# GENETIC DIVERSITY AND POPULATION STRUCTURE OF SIMILAR NAMED AROMATIC RICE (Oryza sativa L.) LANDRACES OF BANGLADESH

M.Z. Islam\*, M. Khalequzzaman, T. Chakrabarty, N. Akter, M.F.R. Khan A. Bhuiya, M.A. Siddique

Genetic Resources and Seed Division Bangladesh Rice Research Institute, Gazipur, Bangladesh

#### **ABSTRACT**

Assessment of thirty-six similar named aromatic rice landraces of Bangladesh was analyzed using 36 microsatellite markers to characterize the landraces and also to establish the sovereignty of the Bangladeshi rice gene pool. With an average of 3.03 per locus, overall 109 alleles differed from 2 to 5 were detected at 36 microsatellite loci across the 36 aromatic rice landraces. With an average of 0.48, the diversity of genes ranged from 0.15 to 0.74. The polymorphic information content (PIC) values ranged from 0.14 (RM500, RM554) to 0.69 (RM496), with an average of 0.41, revealed many variations among the studied landraces. The recurrence of the most prevalent allele at each locus ranged from 31.00% (RM496) to 96.00% (RM500 and RM554). At any given locus, on average 64.33% landraces out of 36 contributed a familiar major allele. For identification and diversity estimation of aromatic rice landraces, RM496 was the finest marker as affirmed by PIC values. Two clusters were revealed with a similarity coefficient of 0.45 by a UPGMA dendrogram in SSR. All the landraces were also divided into two groups (A and B) through the model-based clustering method, confirmed by UPGMA cluster analysis. Some of the SSR markers (RM1, RM489, RM39, RM474, RM2, RM214, RM21, and RM206) generated unique alleles that were specific to particular landraces and were useful for varietal identification. Besides, the evaluation of genotypic data demonstrated the landraces under this study provided noticeable genetic diversity. Meanwhile, for the future breeding program, the similarly named landraces need to be safeguarded.

**Keywords:** Microsatellite markers, Cluster analysis, Polymorphic information content, Aromatic rice.

Received: 21.10.2021 Accepted: 23.12.2021

<sup>\*</sup> Corresponding Author: zahid.grs@gmail.com

#### INTRODUCTION

One of the most popular food crops in the world is rice (*Oryza sativa* L.). In the case of special occasions and export purposes, aromatic rice is preferred over non-aromatic rice for its notable features. Also, due to its superior grain qualities and pleasant aroma aromatic rice is a notable class of rice with high market value (Singh et al., 2000). It's believed that the foothills of Himalayas, covering Bihar, Uttar Pradesh in India, and the Tarai region of Nepal are the center of diversity of aromatic rice landraces. After that, aromatic rice germplasm might be dispersed to the other different states within India and also neighboring countries like Bangladesh and excellently adapted to the local environments of those areas (Khush, 2000). At present, Bangladesh has more than 8000 rice germplasm of which more than 100 identified as aromatic landraces (Islam et al., 2016).

Aromatic rice landraces of Bangladesh generally have short and medium bold grain types with pleasurable aroma (Shahidullah et al., 2009). Usually, aromatic rice landraces contained tall-statured, the smaller number of panicles, high stem weight, lower yields, and are also susceptible to lodging and pests. Due to the presence of a non-functional betaine aldehyde dehydrogenase 2 (BADH<sub>2</sub>) aromatic germplasm effuses fragrance which also responsible for low grain yield (Bradbury et al., 2005; Bradbury et al., 2008). Most of the aromatic rice landraces in Bangladesh are locally adapted, photoperiod-sensitive, and grown under rainfed lowland ecosystem during Transplanted Aman season (July to December). The average yield of high yielding rainfed lowland rice in Bangladesh is 3.4 t ha<sup>-1</sup>, whereas the average yield of aromatic rice is 2.0-2.3 t ha<sup>-1</sup>.

