the low decamethrin rate and 60 BPH per hill in the check. Present findings showed 20-50% increase in the levels of resurgence when low dose was used. Further lower dose might increase the resurgence ratio to some higher degree. The efficacy study of isoprocarb/MIPC showed it was an effective insecticide but when applied with sub-lethal dose was found to be developed resurgence. It clearly indicates that any recommended product or chemical could also be cause of resurgence development under improper doses. Bao et al. (2009) conducted an experiment on the effects of sublethal doses of four insecticides viz. triazophos, fenvalerate, imidacloprid and dinotefuran on the reproduction of BPH. Imidacloprid and dinotefuran reduced the fecundity of BPH to 68.8% and 52.4% in macropterous families, and to 57.9% and 43.1% in brachypterous families, when compared with the untreated controls. By contrast, triazophos and fenvalerate increased fecundity and consequently resurgence ratio increased.

Table 4. Effect of different doses of selected insecticides on resurgence development

Treatment	Low, Recommended and higher dose (g or ml/L water)	No. of BPH/pot at different time intervals			Resurgence ratio	
		Before spray (Pre-treatment)	After 72 hours of spray	After one month of spray		
T ₁	Imidacloprid (Admire 20 SL)	0.20 ml	104.67a	52.67bc	192.67b	1.39b
T ₂	Imidacloprid (Admire 20 SL)	0.25 ml	104.00a	49.00c	136.33fg	0.98fg
T ₃	Imidacloprid (Admire 20 SL)	0.30 ml	103.67a	30.00de	127.67gh	0.92gh
T ₄	Cartap (Suntap 50 SP)	0.20 g	104.33a	47.00c	169.67d	1.22d
T ₅	Cartap (Suntap 50 SP)	2.40 g	105.67a	11.00g	126.33h	0.91h
T ₆	Cartap (Suntap 50 SP)	2.80 g	104.00a	8.00g	117.00i	0.84i
T ₇	Isoprocarb/MIPC (Chabi 75 WP)	2.20 g	103.67a	23.00ef	174.67cd	1.26cd
T ₈	Isoprocarb/MIPC (Chabi 75 WP)	2.60 g	104.00a	13.00g	126.33h	0.91h
T ₉	Isoprocarb/MIPC (Chabi 75 WP)	2.80 g	104.67a	11.33g	135.00fgh	0.97fgh
T ₁₀	Pymetrozine (Plenum 50 WG)	0.30 g	104.67a	29.00de	142.00ef	1.02ef
T ₁₁	Pymetrozine (Plenum 50 WG)	0.60 g	105.00a	15.00fg	96.33j	0.69j
T ₁₂	Pymetrozine (Plenum 50 WG)	0.90 g	103.67a	16.00fg	90.67j	0.65j
T ₁₃	Thiamethoxam (Spike 25 WG)	0.10g	104.00a	60.33b	223.33a	1.61a
T ₁₄	Thiamethoxam (Spike 25 WG)	0.12 g	103.67a	50.33c	180.00c	1.30c
T ₁₅	Thiamethoxam (Spike 25 WG)	0.14 g	104.33a	32.00d	150.00e	1.08e
T ₁₆	Control	Only water	106.67a	79.67a	139.00f	-
Level of significance			NS	**	**	**
LSD _{0.05}			7.86	8.71	9.00	0.06
CV%			4.51	15.85	3.71	3.69

**= Significant at 1% level of probability, *= Significant at 5% level of probability

NS= Non-Significant, Values in a column followed by different letters are significantly different

Effect of insecticide on different stages of BPH for the development of resurgence

Five different growth stages including combination of all stages were sprayed with a most common insecticide— Isoprocarb/MIPC (Chabi 75 WP) for the development of resurgence (Table 5). In treatment T₁, the number of BPH/pot was nil (only bearing eggs) and there was no significant difference among the treatments (T₂ to T₆) before insecticide application. After 72 hours of spraying, significantly the highest number of BPH was found in control treatment; but after one month it was significantly higher with T₁ (303) followed by T₆ (178). Zero population was found in T₂ and T₃ treatments. Resurgence ratio was also significantly higher in T₁ (1.71) treatment compared to other treatments. But no significant difference was found between T₄ (0.78) and T₅ (0.82) treatments.

