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Cytotoxicity and Antifungal Activities of Root Bark of Calotropis gigantea

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ABSTRACT

In this study, methanol extract from the root bark of Calotropis gigantea L. and its petroleum ether (40° C- 60° C), chloroform and ethyl acetate soluble fractions were tested for their cytotoxic activity against brine shrimp nauplii (Artemia salina, Leach) and for antifungal activity against Aspergillus flavus, Aspergillus niger, Penicillium sp and Trichoderma harzianum. Thin layer chromatography (TLC) screening showed that methanol extract and its different fractions contained different type compounds such as steroid, terpene, glycoside, heterocyclic and flavonoid. In brine shrimp lethality bioassay, it was found that chloroform fraction was highly cytotoxic (LD₅₀ 14.72 µg/ml) among the tested samples. Though methanol extract and ethyl acetate fraction have no activity against all the tested fungi but petroleum ether and chloroform fractions showed potent activity against Aspergillus niger, Penicillium sp, Trichoderma harzianum, respectively, in antifungal activity test.

Key words: Calotropis gigantea, Methanol extract, Antifungal activity, Cytotoxicty.

INTRODUCTION

Vast natural resources of medicinal plants are being used for thousands of years for the cure of many diseases in all over the world. If we could use medicinal plants properly we could get medicines at low cost and then it might be possible to fulfill the demand of our medication. This will supply low cost medicine to our poor people and we could establish a better health care system (Ghani, 1998). Calotropis gigantea L. (Family: Asclepiadaceae), a wildly growing plant, has been reported to possess number of medicinal properties and other purposes (Chitme et al., 2004). Roots of *Calotropis gigantea* have been used in leprosy, eczema, syphilis, elephantiasis, ulceration and cough in the Indian system of traditional medicine (Chitme et al., 2005). Some biologically active compounds such as calotropin, frugoside, 4/-O-beta-D-glucopyranosylfrugoside, giganticine, isorhamnetin-3-O-rutinoside, calotroposide Α and Β, isorhamnetin-3-Oglucopyranoside, taraxasteryl acetate, 19-Nor- and 18, 20-epoxy-cardenolides etc. have been isolated from different parts of Calotropis gigantea (Kiuchi et al., 1998; Kitagawa et al., 1992; Pari et al., 1998; Sen et al., 1992; Lhinhatrakool and Sutthivaiyakit, 2006). In previous study extract of Calotropis gigantea showed poor fungitoxicity on the pineapple fruit-rotting fungus Ceratocystis paradoxa (Damayanti et al., 1996). So our present attention was concentrated on the root bark of Calotropis gigantea to evaluate their bioactivity against brine shrimp nauplii and some fungi other than Ceratocystis paradoxa. This will ultimately lead to proper use of this important medicinal plant without having side effects and toxicity for various diseases.

MATERIALS AND METHODS

Plant material

The root bark of *Calotropis gigantea* (Linn) (Family: Asclepiadaceae) were selected for the biological investigations. Roots of *Calotropis gigantea* (Linn) were collected from the relevant areas (Meherchandi) of Rajshahi University campus. The plant was taxonomically identified by Professor A.T.M Naderuzzaman, Department of Botany, University of Rajshahi. Voucher specimen (No. 1A. Alam, collection date 15.08.2004) was kept in the Department of Botany, University of Rajshahi.

Extraction

The powdered plant materials (900gm) were extracted with methanol in Soxhlet extractor. The extraction process was performed repeating 8 cycles. The extract was then filtered through Whatman No.1 filter paper. The filtrate was concentrated with a rotary evaporator under reduced pressure at 60° C to afford crude methanol extract (40 gm). This crude methanol extract (30 gm) was then fractioned into petroleum ether (3 g), chloroform (10 g) and ethyl acetate soluble fraction (2 g) by solvent-solvent partitioning (Bahl and Bahl, 1992).

Thin Layer chromatography (TLC) screening

A small portion of methanol extract and its petroleum ether, chloroform and ethyl acetate fractions were dissolved in respective solvents and the solutions were spotted on TLC plates. Then the TLC plates were run by specific solvent system and viewed individually under UV light and vanillin- H_2SO_4 reagent was used as spray reagent (Bobblt, 1963). Different colored spots on TLC plate indicated the presence of different type of compound in each fractions.