Currently, the valuable gene pool of aromatic rice landraces of Bangladesh is being eroded day by day because of the introduction of high yielding varieties (HYV) and their poor yield performance. Exploring diversity in the landrace collection is very essential for identifying new genes and further improvement of the germplasm (Thomson et al., 2007). However, similarly named rice germplasm was cultivated all over Bangladesh was identified (Hamid et al., 1982; Ahmed et al., 2016). Besides, different genotypes maybe got the same name given by many farmers or a particular genotype acquired several slightly deviated names. Hence, it is very important to study a similar named aromatic rice germplasm to identify whether they are the same or different. Some small and medium-grained of Bangladeshi aromatic rice landraces have the excellent aroma and few other quality traits like elongation after cooking, taste, etc. For measuring genetic diversity in crop germplasm and evaluating evolutionary relationships the commencement of PCR-based molecular marker technology provides highly effective and reliable tools (Islam et al., 2018a). Simple sequence repeat (SSR) markers can serve as the marker for selection and affords several advantages over other markers across various molecular markers (Roy et al.,

2016; Islam et al., 2019). Due to high reproducibility, simplicity, easy scoring ability, multi-allelic nature, hyper-variability, co-dominant inheritance, and genome-wide coverage SSR markers are highly suitable for characterizing rice germplasm (Powell et al., 1996). For genetic diversity analysis, characterization of genotypes, cultivar identification, marker-assisted selection breeding, and population structure assessment in several rice genetic studies, recently many researchers have been used SSR markers (Choudhury et al., 2013; Islam et al., 2018b). With the above background information, the present inquiry was undertaken by using SSR markers to assess the genetic variation in 36 similar named aromatic rice landraces of Bangladesh.

### **MATERIALS AND METHODS**

#### Plant materials and molecular marker

We used 36 aromatic rice accessions representing landraces, farmer's varieties, and pure lines preserved in Bangladesh Rice Research Institute (BRRI) genebank as shown in Table1. These accessions were studied in the Molecular Laboratory of Genetic Resources and Seed Division of BRRI during 2016-17 for diversity analysis. A total of 42 pairs of primers were used from the previous studies on rice (McCouch et al., 2002; Islam et al., 2018a, 2018b); some were selected randomly. Detailed information of the primers is obtained from the websitewww.gramene.org/markers/microsat.

## Molecular analysis using SSR marker

Five grams seeds from each landrace were first germinated and then germinated seeds were sown in the earthen pots. The pots were kept in the net house for collecting 2g fresh leaf samples for DNA extraction. DNA was isolated from young leaves of rice plants using the minor modified miniscale method (Islam et al., 2018a). Polymerase chain reaction (PCR) was carried out in a volume of 10 μL. Each reaction mixture contains 3.0 μL genomic DNA, 1.0 μL of 10 X PCR buffer (MgCl<sub>2</sub> free), 1.35 μL of 25 mmol/L MgCl<sub>2</sub>, 0.2 mM of a dNTPs mix, 0.5 μL of each forward and reverse primers, 1 unit of Taq DNA polymerase and 3.43 μL sterile deionized water. PCR profile was set as follow: 1 cycle at 94°C for 5 min (initial denaturation), followed by 35 cycles of 94°C for 45 s (denaturation), annealing at 55°C for 45 s and extension at 72°C for 1.3 min. Then additional temperature (final extension) of 72 °C for 7 minutes at the end of 35 cycles. The PCR products were subjected to electrophoresis in 0.5X TBE buffer for 1.5 to 2.50 h. The gel was stained with ethidium bromide solution for 25 min. Following this, the gel was viewed under UV light using a gel documentation system (XR System, Uvitec Cambridge, France).