Table 5. Effect of isoprocarb/MIPC (Chabi 75 WP) on different stage of brown planthopper for the development of resurgence

Treatment	No. of BPH/pot at different time intervals			Resurgence ratio
	Before spray (pre-treatment)	After 72 hours of spray	After one month of spray	
T ₁ Egg	0.00b	0.00d	303.00a	1.71a
T ₂ 1 st -2 nd instar nymph	97.00a	1.67cd	0.00d	0.00c
T ₃ 3 rd -4 th instar nymph	100.00a	5.00c	0.00d	0.00c
T ₄ Adult	98.67a	15.67b	138.00c	0.78b
T ₅ Combination of all stages	100.00a	19.00b	144.44c	0.82b
T ₆ Combination of all stages (control)	99.67a	84.33a	177.67b	-
Level of significance	**	**	**	**
LSD _{0.05}	4.97	4.99	24.30	0.19
CV%	3.31	13.09	10.5	15.4

**= Significant at 1% level of probability, *= Significant at 5% level of probability

NS= Non-Significant, Values in a column followed by different letters are significantly different

Many researchers reported that the effectiveness of insecticide depends on the stage of the insect. The reproductive rates of BPH exposed to insecticide-sprayed plants during nymphal or adult stage or both varied significantly (Chelliah, 1979). The reproductive rate was significantly higher when the BPH was exposed to plants sprayed with resurgence-inducing insecticides at the fourth-and fifth-instar stage as well as the adult stage (Chelliah, 1979). The result of the present study clearly showed that the nymph of the BPH was more susceptible to insecticides than the egg

and adult stages. Therefore, care should be taken to apply insecticides depending on stage of the pest. There may be a requirement to spray the crop again after a few days to kill the early insects of pest developed from the primary sprayed egg.

CONCLUSION

The insecticide namely acetamiprid, acephate, chlorpyrifos, cypermethrin, deltamethrin, fenvalerate, lambda-cyhalothrin, thiamethoxam were responsible for higher resurgence of BPH. In contrast, imidacloprid, cartap, dinotefuran, isoprocarb/MIPC, phenthoate, pymetrozine showed low resurgence producing potentials. So, recommendation of insecticides for controlling BPH that produce low resurgence require special attention in selecting right dose at the right time because they may cause high resurgence of BPH when used at sub-lethal dose. A similar high resurgence was evident with insecticide treatment at egg and adult stage. There may be a requirement to spray the crop again after a few days to kill the early insects of pest developed from the primary sprayed egg.

REFERENCES

- Alam, M.Z. (2013). Survey and assessment of insect management technologies and environmental impact on rice ecosystem of Bangladesh. *International Journal of Applied Research and Studies*, 2(4): 1-16.
- Alam, S. and Karim, A.N.M.R. (1986). Brown planthopper situation and its management in Bangladesh. Proceedings of *the Workshop on Experiences with Modern Rice Cultivation in Bangladesh*, 8-11 April 1984. Bangladesh Rice Research Institute, Gazipur, Bangladesh. Pp. 10-29.
- Ali, M.P., Bari, M.N., Ahmed, N., Kabir, M.M.M., Afrin, S., Zaman, M.A.U., Haque, S.S. and Willers, J.L. (2017). Rice production without insecticide in smallholder farmer's field. *Frontiers in Environmental Science*, 5:16.
- Ali, M.P., Bari, M.N., Haque, S.S., Kabir, M.M.M., Afrin, S., Nowrin, F., Islam, M.S. and Landis, D.A. (2019). Establishing next-generation pest control services in rice fields: eco-agriculture. *Scientific Reports*, 9(1):1-9.
- Ali, M.P., Huang, D., Nachman, G., Ahmed, N., Begum, M.A. and Rabbi, M.F. (2014). Will Climate change affect outbreak patterns of planthoppers in Bangladesh? *Plos One*, 9(3): 1-10.
- Anand, K. A., Rama, R. CV., Mallikharjuna, R. N., Krishnam, R. S. and Nafeez, U. SK. (2019). Influence of certain insecticides on the resurgence of rice brown planthopper, *Nilaparvata lugens* (Stal). *Journal of Entomology and Zoology Studies*, 7(3): 874-878.
- Bao, H., Liu, S., Gu, J., Wang, X., Liang, X. and Liu. Z. (2009). Sublethal effects of four insecticides on the reproduction and wing formation of brown planthopper, *Nilaparvata lugens*. *Pest Management Science* 65(2):170-174.
- Bommarco, R., Miranda, F., Bylund, H. and Rkman, C.B. (2011). Insecticides suppress natural enemies and increase pest damage in cabbage. *Journal of Economic Entomology*, 104(3): 782-791.