Brine Shrimp Lethality Bioassay

The experiment was carried out using the method described by Mclaughlin et al. (1982). In brief, Artemia salina Leach (brine shrimp eggs) was allowed to hatch and mature as nauplii (Larvae) in seawater for 48 h at 25°C. Serially diluted test solutions (80 μ L in DMSO from a stock solution of 5 mg/mL DMSO) were added to the seawater (5 mL), containing 10 nauplii. After incubation for 24 h at 25°C, the number of survivors was counted. The LC₅₀ (50% lethal concentration, μ g/ml) was determined from triplicate experiments using probit analysis as described by Finney (1971). Ampicillin trihydrate was used as positive control.

Antifungal Activity Test

In vitro antifungal screening was performed by disc diffusion assay method (Bear et al., 1966; Rios et al., 1988). The methanol extract and its petroleum ether, chloroform and ethyl acetate fractions were tested against four fungi (*Aspergillus flavus*, *Aspergillus niger*, *Penicillium* sp and *Trichoderma harzianum*) at a concentrations of 100 μ g/disc, 200 μ g/disc and 300 μ g/disc for each and the results were compared with flugal (200 μ g/disc). The activity was determined after 72 hours of incubation at 37.5°C. The fungal strains were collected from pathology laboratory, Department of Botany, Rajshahi university, Bangladesh.

RESULTS AND DISCUSSION

On TLC screening methanol extract and its petroleum ether, chloroform and ethyl acetate soluble fractions showed mixture of coloured compounds such as steroid, triterpene, heterocyclic, glycoside and flavonoid when sprayed with vanillin-H₂SO₄ spray reagent. In brine shrimp lethality bioassay, the crude methanol extract and its petroleum ether, chloroform and ethyl acetate soluble fractions showed positive results indicating that all testing samples were biologically active. The mortality was not shown in the solvent control batch. The results obtained from this experiment are shown in Table 1. Among the tested samples, chloroform fraction showed lowest LD₅₀ value (14.72 μ g/ml), whereas positive control ampicillin trihydrate showed LD₅₀ value 12.27 μ g/ml. Rasid et al. (2004) showed that the LD₅₀ for petroleum ether, chloroform and methanol extracts of Wedelia calendulacea against the brine shrimp nauplii were found to be 4.59 µg/ml, 7.99 µg/ml and 14.88 μ g/ml respectively, whereas the positive control, vincristine sulfate showed an LD₅₀ of 0.58 μ g/ml. So cytotoxicity of chloroform fraction was remarkable in respect to ampicillin trihydrate. Chloroform fraction showed activity against Aspergillus niger (16, 24 and 28 mm) and Tricodarma harzianum (14, 16 and 18 mm) at a concentration of 100, 200 and 300 µg/disc, respectively and petroleum ether fraction showed activity against Asperaillus niger (20, 24, and 28 mm), Penicilium sp (20, 22 and 33 mm) and Tricodarma harzianum (20, 26 and 27 mm) at 100, 200 and 300µg/disc, respectively.

Methanol extract and ethyl acetate fraction showed no antifungal activity against all the tested fungi. It might be due to the presence of low concentration of active phytochemicals in methanol extract or due to absence of these phytochemicals in ethyl acetate fraction. The results of antifungal activity (zone of inhibition) of test materials against respective fungi were given in the Table 2.

Treatment	LD ₅₀ (in ppm)
Ampicillin trihydrate	12.27
Methanol extract	66.86
Petroleum ether fraction	39.34
Chloroform fraction	14.72
Ethyl acetate fraction	18.57

Table 1. Cytotoxic effect of the methanol extract and its petroleum ether, chloroform and ethyl acetate fractions.

Table 2. In-vitro antifungal activity of methanol extract and its petroleum ether, chloroform and ethyl acetate fractions.