Table 1. Information on collection site, source and local name of the landraces

Sl. No.	Landraces	Code Name	Acc. No.	Season	District	Origin	1.7% KOH (aroma)
1	Chinigura	C1	6719	T.Aman	Gazipur	Bangladesh	Scented
2	Chinigura	C2	2412	T.Aman	Dhaka	Bangladesh	Light scented
3	Chinigura	C3	4867	T.Aman	Mymensingh	Bangladesh	Light scented
4	Chinigura	C4	7572	T.Aman	Habiganj	Bangladesh	Scented
5	Chiniguri	C5	1424	T.Aman	Dhaka	Bangladesh	Scented
6	Chiniguri	C6	1880	T.Aman	Kishoreganj	Bangladesh	Scented
7	Sakkorkhora	S1	1605	T.Aman	Patuakhali	Bangladesh	Scented
8	Sakkorkhana	S2	4761	T.Aman	Barguna	Bangladesh	Scented
9	Sakkorkhana	<b>S</b> 3	5338	T.Aman	Bagerhat	Bangladesh	Scented
10	Sakkorkhana	S4	7316	T.Aman	Jhalakati	Bangladesh	Light scented
11	Sakkorkhana	S5	7500	T.Aman	Pirojpur	Bangladesh	Light scented
12	Sakkorkhora	S6	7506	T.Aman	Pirojpur	Bangladesh	Scented
13	Kataribhog	K1	232	T.Aman	Mymensingh	Bangladesh	Light scented
14	Kataribhog	K2	1091	T.Aman	Jessore	Bangladesh	Scented
15	Kataribhog	K3	1491	T.Aman	Tangail	Bangladesh	Scented
16	Kataribhog TAPL-78	K4	2505	T.Aman	Gazipur	Bangladesh	Scented
17	Kataribhog TAPL-79	K5	2506	T.Aman	Gazipur	Bangladesh	Scented
18	Kataribhog TAPL-80	K6	2507	T.Aman	Gazipur	Bangladesh	Scented
19	Kataribhog TAPL-81	K7	2508	T.Aman	Gazipur	Bangladesh	Scented
20	Kataribhog TAPL-82	K8	2509	T.Aman	Gazipur	Bangladesh	Scented
21	Kataribhog TAPL-83	K9	2510	T.Aman	Gazipur	Bangladesh	Scented
22	Kataribhog TAPL-84	K10	2511	T.Aman	Gazipur	Bangladesh	Scented
23	Kataribhog TAPL-85	K11	2512	T.Aman	Gazipur	Bangladesh	Scented
24	Kataribhog TAPL-86	K12	2513	T.Aman	Gazipur	Bangladesh	Light scented
25	Kataribhog TAPL-87	K13	2514	T.Aman	Gazipur	Bangladesh	Light scented
26	Kataribhog TAPL-88	K14	2515	T.Aman	Gazipur	Bangladesh	Scented
27	Kataribhog	K15	4362	T.Aman	Gazipur	Bangladesh	Scented
28	Kataribhog	K16	4363	T.Aman	Gazipur	Bangladesh	Scented
29	Kataribhog	K17	4791	T.Aman	Dinajpur	Bangladesh	Light scented
30	Kataribhog	K18	7082	T.Aman	Sylhet	Bangladesh	Light scented
31	Begunbichi	B1	508	T.Aman	Rangpur	Bangladesh	Light scented
32	Begunbichi	B2	740	T.Aman	Rangamati	Bangladesh	Light scented
33	Begunbichi	В3	986	T.Aman	Khulna	Bangladesh	Scented
34	Begunbichi	B4	1465	T.Aman	Dhaka	Bangladesh	Light scented
35	Begunbichi	B5	1678	T.Aman	Faridpur	Bangladesh	Scented
36	Begunbichi	В6	4088	T.Aman	Nilphamari	Bangladesh	Non-scented

## **Data Analysis**

The molecular weight for each of the markers was measured using the AlphaEaseFC 4.0 software. The summary statistics include the number of alleles per locus, major allele frequency, and polymorphism information content (PIC) was obtained by the use of Power Marker V 3.25 (Liu and Muse, 2005). The allele frequency data from Power Marker software was calculated to export data and scored as 1 or 0 indicating the presence and absence of products of a particular size. NTSYS-pc software (Rohlf, 2002) was used for dendrogram construction.

The population structure of 36 aromatic landraces was determined using the STRUCTURE V2.3.4 software (Pritchard et al., 2000; Falush et al., 2003). The number of populations (K) investigated here and ranged (1 - 10), replication: 5, burnin period length: 5000, run-length: 50000, and also a model allowing for admixture and correlated allele frequency. The output of the analysis was harvested using the 'Structure harvester' program (<a href="http://taylor0.biology.ucla.edu">http://taylor0.biology.ucla.edu</a>) and determined the final K value (K = 2 was optimum for this analysis) based on both the LnP (D) and Evanno's  $\Delta$ K (Evanno et al., 2005). In summary, the major patterns of variation in the multilocus dataset, an analysis of molecular variance (AMOVA) was performed using GenAlEx V 6.5 (Peakall and Smouse, 2012).