- Cabauatan, P.Q, Cabunagan, R.C. and Choi, I. (2009). Rice viruses transmitted by the brown planthopper *Nilaparvata lugens* Stål. In Heong, K.L. and Hardy, B. (Eds.). *Planthoppers: New threats to the sustainability of intensive rice production systems in Asia*. International Rice Research Institute, Pp.357-368.
- Chelliah, S. (1979). Insecticide application and brown planthopper, *Nilaparvata lugens* (Stål) resurgence in rice. A report of research conducted from July 8, 1977 to July 7, 1979. Department of Entomology, International Rice Research Institute, Los Baños, Philippines. Pp. 69.
- Chelliah, S. and Uthamasamy, S. (1986). Insecticide induced resurgence of insect pests of rice. *Oryza*. 23: 71-82.
- FAO. (2006). International code of conduct on the distribution and use of pesticides – Guidelines on efficacy evaluation for the registration of plant protection products. Rome. Pp. 61.
- Gao, C.X., Gu, X.H. and Bei, Y.W. (1988). A study on the cause of resurgence of brown planthopper. *Acta Ecologica Sinica*, 8: 155-163.
- Ghosal, A. and Chatterjee, M. (2018). Insecticide induced resurgence study of whitefly in cotton and tomato. *University of Sindh Journal of Animal Sciences*, 2(2): 1-6.
- Heinrichs, E.A. and Medrano, F.G. (1984). *Leersia hexadra*, a weed host of the brown planthopper, *Nilaparvata lugens* (Stal). *Crop Protection*, 3:77-85.
- Heinrichs, E.A., Chelliah, S., Valencia, S.L., Arceo, M.B., Fabellar, L.T. and Aquino, G.B. (1981). Manual for testing insecticides on rice. International Rice Research Institute, Los Baños, Laguna, Philippines. Pp. 134.
- Heinrichs, E.A., Reissig, W.H., Valencia, S.L. and Chelliah, S. (1982). Rates and effect of resurgence-induced insecticides on populations of *Nilaparvata lugens* (Stål) (Homoptera: Delphacidae) and its predators. *Environmental Entomology*, 11: 1267-1273.
- Heong, K.L. and Hardy, B. (2009). *Planthoppers: new threats to the sustainability of intensive rice production systems in Asia*. International Rice Research Institute, Los Baños, Philippines. Pp. 157-1783.
- Islam, Z. and Catling, D. (2012). *Rice pest of Bangladesh: Their ecology and management*. The University Press Limited, Dhaka, Bangladesh. Pp. 320.
- Islam, Z., Rahman, M.A., Barrion, A.T., Polaszek, A., Chancellor, T., Heong, K.L., Ahmed, N., Haq, M. and Kamal, N.Q. (2003). Diversity of Arthropods in irrigated rice in Bangladesh. *Bangladesh Journal of Entomology*, 13(2): 1-25.
- Karns, D.L. and Stewart, S.D. (2003). Sub-lethal effects of insecticides on the intrinsic rate of increase of cotton aphid. *Entomologia Experimentalis et Applicata*, 94(1): 41-49.
- Krishnaiah, N.V., Lakshmi, V.J., Pasalu, I.C., Katti, G.R. and Padmavathi, C. (2008). Insecticides in rice IPM—Past, present and future. Hyderabad (India): Directorate of Rice Research, ICAR. Pp. 148.
- Liu, X. and Sun, Q. (2016). Early assessment of the yield loss in rice due to the brown planthopper using a hyperspectral remote sensing method. *International Journal of Pest Management*, 62 (3): 205-213.

- Muthayya, S., Sugimoto, J.D., Montgomery, S. and Maberly, G. (2014). An overview of global rice production, supply, trade, and consumption. *Annals of the New York Academy of Sciences*, 1324: 7-14.
- Sivasubramaniam, N. and Imthiyas, M.S.M. (2018). Evaluation of the major factors associated in controlling of brown planthopper (*Nilaparvata lugens*) in paddy cultivation during its outbreak in selected divisional secretariats of the Ampara District of Sri Lanka. *Scholars Journal of Agriculture and Veterinary Sciences*, 5(11): 595-600.
- Volodymyr, I.L., Tetiana, M.M., Viktor, V.H., Janet, M.S. and Kenneth. B.S. (2018). Pesticide toxicity: a mechanistic approach. *EXCLI Journal*, 17: 1101-1136.
- Wang, A.H., Wu, J.C., Yu, Y.S., Liu, J.L., Yue, J.F. and Wang, M.Y. (2005). Selective Insecticide-Induced Stimulation on Fecundity and Biochemical Changes in *Tryporyza incertulas* (Lepidoptera: Pyralidae). *Journal of Economic Entomology*, 98(4):1144-9.
- Wojciechowska, M., Stepnowski, P. and Gołębiowski, M. (2016). The use of insecticides to control insect pests. *Information Systems Journal*, 13: 210-220.
- Wu, S., Zeng, B., Zheng, C., Mu, X., Zhang, Y., Hu, J., Zhang, S., Gao, C. and Shen, J. (2018). The evolution of insecticide resistance in the brown planthopper (*Nilaparvata lugens* Stål) of China in the period 2012–2016. *Scientific Report*, 8(1): 1-11.