Tested fungi -	Treatment												
	Methanol extract				Petroleum ether fraction		Chloroform fraction		Ethyl acetate fraction			Flugal	
	A	В	С	A	В	С	A	В	С	A	В	С	В
	Diameter of zone of inhibition (mm)												
Aspergillus	R	R	R	R	R	R	R	R	R	R	R	R	24
flavus	R	R	R	20	24	28	16	24	28	R	R	R	33
Aspergillus	R	R	R	20	22	23	R	R	R	R	R	R	30
niger	R	R	R	20	26	27	14	16	18	R	R	R	31
<i>Penicillium</i> sp													
Trichoderma													
harzianum													

CONCLUSION

The findings of this study indicate that some chemical principles with potent cytotoxic and antifungal activity present in chloroform and petroleum ether fractions. If we can isolate and characterize these bioactive principles from petroleum ether and chloroform fractions, we can use them in various medicinal purposes and our future research plan is going on this direction.

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REFERENCES

Bahl BS, Bahl A. (1992) A Text Book of Organic Chemistry, 13th Ed, pp. 11, Schand Company Ltd.

Beur AW, kirby WMM, Sherris JC, Turck M. (1966) Antibiotic susceptibility testing by standardized single disc method. *Am. J. Clin. Pathol.* 44: 493-496.

Bobblt JM. (1963) Thin layer chromatography, pp. 94, Chapman and Hall Ltd. London.

- Chitme HR, Chandra M, Kaushik S. (2004) Studies on anti-diarrhoeal activity of *Calotropis gigantea* R.Br. in experimental animals. *J. Pharm. Pharm. Sci.* 7(1): 70-75.
- Chitme HR, Chandra M, Kaushik S. (2005) Evaluation of antipyretic activity of *Calotropis gigantea* (Asclepiadaceae) in experimental animals. *Phytother. Res.* 19(5): 454-456.

- Damayanti M, Susheela K, Sharma GJ. (1996) Effect of plant extracts and systemic fungicide on the pineapple fruit-rotting fungus, *Ceratocystis paradoxa*. *Cytobios* 86(346): 155-165.
- Finney DJ. (1971) Probit analysis, 3rd Ed. pp. 18, 37, 77. University Press. Cambridge, UK.
- Ghani A. (2003) Medicinal plants of Bangladesh, 2nd Ed. pp.238. The Asiatic Society of Bangladesh, Dhaka.
- Kiuchi F, Kukao Y, Maruyama T, Obata T, Tanaka M, Sasaki T, Mikage M, Haque ME, Tsuda Y. (1998) Cytotoxic principles of a Bangladeshi crude drug, akond mul (roots of *Calotropis* gigantea L.). Chem Pharm Bull (Tokyo). 46(3): 528-530.
- Kitagawa I, Zhang RS, Park JD, Baek NI, Takeda Y, Yoshikawa M, Shibuya H. (1992) Indonesian medicinal plants.I. Chemical structures of Calotroposides A and B, two new oxypreganeoligoglycosides from the root of *Calotropis gigantea* (Asclepiadaceae). *Chem Pharm Bull* (Tokyo). 40(8): 2007-2013.
- Lhinhatrakool T, Sutthivaiyakit S. (2006) 19-Nor- and 18, 20-epoxy-cardenolides from the leaves of *Calotropis gigantea. J Nat Prod.* 69(8): 1249-1251.
- Malaughlin JL, Chang CJ, Smith DL. (1991) Bench top bioassay for the discovery of bioactive natural products. *Natural Products Chemistry*. **9**: 388-409.
- Pari K, Rao PJ, Devakumar C, Rastogi JN. (1998) A novel insect antifeedant nonprotein amino acid from *Calotropis gigantea*. J Nat Prod. 61(1): 102-104.
- Rashid AM, Rahman KM, Haider MS, Chowdhury R, Mottakin AKM. (2004) Cytotoxicity and antibacterial activity of extractives from *Wedelia calendulacea*. *Fitoterapia* 75(3-4): 355-59.
- Rios JJ, Reico MC, Villar A. (1988) Antimicrobial screening of natural products. J Enthopharmacol. 23: 127-149.
- Sen S, Sahu NP, Mahato SB. (1992) Flavonol glycosides from *Calotropis gigantea*. *Phytochemistry* 31(8): 2919-2921.