#### RESULTS AND DISCUSSION

For the efficient characterization, conservation, documentation and utilization of biodiversity, evaluation of genetic disparity in germplasm collections are obligatory. Genetic diversity in crop material is used as the basis for varietal improvement. The use of germplasm can be measured from the amount of available diversity in the material. The main objective is to know the possibility of classifying individual landraces into dissimilar groups from each genetic diversity study. Landraces which are studied in this context showed remarkable variations among the landraces for distinct agro-morphological traits (data not given). Molecular characterization, on the other hand, is the alternative approach to overcome several limitations of morphological characterization, which are high experimental cost, long evaluation time, and environmental effects. In past times characterization, genetic diversity and population structure of Bangladeshi rice germplasm have been studied by using molecular markers (Ahmed et al., 2016; Siddique et al., 2017; Islam et al., 2018a).

#### Genetic diversity

All the 36 aromatic rice landraces were genotyped with 42 simple sequence repeat (SSR) markers. Six markers (RM224, RM215, RM536, RM537, RM192, RM193) were found monomorphic (data are not shown), exhibiting one allele at each locus for all the landraces among 42 SSR markers. Then again, based on polymorphism 36 SSR markers were selected to use for molecular characterization of the aromatic rice landraces.

A total of 109 alleles were identified at 36 SSR markers over 36 aromatic landraces (Table 2). RM496 (262 bp) produced the maximum amplicon size and RM413 (66 bp) was the minimum. In the case of RM489 (242-315 bp), a maximum range of band sizes was found and succeeded by RM496 (262-314 bp) and RM474 (227-292 bp), respectively. The number of alleles per locus ranged from 2 alleles (RM178, RM507, RM510, RM447, RM282, RM487, RM554, RM542, RM500, RM560, RM342, RM553 and RM20) to 5 alleles (RM474), with an average of 3.03 alleles across the 36 loci. The PIC values differed from 0.14 (RM500, RM554) to 0.69 (RM496), with the 0.41 average. SSRs which have higher PIC values have a higher number of alleles. Lower PIC value shows that the landraces under study are closely allied, whereas the higher value of PIC stipulates the higher array of materials which is the utmost need for the new variety development. RM496 primer had the highest PIC (0.69) value and the number of alleles (5) were highest. It detected the maximum level of polymorphism. Therefore, RM496 marker confirmed that it was the best marker for characterizing the studied landraces. The most common allele was ranged from 31.00% (RM496) to 96.00% (RM500, RM554) at each locus. On average, 64.33% of the 36 rice landraces shared a common major allele at any given locus. The DNA figuration of 36 aromatic rice landraces by RM447 is demonstrated(Fig. 1).

From the present study, the genetic diversity is similar to earlier report (Islam et al., 2018a); where investigators identified 3.11 alleles per locus and an average PIC value of 0.29 among 113 aromatic rice germplasm. Correspondingly, 88 Indian rice varieties were collected from different agro-climatic regions of India having 3 alleles per locus with a mean PIC value of 0.41, also reported by Yadav et al. (2013). Again, the average PIC value of 0.44 was observed by Chakhonkaen et al. (2012) among 43 Thai and 57 IRRI germplasm of rice. Further, Ahmed et al. (2016) found 350 alleles from similarly named rice germplasm using 45 SSR markers and several alleles per locus ranged from 3 to 14 with an average of 7.8 which was higher than the present study. On the other hand, a marginally lower genetic diversity was disclosed among 40 rice accessions of Pakistan with an average of 2.75 alleles per locus and an average PIC value of 0.38 (Shah et al., 2013).

Again, a lower SSR diversity was found in a study with thirty-six (36) polymorphic HvSSRs where they identified 2.22 alleles per locus and mean PIC value of 0.25 in 375 Indian rice genotypes gathered from totally different areas of India (Singh et al., 2013). In this study, the PIC values projected that RM496 might be the leading marker for diversity analysis of aromatic rice germplasm, followed by RM214, RM474 and RM567 were probably the least potent markers (>0.60). So, these microsatellite markers may be useful tools for the upcoming genetic studies of rice germplasm.

Table 2. Allele number, allele size and frequency, gene diversity and polymorphism information content (PIC) found among 36 similar named aromatic rice landraces for 36 microsatellite markers

Markers	Chromoso me No.	Position (cM)	Motif*	Allele No.	No. of Unique allele	Size range (bp)	Size (bp)	Freq (%)	Gene diversity	PIC
RM1	1	29.7	(GA)26	5	1	82-122	122	64.00	0.55	0.51
RM489	3	29.2	(ATA)8	4	2	242-315	242	78.00	0.37	0.33
RM39	5	87.7	(CT)17CCA(TC)3	3	1	102-118	118	69.00	0.44	0.36
RM178	5	118.8	(GA)5(AG)8	2	0	116-121	121	86.00	0.24	0.21
RM507	5	-	(AAGA)7	2	0	242-250	250	83.00	0.28	0.24
RM510	6	20.8	(GA)15	2	0	112-120	120	89.00	0.20	0.18
RM223	8	80.5	(CT)25	4	0	153-168	168	56.00	0.61	0.56
RM447	8	124.6	(CTT)8	2	0	105-112	112	67.00	0.44	0.35
RM105	9	32.1	(CCT)6	3	0	128-140	140	53.00	0.61	0.54
RM316	9	1.8	(GT)8- (TG)9(TTTG)4(TG)4	4	0	184-205	184	50.00	0.65	0.59
RM474	10	-	(AT)13	5	1	227-292	227	47.00	0.68	0.64
RM552	11	40.6	(TAT)13	3	0	172-224	172	53.00	0.54	0.45
RM262	2	103.3	(CT)16	3	0	149-165	149	69.00	0.47	0.43
RM16	3	131.5	(TCG)5(GA)16	3	0	186-237	186	72.00	0.43	0.37
RM282	3	100.6	(GA)15	2	0	138-145	136	75.00	0.38	0.30
RM487	3	127.9	(AC)10	2	0	175-181	175	78.00	0.35	0.29
RM554	3	100.6	(GA)14	2	0	250-257	257	92.00	0.15	0.14
RM518	4	25.5	(TC)15	3	0	154-169	169	75.00	0.40	0.35
RM567	4	153.6	(GA)21	4	0	240-266	261	36.00	0.69	0.63
RM334	5	141.8	(CTT)20	3	0	178-191	178	64.00	0.50	0.42
RM413	5	26.7	(AG)11	4	0	66-106	66	64.00	0.54	0.51
RM190	6	7.4	(CT)11	3	0	115-135	135	56.00	0.57	0.51
RM2	7	36.1	(GA)13	3	1	140-155	155	78.00	0.36	0.31
RM214	7	34.7	(CT)14	5	1	120-142	120	36.00	0.73	0.68
RM542	7	34.7	(CT)22	2	0	95-138	138	58.00	0.49	0.37
RM320	7	62.5	(AT)11GTAT(GT)13	3	0	135-170	170	67.00	0.49	0.42
RM500	7	36.1	(AAG)9	2	0	240-252	252	92.00	0.15	0.14
RM560	7	54.2	(CT)12	2	0	228-235	235	56.00	0.49	0.37
RM72	8	60.9	(TAT)5C(ATT)15	3	0	180-198	180	61.00	0.51	0.43
RM342	8	78.4	(CAT)12	2	0	134-157	141	75.00	0.38	0.30
RM553	9	76.7	(CT)10	2	0	174-184	172	61.00	0.48	0.36
RM496	10	113	(TC)14	4	0	262-314	262	31.00	0.74	0.69
RM21	11	85.7	(GA)18	4	1	135-160	160	53.00	0.62	0.55
RM206	11	102.9	(CT)21	4	1	134-172	172	61.00	0.54	0.48

Markers	Chromoso me No.	Position (cM)	Motif*	Allele No.	No. of Unique allele	Size range (bp)	Size (bp)	Freq (%)	Gene diversity	PIC
RM224	11	120.1	(AAG)8(AG)13	3	0	136-156	156	53.00	0.54	0.47
RM20	12	-	(ATT)14	2	0	220-235	235	58.00	0.49	0.37
Total				109	9		6236	2316	17.1	14.85
Min				2			66	31	0.15	0.14
Mean				3.03			173. 22	64.33	0.48	0.41
Max				5			262	92	0.74	0.69

Unique alleles are valuable as they may be effectively indicative of particular landraces and also for a breeding purpose. Additionally, eight markers magnified nine unique alleles among 36 markers. Here, the RM1 magnified the 122 bp allele which was distinct to the Begunbichi landrace (B5). Again, the RM489 intensified the particular alleles of 248 and 315 bp in the landraces namely Chiniguri (C6) and Kataribhog (K1), respectively. The prominent aromatic rice landraces "Chinigura (C2)" was exclusively identified by RM39, "Kataribhog (K3)" by RM474 and "Kataribhog (K18)" by RM2. Normally, the higher number of unique alleles in germplasm indicates as a reservoir of novel alleles. Some unique alleles for molecular characterization of crop has been reported earlier (Das et al., 2013; Islam et al., 2018a). Among Basmati and non-Basmati rice varieties, Saini et al. (2004) noticed 58 unique alleles (36.2%).

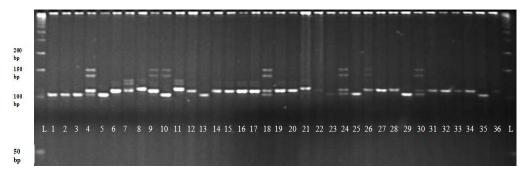


Figure 1. Gel picture of marker RM447 showing banding pattern in 36 aromatic rice landraces (L= DNA Laddaer 50 bp)

#### Genetic distance-based analysis

The genetic distance-based results in the UPGMA cluster analysis revealed two major clusters in the 36 genotypes at a coefficient of 0.45 in SSR and the similarity coefficient value ranged from 0.33 to 0.97 in SSR which is an indication of the

genetic variation among the accessions based on the SSR primers (Fig. 2). Cluster I consisted of 11 aromatic landraces (C1, C2, C3, C5, S4, B5, K1, K11, K12, K13 and K17). Again, cluster I was further divided into 2 sub-clusters (A and B). Sub-cluster A, incorporated popular landraces viz, Chinigura group (C1, C2, C3 and C5), S4 and B5. Sakkorkhana (S4) and Begunbichi (B5) are popular T. Aman landraces from Jhalakati and Faridpur districts of Bangladesh. Sub cluster-B consisted of Kataribhog group (K1, K11, K12, K13 and K17) from Mymensingh, Gazipur and Dinajpur district. Again, cluster II was further divided into 2 subclusters (A and B). Sub cluster-A, included 24 local popular aromatic landraces and sub-cluster-B consisted of one landrace namely Kataribhog (K18) from Sylhet. Kataribhog group (K4 and K5) aromatic landraces have high similarity and close placed in phylogenetic tree analyses. In previous study, Islam et al. (2018a) found three (3) major clusters and few sub-clusters through the UPGMA cluster analysis among 113 aromatic rice landraces.

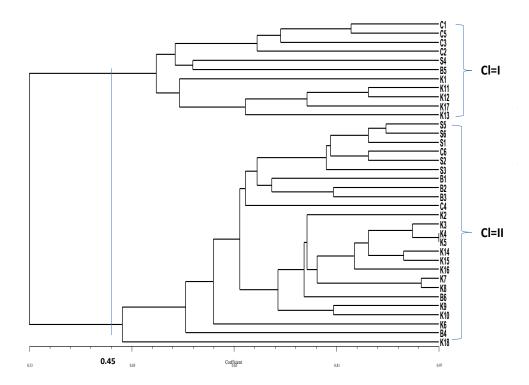


Figure 2. An UPGMA cluster dendrogram showing the genetic relationships between 36 aromatic rice landraces of Bangladesh based on the alleles detected by 36 microsatellite markers.

## Model-based population structure

The population structure analysis declared the log-likelihood value ( $\Delta K$ ) maximized to the highest value of at K=2 (Fig. 3), demonstrating a sharp peak expressing the classification of entire landraces into two specific sub-groups, here denoted as Group A and Group B, respectively.

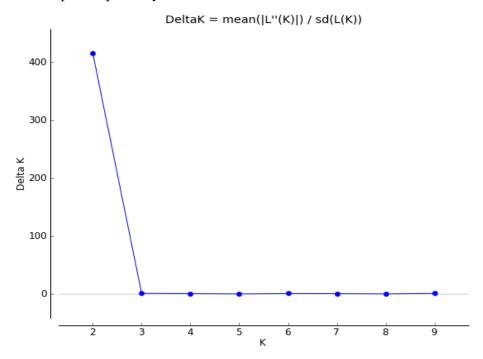


Figure 3. Representation of population structure dividing the landraces in two subgroups based on K value.

To find out the number of pure and admixed individuals, populations were studied. The population Group A (red colour, Fig. 4) and Group B (green colour, Fig. 4) representing 27.78% (10) and 72.22% (26) of aromatic landraces used in structure analysis, respectively. Overall, 05 (13.89%) admixed landraces were found at K=2. It may be noted that Group A had 12 aromatic accessions with 10 pure and 2 admixed landraces and Group B had 24 aromatic accessions with 21 pure and 3 admixed landraces. The population grouping through structural analysis and distance-based clustering demonstrated a similar result.

The Bayesian clustering approach implements to choose the number of groups with peak log-likelihood (Lu et al., 2005). From different rice diversity panels, population structure analysis has marked different numbers of sub-groups, ranged from 2 to 8 (Garris et al., 2005; Das et al., 2013; Islam et al., 2018b). With the help of structure

analysis, the 36 aromatic accessions of this study were investigated into two important groups and disclosed an adequately consistent genetic liaison with the dendrogram.

From this study, QTLs/genes mapping can be constructed by using the diverse landraces and highly polymorphic SSR markers that have been determined for different physicochemical quality traits. Finally, it can be wrapped up that similarly named aromatic rice landraces need to be preserved in genebank.

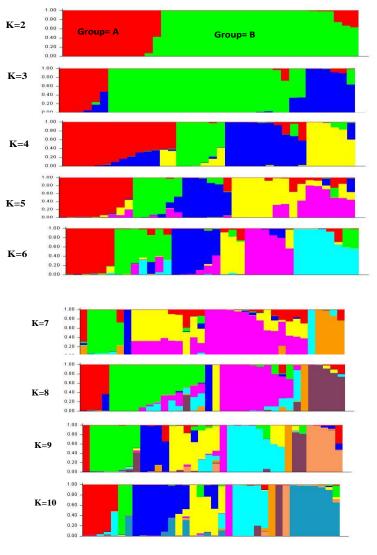


Figure 4. Population structure at K=2 to K=10 of 36 aromatic rice landraces based on genotypic data using 36 microsatellite markers.

### Analysis of molecular variance (AMOVA) from the model-based approach

From the structural analysis, two populations were consequently demonstrating to AMOVA to determine the fluctuation across and within populations. In the time, 11% variance was recorded across populations for individuals, 87% variance among, and 1% within were found (Fig. 5, Table 3). Mostly, by using the phylogenetic tree-based similarity coefficient distribution as well as the structure analysis, results from the AMOVA complied with findings achieved.

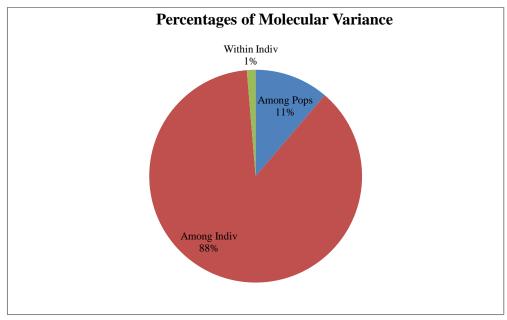


Figure 5. Analysis of molecular variance (AMOVA) of 36 aromatic rice landraces using 36 SSR markers.

Table 3. Analysis of molecular variance (AMOVA) of aromatic landraces available in Bangladesh

Source	Degree freedom	Sum Square	Mean Square	Estimated Variance	% Variance
Among Population	1	50.521	50.521	1.060	11%
Among Individual	34	564.104	16.591	8.233	87%
Within Individual	36	4.500	0.125	0.125	1%
Total	71	619.125		9.418	100%

#### **CONCLUSION**

An enormous genetic variability at molecular level revealed in this study. Analysis of population structure from the SSR markers grouped all the landraces into 2 groups according to geographical district or origin. Besides, 2 groups were also constructed using SSR markers data clustering analysis. Hence, the most divergent landraces obtained in this study can be utilized for the future aromatic rice breeding programme. Also, from this study, the diverse landraces and highly polymorphic SSR markers which have been identified can be used for QTLs/genes mapping for different physicochemical quality traits. Finally, it can be concluded that similarly named aromatic rice landraces need to be preserved in Genebank as they contain profuse genetic variation and can exploit tremendously in future breeding program.

#### ACKNOWLEDGMENT

The authors are grateful for the financial support provided by the Bangladesh Rice Research Institute (BRRI) through the Ministry of Agriculture, Bangladesh.

### CONFLICT OF INTEREST

There is no conflict of interest.

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